

Chapter 6. Ocean Systems

Coordinating Lead Authors

Hans-O. Pörtner (Germany), David Karl (USA)

Lead Authors

Philip W. Boyd (New Zealand / Australia), William Cheung (Canada), Salvador E. Lluch-Cota (Mexico), Yukihiro Nojiri (Japan), Daniela Schmidt (UK), Peter Zavialov (Russian Federation)

Contributing Authors

Jürgen Alheit (Germany), Javier Aristegui (Spain), Claire Armstrong (Norway), Gregory Beaugrand (France), Vsevolod Belkovich (Russian Federation), Chris Bowler (France), Peter Brewer (USA), Matthew Church (USA), Sarah Cooley (USA), Pablo Del-Monte (Mexico), Martin Edwards (UK), Michael Flint (Russian Federation), Mick Follows (USA), Thomas Frölicher (USA), Beth Fulton (Australia), Jean-Pierre Gattuso (France), Ove Hoegh-Guldberg (Australia), Eileen Hofmann (USA), Andrew Knoll (USA), Lisa Levin (USA), Lena Menzel (Germany), Coleen Moloney (South Africa), Ian Perry (Canada), Elvira Poloczanska (Australia), J. Murray Roberts (UK), Björn Rost (Germany), Jorge Sarmiento (USA), Jan Sedláček (Switzerland), Daniela Storch (Germany), Christian Wiencke (Germany), Astrid Wittmann (Germany)

Review Editors

Ken Drinkwater (Norway), Alexander Polonsky (Ukraine)

Volunteer Chapter Scientists

Lena Menzel (Germany), Astrid Wittmann (Germany)

Contents

Executive Summary

- 6.1. Introduction: Point of Departure, Observations, and Projections
 - 6.1.1. Changes in Physical and Chemical Variables
 - 6.1.1.1. Temperature and Salinity
 - 6.1.1.2. CO₂-Induced Acidification
 - 6.1.1.3. Hypoxia
 - 6.1.1.4. Light and Nutrients
 - 6.1.2. Historical and Paleo-Records
 - 6.1.2.1. Historical Observations
 - 6.1.2.2. Paleontological Records
- 6.2. Diversity of Ocean Ecosystems and their Sensitivities to Climate Change
 - 6.2.1. Pelagic Biomes and Ecosystems
 - 6.2.2. Benthic Habitats and Ecosystems
- 6.3. Climate Change Impacts from Organism to Ecosystem
 - 6.3.1. Temperature Effects
 - 6.3.1.1. Principles
 - 6.3.1.2. Microbes
 - 6.3.1.3. Macroalgae and Seagrasses
 - 6.3.1.4. Animals
 - 6.3.1.5. Ecosystems
 - 6.3.2. CO₂ Effects
 - 6.3.2.1. Principles
 - 6.3.2.2. Microbes

- 6.3.2.3. Macroalgae and Seagrasses
- 6.3.2.4. Animals
- 6.3.2.5. Ecosystems
- 6.3.3. Life in Hypoxia and Anoxia
 - 6.3.3.1. Principles
 - 6.3.3.2. Microbes
 - 6.3.3.3. Animals and Plants
 - 6.3.3.4. Ecosystems
- 6.3.4. Mixed Layer Depth and Light Shaping Net Primary Production
- 6.3.5. Concurrent Responses to Multiple Drivers
 - 6.3.5.1. Principles
 - 6.3.5.2. Microbes
 - 6.3.5.3. Animals and Plants
 - 6.3.5.4. Ecosystems
- 6.3.6. Food Web Consequences
- 6.3.7. Marine Reptiles, Mammals, and Birds
 - 6.3.7.1. Principles
 - 6.3.7.2. Field Observations
- 6.3.8. Summary and Conclusions

- 6.4. Human Activities in Marine Ecosystems: Adaptation Benefits and Threats
 - 6.4.1. Ecosystem Services
 - 6.4.1.1. Food from the Sea
 - 6.4.1.2. Other Provisioning Services
 - 6.4.1.3. Climate Regulation and Extreme Events
 - 6.4.1.4. Cultural Services
 - 6.4.1.5. Supporting Services
 - 6.4.2. Management-Related Adaptations and Risks
 - 6.4.2.1. Ecosystem Management
 - 6.4.2.2. Geoengineering Approaches
 - 6.4.2.3. Health Issues
 - 6.4.3. Conclusions

- 6.5. Projections of Future Climate Change Impacts through Modeling Approaches
 - 6.5.1. Oceanic Primary Production
 - 6.5.2. Higher Trophic Levels
 - 6.5.3. Ecosystems and Fisheries
 - 6.5.4. Conclusions

- 6.6. Chapter Conclusions and Key Uncertainties
 - 6.6.1. Key Risks Related to Climate Change: Constraints on Ecosystem Services
 - 6.6.2. Key Uncertainties

References

Chapter Box

- 6-1. An Atlantic Ocean Example: Long-Term Responses of Pelagic Organisms and Communities to Temperature

Frequently Asked Questions

- 6.1: Why are climate impacts on oceans and their ecosystems so important?
- 6.2: What is different about the effects of climate change on the oceans compared to the land, and can we predict the consequences?
- 6.3: Why are some marine organisms affected by ocean acidification?

6.4: What changes in marine ecosystems are likely because of climate change?

Executive Summary

Ocean ecosystems have responded and will continue to respond to climate changes of different rates, magnitudes, and durations (*virtually certain*). Human societies depend on marine ecosystem services, which are sensitive to climate change (*high confidence*), in particular the provisioning of food (fisheries and aquaculture) and other natural resources, nutrient recycling, regulation of global climate (including production of oxygen and removal of atmospheric CO₂), protection from extreme weather and climate events, aesthetic, cultural, and supporting services. [6.3, 6.4, 6.5]

Climate change alters physical, chemical, and biological properties of the ocean (*very high confidence*). Oceanic drivers include salinity, circulation, temperature, carbon dioxide (CO₂), oxygen (O₂), nutrients, and light. These drivers shape the physiological performance of individual cells and organisms and ultimately determine ecosystem composition, spatial structure and functioning. [6.1.1, 6.3]

The fossil record and present field and laboratory observations confirm links between key environmental drivers and responses of ocean ecosystems to climate change (*high confidence*). For millions of years in Earth history, natural climate change at rates slower than today's anthropogenic change has led to significant ecosystem shifts (*high confidence*), including species emergences and extinctions (*high confidence*). Contemporary multidecadal natural climate variations associated with regional transient warming periods by 1°C have led to fundamental restructuring of ecosystems and large socioeconomic implications (*high confidence*). [6.1.2, 6.3.1, 6.4]

Vulnerability of most organisms to warming is set by their physiology which defines their limited temperature ranges and hence their thermal sensitivity (*high confidence*). Temperature defines the geographic distribution of many species and their responses to climate change. Shifting temperature means and extremes alter habitat (e.g., sea ice and coastal), and cause changes in abundance through local extinctions and latitudinal expansions or shifts (*very high confidence*). Vulnerability is greatest in polar animals due to their narrow temperature ranges (*medium confidence*) and in tropical species living close to upper thermal limits (*medium confidence*). Although genetic adaptation occurs (*medium confidence*), the capacity of present-day fauna and flora to compensate for or keep up with the rate of ongoing thermal change is limited (*low confidence*). [6.3.1, 6.3.5, 6.5.2]

The warming-induced shifts in the abundance, geographic distribution, migration patterns, and timing of seasonal activities of species (*very high confidence*) have been and will be paralleled by a reduction in their maximum body size (*medium confidence*). This has resulted and will further result in changing interactions between species, including competition and predator-prey dynamics (*high confidence*). Numerous observations over the last decades in all ocean basins show global-scale changes including large-scale distribution shifts of species (*very high confidence*) and altered ecosystem composition (*high confidence*) on multidecadal time scales, tracking climate trends. The distribution and abundance of many fishes and invertebrates have shifted poleward and/or to deeper, cooler waters (*high confidence*). Poleward displacements of phyto- and zooplankton have occurred by hundreds of km's per decade (*high confidence*). Some warm-water corals and their reefs have responded with species replacement, bleaching and a decreased coral cover causing habitat loss (*high confidence*). While marine reptiles such as turtles encounter direct effects of warming, impacts to seabirds and marine mammals are mostly indirect through effects of warming on their prey (*high confidence*) [6.3.1, 6.3.7, 6.5, CC-CR, CC-MB].

In response to further warming by 1°C or more by the mid 21st century and beyond, ocean-wide changes in ecosystem properties are projected to continue (*high confidence*). Large irreversible shifts in the spatial distribution of species and seasonal timing of their activities (feeding, growth, development, behaviors, and productivity) will have implications for species composition, and ecosystem goods and services. [6.3.1, 6.4, 6.5, 6.6]

By the mid 21st century, the spatial shifts of marine species will cause species richness to increase at mid and high latitudes (*high confidence*) and to decrease at tropical latitudes (*medium confidence*), resulting in global redistribution of catch potential for fishes and invertebrates, with implications for food security (*medium*

confidence). Animal displacements are projected to lead to high-latitude invasions and high local-extinction rates in the tropics and semi-enclosed seas. This will cause a 30–70% increase in the fisheries yield of some high-latitude regions by 2055 (relative to 2005), a redistribution at mid latitudes, but a drop of 40%- 60% in the tropics and the Antarctic, based on 2°C warming above pre-industrial values (*medium confidence* in the direction of trends in fisheries yields, *low confidence* in the magnitude of change). If a decrease in global net primary production (NPP) or a shift towards smaller primary producers occurs, the overall fisheries catch potential may also decrease. [6.3.1-4, 6.4.1, 6.5.1-4]

Open ocean NPP is projected to fall globally depending on RCP scenario (*medium confidence*). The estimated decrease will occur by up to 9% by 2100 under the RCP 8.5 business-as-usual climate scenario (relative to 1990, *low confidence*). The oceans currently provide about half of global net primary production (NPP). Environmental controls on NPP include temperature, CO₂, nutrient supply and light (through cloud cover, mixed layer depth), all of which will be altered (WGI, Ch. 6.3). Present observations indicate increasing NPP at high (Arctic) latitudes (*medium confidence*), projected to continue beyond 2100 (*medium confidence*). This increase is offset by a decrease at temperate and tropical latitudes (*medium confidence*). Poor representation of shelf and coastal regions hamper projections in global NPP models for near-shore waters, reducing confidence in global projections. [6.3.4, 6.5.1, CC-PP]

Large-scale processes and climatic feedbacks sustained by microbes (bacteria, archaea, unicellular algae and protozoans) play key roles in marine ecosystems (e.g., carbon and nitrogen fixation, nutrient recycling) and will be altered by climate change (*medium confidence*). Identifying which microbial species, groups and processes are being affected and how these will be altered is difficult as these organisms and their responses to environmental change are extremely diverse and often modulated by biological interactions or changes in circulation and nutrient supply (*limited evidence* and *low agreement*). Warming will cause species-specific responses, such as enhancing metabolic rates and exceeding thermal tolerances, which will affect abundance, distribution, and community structure. Warmer, CO₂- and nutrient-enriched coastal oceans may stimulate harmful algal blooms (*medium confidence*), and the redistribution of certain microbes causing diseases like cholera (*medium confidence*). [6.3, 6.4.2]

Rising atmospheric CO₂ over the last century and into the future not only causes ocean warming but also changes carbonate chemistry in a process termed ocean acidification (WGI, Chs. 3.8.2, 6.4.4). Impacts of ocean acidification range from changes in organismal physiology and behavior to population dynamics (*medium to high confidence*) and will affect marine ecosystems for centuries if emissions continue (*high confidence*). Laboratory and field experiments as well as field observations show a wide range of sensitivities and responses within and across organism phyla (*high confidence*). Most plants and microalgae respond positively to elevated CO₂ levels by increasing photosynthesis and growth (*high confidence*). Within other organism groups, vulnerability decreases with increasing capacity to compensate for elevated internal CO₂ concentration and falling pH (*low to medium confidence*). Among vulnerable groups sustaining fisheries, highly calcified corals, mollusks and echinoderms, are more sensitive than crustaceans (*high confidence*) and fishes (*low confidence*). Trans-generational or evolutionary adaptation has been shown in some species, reducing impacts of projected scenarios (*low to medium confidence*). Limits to adaptive capacity exist but remain largely unexplored. [6.3.2, CC-OA]

Few field observations conducted in the last decade demonstrate biotic responses attributable to anthropogenic ocean acidification, as in many places these responses are not yet outside their natural variability and may be influenced by confounding local or regional factors. Shell thinning in planktonic foraminifera and in Southern Ocean pteropoda has been attributed fully or in part to acidification trends (*medium to high confidence*). Coastward shifts in upwelling CO₂-rich waters of the Northeast-Pacific cause larval oyster fatalities in aquacultures (*high confidence*) or shifts from mussels to fleshy algae and barnacles (*medium confidence*), providing an early perspective on future effects of ocean acidification. This supports insight from volcanic CO₂ seeps as natural analogues that macrophytes (seaweeds and seagrasses) will outcompete calcifying organisms. During the next decades ecosystems, including cold- and warm-water coral communities, are at increasing risk of being negatively affected by ocean acidification (OA), especially as OA will be combined with rising temperature extremes (*medium to high confidence*, respectively). [6.1.2, 6.3.2, 6.3.5]

The expansion of hypoxic regions termed Oxygen Minimum Zones (OMZs) and anoxic “dead” zones, observed over the last 50 years and projected into the future under climate change, especially if combined with nutrient enrichment (eutrophication), will constrain the habitat of O₂-dependent organisms and benefit anaerobic microbes (*medium confidence*). Hypoxia tolerance varies among species and is influenced by temperature, elevated CO₂, food consumption, and O₂-demand (*high confidence*). Warming-induced stratification limits the exchange of gases between water layers. Enhanced oxygen consumption by heterotrophic organisms depletes the oxygen further, causing a community shift toward lower species richness and hypoxia-tolerant specialists. Under extreme hypoxia ecosystems are dominated by microbes. These OMZs are also characterized by microbial removal of fixed nitrogen (denitrification), which can significantly reduce the low latitude nutrient inventories with implications for regional productivity. [6.3.3, 6.3.5]

The climate-change-induced intensification of ocean upwelling in some eastern boundary systems, as observed in the last decades, may lead to regional cooling rather than warming of surface waters and cause enhanced productivity (*medium confidence*), but also enhanced hypoxia, acidification, and associated biomass reduction in fish and invertebrate stocks. Due to contradictory observations there is currently uncertainty about the future trends of major upwelling systems and how their drivers (enhanced productivity, acidification, and hypoxia) will shape ecosystem characteristics (*low confidence*). [6.1.1, 6.3.2, 6.3.3, 6.3.5-6, CC-UP]

Environmental drivers acting simultaneously on ocean biota* often lead to interactive effects and complex responses (*high confidence*). Interactions of temperature, ocean acidification, and hypoxia narrow thermal ranges and enhance sensitivity to temperature extremes in organisms like corals, coralline algae, mollusks, crustaceans, and fishes (*high confidence*). In primary producers light and individual nutrients can also interact with temperature and acidification. Combined warming and ocean acidification reduce calcification in warm-water corals (*high confidence*). Ocean acidification will alter availability of trace metals (*low confidence*). (*The term biota encompasses the organisms of a region, habitat, or geological period.) [6.3.2.2, 6.3.5, 6.5.2]

The combination and often amplification of global and regional climate change and local anthropogenic drivers result in enhanced vulnerability of natural and human systems (*high confidence*). Major regional and local drivers include fishing, pollution, and eutrophication. [6.3.5, 6.4, 6.5]

The progressive redistribution of species and the reduction in marine biodiversity in sensitive regions and habitats puts the sustained provision of fisheries productivity and other ecosystem services at risk, which will increase due to warming by 1°C or more by 2100 compared to the present (*high confidence*). Human societies respond with limited adaptive capacity. Socio-economic vulnerability is highest in developing tropical countries involving a risk of reduced supplies, income, and employment from marine fisheries (*high confidence*). This emphasizes disparities in food security between developed and underdeveloped nations. [6.4.1, 6.5]

With continuing climate change, local adaptation measures (such as conservation) or a reduction in human activities (such as fishing), may not sufficiently offset global-scale effects on marine ecosystems (*high confidence*). Effects of climate change will thus complicate management regimes such as of marine protected areas once species undergo distributional shifts. This increases the vulnerabilities of marine ecosystems and fisheries. [6.4.2.1]

Geoengineering approaches involving manipulation of the ocean to ameliorate climate change (such as nutrient fertilization, binding of CO₂ by enhanced alkalinity, or direct CO₂ injection into the deep ocean) have very large environmental and associated socioeconomic consequences (*high confidence*). Some actually require purposeful alteration of ocean ecosystems for implementation. Alternative methods focusing on solar radiation management (SRM) leave ocean acidification largely unabated as they cannot mitigate CO₂ emissions. [6.4.2]

6.1. Introduction: Point of Departure, Observations, and Projections

The oceans cover about 71% of Earth's surface to an average depth of 3,700 m. Their importance for life on Earth, including humans, is vast (FAQ 6.1). Marine habitats display natural variability on various spatial and temporal scales but a dearth of long-term observational data from the vast open oceans limits our understanding of the causes and ecological consequences of this variability. The available information indicates that climate controls ocean temperatures, chemistry, circulation, upper ocean stratification, nutrient supply, and sunlight exposure. These drivers affect marine ecosystems through direct effects on organisms, amplified by their changing interactions with other species. Food webs are modified by changes in phytoplankton growth and the availability of live organisms or their decomposing bodies, i.e. debris or dissolved organic matter, as food to (chemo-) heterotrophs (organisms gaining energy by feeding on organic matter). Organismal responses lead to changes in biogeochemical processes, such as the carbon cycle, and in biological diversity and the services the oceans provide.

Some impacts of climate change on marine ecosystems and their services were addressed in the 4th IPCC assessment report (AR4), WGII, chapters 4-6 (ecosystems, food, coastal areas), and regional chapters, e.g., 15 (polar regions) and 16 (small islands). The ecosystem assessment in chapter 4 focused on terrestrial, coastal, and marine systems, their properties, goods, and services. It emphasized the difficulty in assessing future ecosystem responses as a result of ecosystem complexity, different vulnerabilities of species, and ecosystem-specific, critical thresholds associated with non-linear responses to environmental change. Focusing on terrestrial ecosystems, Ch. 4 concluded that more than 2-3°C warming above pre-industrial levels causes high extinction risks to 20 to 30% of present-day species (*medium confidence*), paralleled by substantial changes in ecosystem structure and functioning (*high confidence*). Authors projected that a wide range of planktonic and benthic calcifiers will be impacted by ocean warming (*very high confidence*) and acidification (*medium confidence*), particularly in the Southern Ocean. They characterized sea ice and coral reef biomes as highly vulnerable. Key uncertainties identified in AR4 were the incomplete knowledge of ocean acidification (addressed in present chapter 6.3.2), synergistic effects and their mechanisms (6.3.5), biotic feedbacks to the climate system (6.4), and the impacts of interactions between climate change, human uses, and ecosystem management (6.4.2).

Much more than in previous IPCC reports (Figure 1-2), impacts on the oceans are a focus in AR5. This allows for a more comprehensive discussion of phenomena and impacts, as well as the associated uncertainties and the levels of confidence in observed and projected changes. The present chapter focuses on the general principles and processes characterizing climate change impacts on ocean systems and on the uses of these systems by human societies. For projections of responses to climate change, the chapter also assesses our understanding of underlying functional mechanisms causing change across all levels of biological organization, from molecules to organisms to ecosystems. As the ocean is a heterogeneous environment, the comparison of major ocean regions is required to understand variability and differences in key processes and carbon inventories (Box CC-PP, Figure 1). We discuss the changes and variability in the ocean's principal physical and chemical properties and assess knowledge drawn from paleo- and historical to present observations. We develop a conceptual framework for analyzing effects on organisms and ecosystems and assess present knowledge derived from experiments, field studies, and numerical model projections mostly using Representative Concentration Pathways (RCP) of climate change scenarios to provide trajectories of climate change drivers (Moss *et al.*, 2010). Finally, we assess the implications of such changes for ecosystem services, and identify plausible socioeconomic consequences.

Assessing climate change impacts on coastal systems is the topic of chapter 5. An integrative treatment of regional climate changes and impacts in seven key ocean regions is the focus of regional chapter 30. Marine issues are also included in regional chapters 22 to 29, with a focus on polar oceans (Ch. 28) and small island nations (Ch. 29). Topics important to several chapters, such as ocean acidification, upwelling systems, primary productivity, changes in biogeography, and coral reefs, are discussed in joint assessments presented in the respective cross chapter boxes.

6.1.1. Changes in Physical and Chemical Variables

Trends in ocean conditions over the last 60 years reflect significant human impacts beyond natural variability on temperature, salinity, dissolved inorganic carbon and oxygen content, pH, and other properties of the upper ocean

(e.g., Pierce *et al.*, 2012; Sen Gupta and McNeil, 2012; WGI, Ch. 3.8., WGI, Table 10.1). With climate change, marine ecosystems are and will be exposed to rising temperature, ocean acidification, expansion of hypoxic zones, and other environmental drivers changing concomitantly.

6.1.1.1. Temperature and Salinity

Over the last 39 years, oceans have warmed at average rates of >0.1 °C per decade in the upper 75 m and 0.015 °C/decade at 700 m depth (WGI, Ch. 3.2.2, Figure 3.1). Trends differ regionally, seasonally, and interannually (WGI, Ch. 2.7, for ocean regions see 30.5). Temperature changes are particularly large at ENSO (El Niño-Southern Oscillation) with high (3 to 4 year) and low (5 to 7 year) frequencies, and on multidecadal scales (>25 years, Figure 6-1). The strongest warming trends are found at high latitudes where most of the interdecadal variability occurs, while tropical oceans are dominated by interannual frequencies. Global climate models have explored changes in different frequency domains, but their spatial resolution is poor (WGI, Ch. 11.3.3, WGI, Ch. 12.4.7).

Temperature variations are often accompanied by changes in salinity. Increased salinity results from reduced precipitation relative to evaporation, e.g., above the thermoclines (layer separating the upper mixed layer from deeper water where temperature and density change rapidly with depth) of subtropical gyres at mid to low latitudes since 1950 (WGI, Ch. 3). Decreased salinity due to enhanced precipitation relative to evaporation has occurred at some tropical and higher latitudes, exacerbated by sea-ice melt (Durack *et al.*, 2012). Both warming and freshening cause enhanced density stratification, a trend projected to continue into the 21st century (WGI, Figure 12.34; Helm *et al.*, 2010; WGI, Ch. 3, Ch. 11.3.3). Mean sea surface temperature in 2090 will be by 2.7 °C warmer than in 1990 (RCP8.5, WGI, Ch.12, Bopp *et al.*, 2013).

[INSERT FIGURE 6-1 HERE

Figure 6-1: Sea surface temperature variability between 1911 and 2011. The top left map shows the sea surface temperature average for the period. The top right map illustrates the temperature range calculated as the difference between the maximum and minimum values for each grid component during the century. The spatial distribution of variability by time scales (left hand map series, based on the Extended Reynolds Sea Surface Temperature, NOAA, 2012) corresponds to the multidecadal (25 to 40 years), bidecadal (15 to 25 years), decadal (8 to 15 years), low ENSO (El Niño Southern Oscillation) frequency (5 to 8 years), high ENSO frequency (3 to 5 years), and very high frequency (2 to 3 years) scales. The summed variabilities from the same $2^{\circ}\times 2^{\circ}$ box in all six maps corresponds to 100% of the time series variability. The areas of the right hand bubbles show the spectral density of some of the most widely used climate indices, accumulated in the same frequency windows: the Global Average Temperature and SST (sea surface temperature) Anomalies (GSST), the Southern Oscillation Index (SOI), the North Atlantic Oscillation (NAO), the Multidecadal Atlantic Oscillation (AMO), and the Pacific Decadal Oscillation (PDO). The sum of bubble surface areas for each vertical column (each climate index) corresponds to 100% of the time series variability between the 2 and 40 year periods. Climate indices were obtained from the NOAA ESRL Physical Sciences Division website.]

6.1.1.2. CO₂-Induced Acidification

Rising carbon dioxide (CO₂) concentrations in air (given as partial pressures, $p\text{CO}_2$, in μatm) cause increasing upper ocean CO₂ levels (Watson *et al.*, 2009). Starting from a pre-industrial value of 280 μatm atmospheric $p\text{CO}_2$ levels will have reached around 500 μatm by 2050 under SRES scenarios (IPCC, 2000) and all RCPs (Moss *et al.*, 2010; Meinshausen *et al.*, 2011). By 2100 values are projected to reach between 420 μatm and 940 μatm depending on the RCP. The rise in $p\text{CO}_2$ causes ocean acidification (OA), measured as a decline in water pH (negative log of proton concentration), accompanied by a fall in both carbonate ion (CO₃²⁻) concentration and the saturation states (Ω) of various calcium carbonates (CaCO₃; Zeebe and Westbroek, 2003; WGI Ch. 3.8.2, Box 3.2, Ch. 6, Figure 6.29). Hence, the seawater solubilities of three forms of calcium carbonate, namely calcite, Magnesium-calcite, and aragonite, increase. These minerals are important components of shells and skeletons of many marine organisms (6.3.2).

Ocean acidification occurs on a background of natural temporal and spatial variability of pH, $p\text{CO}_2$, and Ω . In the open ocean, the mean pH (total scale, pH_T) of surface waters presently ranges between 7.8 and 8.4 (WGI Ch. 3.8.2). In stratified mid-water layers, largely isolated from gas exchange between surface waters and air, decomposition of organic material leads to lowered oxygen (O_2) and elevated CO_2 levels (Paulmier *et al.*, 2011) associated with lower pH values. The few existing field data of sufficient duration, resolution, and accuracy (WGI Figure 3.18) show that trends in anthropogenic OA clearly deviate from the envelope of natural variability (Friedrich *et al.*, 2012). OA presently ranges between -0.0013 and -0.0024 pH_T units per year (WGI, Ch. 3.8.2, Table 3.2, Box 3.2; Dore *et al.*, 2009). Average surface ocean pH has decreased by more than 0.1 units below the pre-industrial average of 8.17. By 2100 pH is expected to change by -0.13, -0.22, -0.28, -0.42 pH_T units, at CO_2 levels of 421, 538, 670, and 936 ppm under RCP 2.6, 4.5, 6.0 and 8.5 climate scenarios, respectively (WGI, Figure 6.28). The rate of acidification in surface waters varies regionally and is 50% higher in the northern North Atlantic than in the subtropical Atlantic (Olafsson, 2009). Salinity reduction caused by ice melt or excess precipitation (Jacobs and Giulivi, 2010; Vélez-Belchí *et al.*, 2010) exacerbates OA by diluting the concentrations of substances acting as buffers (Steinacher *et al.*, 2009; Denman *et al.*, 2011). At high sustained CO_2 concentrations the changes in ocean chemistry will take thousands of years to be buffered by the natural dissolution of calcium carbonate from sediments and tens to hundreds of thousands of years to be eliminated completely by the weathering of rocks on land (Archer *et al.*, 2009).

6.1.1.3. Hypoxia

The average dissolved oxygen concentration in the ocean is presently $162 \mu\text{mol kg}^{-1}$ (Sarmiento and Gruber, 2006). Concentrations range from over $500 \mu\text{moles kg}^{-1}$ in productive Antarctic waters super-saturated with oxygen (Carrillo *et al.*, 2004) to zero in coastal sediments and in permanently anoxic deep layers of isolated water bodies, such as the Black Sea and the Cariaco Basin. Hypoxia results from oxygen depletion in excess of supply as in stratified water bodies (6.1.1.2). Vast oxygen minimum zones (OMZ) exist between less than 100 and more than 900 m depths in Eastern Atlantic and Pacific tropical oceans. The ecological literature applies the term hypoxia (see 6.3.3) to O_2 concentrations below $60 \mu\text{moles kg}^{-1}$ (estimated at ~5 % of global ocean volume; Deutsch *et al.*, 2011). Pacific OMZs regularly reach oxygen levels below $20 \mu\text{moles kg}^{-1}$ (about 0.8% of global ocean volume; Paulmier and Ruiz-Pino, 2009), lower than Atlantic ones. Suboxic waters at $< 4.5 \mu\text{moles O}_2 \text{ kg}^{-1}$ occupy about 0.03 % of the ocean volume, mainly in the northeastern tropical Pacific (Karstensen *et al.*, 2008).

OMZs are naturally present in many habitats including marine sediments, but are also expanding due to anthropogenic influences. Over the past 50 years, open ocean O_2 concentrations have decreased by a mean rate of 0.1 to over $0.3 \mu\text{moles kg}^{-1} \text{ year}^{-1}$ (WGI, Ch. 3.8.3, Stramma *et al.*, 2008). In some OMZs the rate has been much higher due to warming, increased stratification, and rising biological O_2 demand (WGI, Box 6.5, Figure 1). Long-term declines in O_2 by about $7 \mu\text{moles kg}^{-1} \text{ decade}^{-1}$ have been documented at mid-water depths over much of the subarctic North Pacific (Keeling *et al.*, 2010). In coastal regions, extremely hypoxic ‘dead zones’ that exclude animal life, have increased from 42 reported in the 1960s to over 400 in 2008 and been attributed to high oxygen demand from eutrophication, the local enrichment of nutrients, resulting in organic matter loading and its decay as well as nitrous oxide formation and release (Naqvi *et al.*, 2000; Díaz and Rosenberg, 2008; Zhang *et al.*, 2010).

Future warming will *likely* accelerate the spread of hypoxic zones, especially in temperate to subpolar regions. Most models project decreasing global ocean oxygen contents by 1 to 7 % from present-day concentrations in 2100 (Keeling *et al.*, 2010; WGI Figure 6.30 under RCP 8.5), with a mean decline by 3.4 % in 2090 compared to the 1990s (Bopp *et al.*, 2013). Warming and freshening of the surface layer will increase stratification and reduce the depth of winter mixing. The evolution of low O_2 zones will be linked to changes in fluvial runoffs (e.g. Milly *et al.*, 2008; 5.3.4.3), the wind regime (e.g., Vecchi and Soden, 2007), as well as the intensity, duration and seasonal timing of upwelling events (Snyder *et al.*, 2003, 30.5.2). The potential contributions of destabilized methane hydrates and bacterial methane oxidation to exacerbate hypoxia and acidification at high latitudes remain to be explored (Westbrook *et al.*, 2009). Currently, there is no consensus on the future volumes of hypoxic and suboxic waters because of large uncertainties in potential biogeochemical effects and in the evolution of tropical ocean dynamics due to both natural and anthropogenic causes (WGI, Ch. 6). While volumes with O_2 concentrations below $80 \mu\text{moles kg}^{-1}$ are projected to increase by several %, suboxic waters $< 5 \mu\text{moles O}_2 \text{ kg}^{-1}$ may undergo a 30 % increase by 2100 compared to 2005 (*low confidence*, Bopp *et al.*, 2013).

6.1.1.4. Light and Nutrients

Most models project that the mixed layer at the ocean surface (see Figure 6-2) will become shallower in the coming decades through a strengthening of the vertical density gradient (e.g. Sarmiento *et al.*, 1998; Sallée *et al.*, 2013). Mean light levels encountered by phytoplankton are set by incoming light from solar radiation, the depth of the mixed layer, and the degree to which underwater light is attenuated by living and non-living particles (Kirk, 1994). A shallower mixed layer will *likely* result in the resident phytoplankton receiving higher mean underwater light levels if the organisms are physically mixed through this stratum (Figure 6-2).

Enhanced, seasonally prolonged stratification (Holt *et al.*, 2010), especially in the tropics, the North Atlantic, the Northeast Pacific, and the Arctic (Capotondi *et al.*, 2012), will lead to decreased vertical transport of nutrients to surface waters (Doney, 2010; Figure 6-2). River plumes (Signorini *et al.*, 1999), nutrient accumulation in the pycnocline as reported for North Pacific waters (Whitney, 2011), human-induced eutrophication, enhanced upwelling (Box CC-UP), and tidal mixing and estuarine circulation in coastal oceans could partly compensate for the projected reduction in nutrient supply in the oceans (*limited evidence, medium agreement*).

[INSERT FIGURE 6-2 HERE

Figure 6-2: Projected alteration (magnitude and frequency) of oceanic fluxes and atmospheric events due to a changing climate in the coming decades. Ocean properties will be altered from the sunlit surface layer to the mid-water stratum. In the surface ocean, the depth of the mixed layer (solid horizontal line) will shallow resulting in higher mean light levels. Increased density stratification (i.e., a strengthening sea water density gradient represented by the increasing thickness of the solid horizontal line) will reduce the vertical supply of nutrients for photosynthesizing organisms residing in the mixed layer. Anthropogenic CO₂ will acidify, i.e., lower the pH of the surface ocean (note this happens in a pH range higher than 7 such that oceans will remain alkaline but less so due to acidification). The penetration of acidified waters to depth will result in a shallower depth (dashed horizontal line) at which calcium carbonate structures, such as shells, dissolve. At depth, the location of low oxygen waters will progressively become shallower. In addition, changes in storm activity and dust deposition will influence ocean physics and chemistry, with consequent effects on ocean biota and hence ecosystems (courtesy of Reusch and Boyd, 2013).]

6.1.2. Historical and Paleo-Records

6.1.2.1. Historical Observations

Ocean ecosystems are variable in time and space, and in a non-steady-state, reflected in indices such as the North Atlantic Oscillation Index (NAO), the Atlantic Multidecadal Oscillation (AMO), the Arctic Climate Regime Index (ACRI), Pacific Decadal Oscillation (PDO), or the El Niño-Southern Oscillation (ENSO) (WGI, Box 2.5; Figure 6-1, 30.5). The combination of large, global data sets such as Reynolds, NCAR (National Center for Atmospheric Research), ICOADS (International Comprehensive Ocean-Atmosphere Data Set), with multidecadal time-series, for example near Hawaii (HOT), Bermuda (BATS), the Ligurian Sea (DYFAMED), the Canaries (ESTOC), Kerguelen Island (KERFIX), Hokkaido Island (KNOT), and Taiwan (SEATS) has provided data on the physical and biogeochemical state of the oceans (Karl *et al.*, 2003). These have been augmented by the limited-term, high resolution programs World Ocean Circulation Experiment (WOCE) and Joint Global Ocean Flux Study (JGOFS).

Historical data sets provide baseline information on ecosystem states and document the responses of biota to both, natural variability in the ocean system and surface ocean warming since the 1970s (Figure 6-3; 6.3.1). Such datasets are rare and regionally biased. Examples include changes in geographic ranges of plankton and seasonal timing (phenology) of different components of the ecosystem detected by the Continuous Plankton Recorder (CPR: e.g., Edwards *et al.*, 2001; Richardson *et al.*, 2006; Box 6-1) or multidecadal shifts in pelagic ecosystems (CalCOFI) including higher parts of the food chain such as sardines and anchovies (Brinton and Townsend, 2003; Lavaniegos

and Ohman, 2003; Chavez *et al.*, 2003, 6.3.1) and the skeletal archives of long-lived organisms such as coralline algae (Halfar *et al.*, 2011), bivalves (Schöne *et al.*, 2003), and corals (De'ath *et al.*, 2009).

Systematic, long-term interdisciplinary observations using repeated, highly calibrated measurements at a given field site are required to capture high- and low-frequency events, e.g., regime shifts (abrupt changes between contrasting, persistent states of any complex system, deYoung *et al.*, 2008). Direct observations are complemented by satellite remotely-sensed datasets. Ocean color data (e.g. Coastal Zone Color Scanner [1978-1986], SeaWiFS [1997-2010], and MODIS-AQUA [2002-present], McClain, 2009) provide estimates of chlorophyll concentrations (a proxy for phytoplankton stocks and Net Primary Production, NPP, 6.2.1, 6.3.4; Saba *et al.*, 2011). Total chlorophyll cannot be measured from space; therefore, the near-surface value (approximately one optical depth) is extrapolated to whole water-column chlorophyll based on vertical distribution using region-specific algorithms. Large uncertainties persist, as these estimates reflect both phytoplankton stocks and their physiological status (Dierssen, 2010; Behrenfeld, 2011). The ~15 year archived time series of SeaWiFS is too short to reveal trends over time and their causes. It is an example for the general issue that undersampling of ocean phenomena in time and space limits our current ability to assess present states, to distinguish effects of anthropogenic change from natural variability and to project future changes (Henson *et al.*, 2010; Beaulieu *et al.*, 2013; Box CC-PP).

6.1.2.2. Paleontological Records

Paleontological records in marine sediments provide long-term, low-resolution data on the spatial distributions of organisms and their abundances from all ages and latitudes. This information can be readily related to the concurrent shifts in multiple environmental properties that are also recorded in these sediments. The records provide insights into shifts, expansions and contractions of biogeographic ranges, species extinctions and emergences, and changes in species abundance, as well as the environmental forcings to which organisms respond. Temporal trends reveal influences of temperature, hypoxia, CO₂, and food availability on organisms and ecosystems (6.1.1, Figure 6-3).

Due to insufficient resolution the geological record often does not allow the direct attribution of a biological change to a single driver or the identification of various drivers and their relative importance. Support for projections of future changes in present-day ecosystems and their services is thus limited (6.4, 6.5, *low confidence*). Nonetheless, information gained from the geological record is invaluable as both paleo and present climatic shifts share the same combination and sign of environmental changes: increasing atmospheric CO₂ causing warming and CO₂ enrichment in the surface ocean, leading to enhanced stratification of the upper ocean and a decrease in dissolved O₂ (WGI, Chs 3, 5.3). A combination of models (WGI, Chs 3, 6, 12) and geological data can be used to forecast future impacts on ocean biota (*medium confidence*).

[INSERT FIGURE 6-3 HERE]

Figure 6-3: Environmental changes (bottom) and associated biological responses (top) for the Paleocene Eocene Thermal Maximum (PETM, left) and the industrial era (right). The PETM represents the best geological analogue for the future ocean in terms of biological responses. Episodes of largest environmental change are indicated with yellow bars. Note the different time scale between the two columns. Both time intervals are characterized by rapid warming both on land and in the ocean and increases in CO₂. Atmospheric CO₂ and temperature are described with direct measurements (black), proxy reconstructions (grey) and model results (light grey). For the recent anthropogenic record, the AMO is shown to highlight high frequency natural temperature fluctuations (Enfield *et al.*, 2001) and their influence on marine biota. Biological responses to the environmental forcing are divided into calcification, extinction and migration. Note the highly group-specific responses to the environmental change, especially with regards to calcification with decreases, increases and high variability. While there was extinction during the PETM, there is currently no evidence for climate-related extinction in the marine record. The warming led to migration of warm water species into previous cold water habitats indicating range expansion due to warming (modern: planktic foraminifera, *G. ruber* Field *et al.*, 2006; PETM: coccolithophores, *Discoaster* spp. Bralower, 2002). CO₂ data: Mauna Loa (Keeling *et al.*, 2005), ice core records (Etheridge *et al.* 1996) and model output for the PETM (Zeebe *et al.*, 2009; Ridgwell and Schmidt, 2010). Sea surface temperatures modern: Wilson *et al.*, 2006 and PETM: Kennett and Stott, 1991. Biotic responses: growth changes in coralline algae (Halfar *et al.*, 2011, bottom)

and corals (Vásquez-Bedoya *et al.*, 2012, middle, De'ath *et al.*, 2009 [corrigendum], top); calcification changes (Foster *et al.*, 2013) and extinction in benthic foraminifers (Thomas, 2003) for the PETM.]

The last glacial-interglacial transition is associated with an average increase in atmospheric CO₂ of ~1 µatm/century between 18 and 10 thousand years before present (kyr BP) (WGI Ch. 5), a significantly slower increase than the ~90 µatm in the last century (WGI Chs. 5, 6). Consequently, the average pH change of 0.002 pH units per century during the glacial-interglacial transition is small relative to the ongoing anthropogenic perturbation of >0.1 pH unit per century (WGI, Ch. 3.8.2). Overall the upper glacial ocean was more O₂ rich than today's ocean (Jaccard and Galbraith, 2012) and between 0.7 and 2.7°C colder, with strong regional differences of up to 10°C cooling in the North Atlantic and 2-6°C in the Southern Ocean (WGI Ch. 5, Table 5.2). During warming from the glacial into the interglacial marine plankton such as foraminifera, coccolithophores, diatoms, dinoflagellates, and radiolarians showed marked poleward range expansion (*high confidence*, see WGI, Ch. 5.7; CLIMAP Project Members, 1976; MARGO Project Members, 2009). Under the lower glacial CO₂ concentrations, calcification in planktonic foraminifera was higher (*limited evidence, medium agreement*).

The most prominent abrupt climate change periods in the recent geological record, developing within 10-100 years, are associated with Dansgaard-Oeschger (DO) and Heinrich events (WGI, Ch. 5.7), which occurred repetitively during the last 120 kyrs. While the atmospheric changes happened within a few decades, the sea surface temperature in the North Atlantic changed by up to 5°C within decades to centuries (WGI, Ch. 5.7). Southern Ocean temperature changes were slower (hundreds to thousands of years, Barker *et al.*, 2009). The cold phase of a DO event led to the migration of polar foraminiferal species towards the equator, in the North Atlantic as far south as the Iberian Peninsula (Martrat *et al.*, 2004). Abrupt (~100 year) abundance changes in the Southern Ocean were associated with latitudinal shifts in the Antarctic Circumpolar Current and associated species (Barker *et al.*, 2009) akin to modern changes in plankton range due to warming (Box CC-MB, Box 6-1). During the DO warm phases the Monsoon-driven Arabian Sea upwelling records show enhanced primary and export production, reduced oxygenation, and denitrification, all within ~200 years (Higginson *et al.*, 2004).

The last time the atmospheric CO₂ content approached that of today was during the Pliocene warm period (3.3 to 3.0 Ma, i.e., million years ago), with long periods of atmospheric CO₂ levels between 330-400 µatm (Pagani *et al.*, 2010; Seki *et al.*, 2010) and equilibrated temperatures ~2°C warmer than today (Haywood *et al.*, 2009, WGI Ch. 5) (*medium confidence*). The Mid-Pliocene Warm Period saw a poleward expansion of tropical planktonic foraminifera (Dowsett, 2007) (*high confidence*). Coccolithophores (Bown *et al.*, 2004), corals (Jackson and Johnson, 2000) or mollusks (Vermeij and Petuch, 1986) remained unaffected with respect to rates of species extinction or emergences compared to background rates.

Perhaps the best analogue for the future ocean is the Paleocene-Eocene Thermal Maximum (PETM, 55.3 Ma). The PETM was an event of warming (Dunkley Jones *et al.*, 2013), and ocean acidification (Zachos *et al.*, 2005) over millennia (Stassen *et al.*, 2012; Cui *et al.*, 2011) with increased runoff and nutrients into the shelf ecosystems. Model simulations for the PETM show 10 times lower rates of CO₂ input and hence ocean acidification compared to today (Ridgwell and Schmidt, 2010) (*medium confidence*). Depending on the assumed rate and magnitude of the CO₂ release, models project pH declined by 0.25 to 0.45 units in PETM surface waters and a reduction in surface ocean aragonite saturation from Ω=3 to Ω=2 or even as low as 1.5 (Ridgwell and Schmidt, 2010). Warming caused range expansions of warm-water taxa towards higher latitudes (*high confidence*). The composition of plankton assemblages changed both within and between phytoplankton groups (Gibbs *et al.*, 2006; Sluijs and Brinkhuis, 2009), possibly reflecting the warming trend and/or changes in nutrient availability (6.2.2.-3). There was no bias in extinction towards more heavily calcifying species, possibly as slow CO₂ input led to minor surface water acidification. By contrast, benthic foraminifera, the dominant deep water eukaryote, recorded up to 50% extinction (Thomas, 2007). In contrast to sediment dwellers, more mobile pelagic crustaceans (ostracods) did not show any significant change in species composition (Webb *et al.*, 2009). In shallow coastal waters, calcareous algae and corals were replaced by symbiont-bearing benthic foraminifera (Scheibner and Speijer, 2008) (*medium confidence*).

The warm climates of the Mesozoic (251 to 65 Ma) led to a number of anoxic events in the oceans (Jenkyns, 2010). In some cases, OMZs expanded vertically leading to anoxia in upper water layers (Pancost *et al.*, 2004). Some of the Cretaceous oceanic anoxic events were associated with extinctions or increased species turnover (normalized sum of

originations and extinctions) of planktonic foraminifera and radiolarians (30%). Such turnover was very small in other groups of organisms, (e.g., a maximum of 7% of coccolithophores, Leckie *et al.*, 2002). The attribution of these evolutionary changes to reduced O₂ is tenuous as warming, changes in nutrient supply and, possibly, ocean acidification occurred concomitantly (Hönisch *et al.*, 2012).

Global-scale collapse of marine ecosystems is rare, even in the geological record. Some mass extinctions, in particular the Permian Period extinction 251 Ma ago, have been associated with large-scale inputs of carbon into the atmosphere and ocean, with associated warming and deep-sea O₂ decline (Knoll *et al.*, 2007; Kiessling and Simpson, 2011). The end-Permian mass extinction preferentially affected reef organisms such as corals and sponges resulting in a 4 Myr period without reef builders (Kiessling and Simpson, 2011), and underscores that vulnerabilities differ among organisms depending on anatomy, physiology, and ecology (Knoll and Fischer, 2011). The rates of environmental change and any potential acidification have not yet been accurately constrained for these events.

Of the last 100 Myr, only the last 2 Myr had CO₂ levels of ~190 to 280 ppm, comparable to preindustrial values. Values like those predicted by the mid and end of this century can solely be found in the geological record older than 33 Ma, with large uncertainties in the absolute numbers (WGI, Ch. 5.3; Hönisch *et al.*, 2012). That marine biota thrived throughout high CO₂ times cannot imply that marine organisms will remain unaffected in a future warm, high-CO₂ world. The key environmental issue of the 21st century is one of an unprecedented rate of change, not simply magnitude, of CO₂ levels (Hönisch *et al.*, 2012). The current rate and magnitude of ocean acidification are at least 10 times faster than any event within the last 65 Ma (*high confidence*, Ridgwell and Schmidt, 2010) or even 300 Ma of Earth history (*medium confidence*, Hönisch *et al.*, 2012). The slower events in geological history provide *robust evidence (high agreement)* for environmentally-mediated changes in biogeographic ranges of fauna and flora, their compositional changes, extinctions, and, to much lesser degree, emergences (*very high confidence*). No past climate change event perfectly parallels future projections of anthropogenic climate change, which is unprecedented in evolutionary history. Existing similarities indicate, however, that future challenges (6.1.1, 6.3.1-8) may be outside the adaptive capacity of many organisms living in today's oceans (*low to medium confidence*).

6.2. Diversity of Ocean Ecosystems and their Sensitivities to Climate Change

Global-scale observation and modeling studies provide *robust evidence* of present and future climate-mediated alterations of the ocean environment (*high agreement*, 6.1.1, WGI, Chs. 3, 6; Bopp *et al.*, 2013), which in turn impact ocean ecosystems (*high confidence*; Boyd and Doney, 2002; Hoegh-Guldberg and Bruno, 2010; Drinkwater *et al.*, 2010). An assessment of present findings and projections requires knowledge of the characteristics of ocean biota and ecosystems and their climate sensitivity.

Life on Earth is diverse as a result of nearly 4 billion years of evolutionary history. Marine microorganisms are the oldest forms of life and the most functionally diverse; multicellular organisms are constrained to limited functional abilities. Knowledge of overarching similarities across the organism domains Archaea, Bacteria, and Eukarya (Woese *et al.*, 1990) or kingdoms Bacteria, Protozoa, Fungi, Plantae, Animalia, and Chromista (Cavalier-Smith, 2004) would facilitate projections of climate impacts. The phylogenetic and metabolic diversity of microbes (i.e., viruses, archaea, bacteria, protists and microalgae) sustains key ecosystem processes such as primary production, CO₂ fixation and O₂ production, the conversion of nitrogen into ammonia (N₂ fixation), and the use of nitrate, sulphate, CO₂, and metals (Fe and Mn) in metabolism instead of O₂ when it is absent. Microbes enhance the horizontal transfer of genetic information between unrelated individuals, thereby enhancing biodiversity (McDaniel *et al.*, 2010). Microbes may respond to climate change by exploiting their large diversity, undergoing species replacements (Karl *et al.*, 2001), and thereby sustain their biogeochemical roles. Species replacements also occur among plants and animals, but in most cases research has focused on their resilience, well-being, abundance, survival, and conservation under climate change (FAQ 6.2).

6.2.1. Pelagic Biomes and Ecosystems

Pelagic organisms are key to biogeochemical processes in the ocean. The base of the marine food web is the photosynthetic fixation of CO₂ by phytoplankton, a process termed (net) primary production (NPP, Box CC-PP). Photosynthesis is controlled by light, temperature, inorganic nutrients (CO₂, nitrate, phosphate, silicate, and trace elements including iron), and the density-dependent stability of the surface mixed-layer depth (MLD) (6.1.1; Figure 6-2; Sverdrup, 1953; González-Taboada and Anadón, 2012). Environmental variability and the displacement of organisms by ocean currents cause variability in phytoplankton productivity, competitiveness, and natural selection (Margalef, 1978) and result in changes in carbon sequestration (Box CC-PP, Figure 6-4). Nutrient limitation leads to a decrease in NPP or chlorophyll levels and a reduction in the amount of energy supplied to higher trophic levels, including fish and invertebrates (*high confidence*, Ware and Thomson, 2005; Brander, 2007), affecting fishery yields (Cheung *et al.*, 2008; Friedland *et al.*, 2012). The wide range of trophic structures in marine food webs and the potentially non-linear changes in energy transfer under different NPP and temperature scenarios (Stock and Dunne, 2010) hamper accurate projections of changes in higher trophic levels.

[INSERT FIGURE 6-4 HERE]

Figure 6-4: A schematic representation of the ocean's biological pump, which will be influenced by climate change and is a conduit for carbon sequestration. It is difficult to project how the pump might be altered (Table 6-1) and whether it would represent a positive or negative feedback to climate change through the cumulative effects of affected processes, surface to depth (Table 6-1): shifts in Net Primary Production, floristic and faunistic community composition in the pelagic realm, and in grazing rates; alterations to the ballasting of settling particles and the proportion of NPP released as DOM (Dissolved Organic Matter); modified bacterial enzymatic rates and particle solubilization; faunistic shifts at depth. Modified from Buesseler *et al.* (2008) by J. Cook (WHOI).]

[INSERT TABLE 6-1 HERE]

Table 6-1: To assess how a changing climate will alter the ocean's biological pump (Figure 6-4) and determine the resulting biogeochemical feedbacks on global climate, changes in a wide range of processes from cellular to ocean basin scale, and from epipelagic to mesopelagic, must be quantified. This table illustrates the complexity of the integrated knowledge platform needed to provide evidence of these biogeochemical ramifications and thus the present limits to clear conclusions about climate-induced effects on the biological pump (C, Carbon; TEP, Transparent Exopolymer Particle; DOM, Dissolved Organic Matter; POM, Particulate Organic Matter).]

6.2.2. Benthic Habitats and Ecosystems

The ocean's primary production is inextricably linked with benthic (sea floor) communities via the biological pump (Figure 6-4), the chemical exchange of nutrients and gases, and the existence of organisms with both pelagic and benthic life history stages. Even in abyssal habitats, a continuous rain of organic detritus serves as the primary source of carbon and energy. Therefore climate impacts on surface marine ecosystems will impact even the deepest benthic communities, even if direct changes to their physical habitat do not occur (Smith *et al.*, 2009).

Benthic organisms living in shallow waters or the intertidal zone (where they encounter temporary exposure to air) are exposed to widely fluctuating and progressively changing means and extremes of environmental variables, such as temperature, oxygen, CO₂, salinity, and sea level (WGI, Chs. 3, 13; 6.3.1-3, 6.3.5). Sessile or slow moving plants and animals may be unable to escape from unfavorable changes except by means of advection of fertilized eggs or planktonic larvae. If climate change harms those species engineering benthic habitats, the entire ecosystem may be impacted. This concerns those ecosystem engineers, which form habitat from the structures they produce (e.g., corals forming skeletons, 6.3.1) and those forming habitat through their behaviour (e.g., worms reworking and irrigating sediment in a process termed bioturbation). Effects on both types of ecosystem engineers (6.3.1 to 8) influence the regeneration of nutrients and affect benthic-pelagic coupling.

6.3. Climate Change Impacts from Organism to Ecosystem

Understanding climate-induced alterations in the functioning of individual organisms, species populations, communities (assemblages of various species) and ecosystems builds on studies in the laboratory, in micro- and mesocosms (closed small- to medium-sized experimental systems approximating natural conditions, holding selected biological communities), and of biota or communities in the field as well as modeling. These data inform us which taxonomic groups in what regions are more susceptible to climate change (Boyd *et al.*, 2011). Empirical studies of marine organism and ecosystem sensitivities have begun identifying the mechanisms and processes linking climate to ecosystem changes (Drinkwater *et al.*, 2010; Ottersen *et al.*, 2010). Changes in ecological community composition, species interactions, and food web dynamics often build on organismal effects elicited by climate forcing (e.g., 6.3.1.5; Boyd *et al.*, 2010; Ottersen *et al.*, 2010). The underlying mechanisms respond to climate-related factors in a hierarchy from organism (highest), tissue, cell to molecular (lowest) levels of biological organization (Pörtner, 2002a; Pörtner and Knust, 2007; Raven *et al.*, 2012). Such knowledge aids the interpretation and attribution to climate change of observed effects and is a major asset for projections of future impacts.

The genetic and physiological underpinning of climate sensitivity of organisms sets the boundaries for ecosystem response and provides crucial information on sensitivities, resilience, and the direction and scope of future change. As anthropogenic climate change accelerates, a key issue is whether and how quickly organisms can compensate for effects of individual or multiple drivers, by short-term acclimatization or long-term evolutionary adaptation across generations. Evolutionary adaptation depends on the genetic variation within a population, from which the environment selects the fittest genotypes (Rando and Verstrepen, 2007; Reusch and Wood, 2007). Genetic variation depends on mutation rates, generation time, and population size (Bowler *et al.*, 2010). However, epigenetic mechanisms, such as modifications of the genome by DNA methylation, can also influence fitness and adaptation (Richards, 2006) and can be remarkably rapid as seen in terrestrial ecosystems (Bossdorf *et al.*, 2008). In plants and animals the rate of evolutionary adaptation is constrained by long generation times, but enhanced by high phenotypic variability and high mortality rates among early life stages as a selection pool (e.g., Sunday *et al.*, 2011). The limits to acclimatization or adaptation capacity are presently unknown. However, mass extinctions occurring during much slower rates of climate change in Earth history (6.1.2) suggest that evolutionary rates in some organisms may not be fast enough to cope.

Comprehensive understanding of climate change effects on ecosystems requires addressing the effects of individual drivers across organism taxa (6.3.1 – 6.3.4), the integrated action of multiple drivers (6.3.5), the consequences for food webs (6.3.6), and the specific effects on animals breathing in air (6.3.7) and operating at the highest trophic levels.

6.3.1. Temperature Effects

The effects of temperature on ecosystems largely result from organismal responses. This requires that information on organisms' thermal sensitivities, limits, and functional properties is used to assess how temperature changes have affected and will continue to affect species distributions, abundances, diversity, trophic interactions, community assemblages, risks of species extinctions, and ecosystem functioning. Organisms also respond to temperature-driven changes in the physical environment such as stratification, reduced sea-ice cover and freshening. Ambient temperature interacts with other drivers such as ocean acidification and hypoxia (6.3.5). Ambient temperature plays a more limited role for marine mammals and seabirds (6.3.7).

6.3.1.1. Principles

All organisms including marine ones have limited temperature ranges within which they live and function. Organismal performance is related to temperature by curves called thermal reaction norms (Figure 6-5) which *likely* apply across all organisms (Chevin *et al.*, 2010), from viruses (Knies *et al.*, 2006), bacteria (Ratkowsky *et al.*, 1983), and phytoplankton (Eppley, 1972; Thomas *et al.*, 2012) to macroalgae and plants (Bolton and Lüning, 1982; Müller *et al.*, 2009; Vitasse *et al.*, 2010), and animals (Huey and Kingsolver, 1989; Angilletta, 2009). Heat tolerance

thresholds differ greatly between organisms and are hypothesized to be lowered by rising organizational complexity and body size (Pörtner, 2002a,b). Maximum heat limits of animals and plants are close to the maximum temperature found in the warmest oceans (Figure 6-6). Knowledge of reaction norms, thermal limits, and underlying mechanisms is most advanced in animals (Pörtner *et al.*, 2012; 6.3.1.4). Their role in underpinning biogeography has not been explored systematically in other organisms (e.g., Green *et al.*, 2008), reducing the confidence level in assessments of thermal impacts. In animals, changes in physiological performances influence growth, body size, behavior, immune defense, feeding, reproductive success, biogeography, phenology and therefore, ecosystem structure and functioning. Shape and width of the curves can shift through acclimatization and evolutionary adaptation (Figure 6-5A) and during life history (Figure 6-5B) with implications for the distribution boundaries of species or populations (6.3.1.5).

For any species, tracking the climate-induced displacement of tolerated ambient temperatures by undergoing shifts in biogeographical ranges to, e.g., higher latitudes during warming (6.3.1.5: Figure 6-7), can be understood as a simple mode of adaptation, implemented through dispersal (e.g., of pelagic life stages), active movements (e.g., of migrating adult fishes) or passive displacement (e.g., of early lifestages or plankton with drifting water masses). Conversely, fully completed acclimatization or evolutionary adaptation (Figure 6-5) would involve shifting thermal tolerance ranges and allow species to resist the temperature trend (e.g., warming) and to sustain fitness in their previous habitat.

[INSERT FIGURE 6-5 HERE

Figure 6-5: Thermal specialization of an organism explains the why, how, when, and where of climate sensitivity. (A) The thermal tolerance range and performance levels of an organism are described by its performance curve (exemplified for an animal). Each Performance (e.g., exercise, growth, reproduction) is maximal at its optimum temperature (T_{opt}), and becomes progressively constrained during cooling or warming. Surpassing the first low and high temperature thresholds (T_p , p, pejus: getting worse) means going into time-limited tolerance. Once further cooling or warming surpasses the next low or high thresholds (T_c , c, critical), oxygen availability becomes insufficient and an anaerobic metabolism begins. Denaturation temperatures (T_d) are even more extreme and characterized by the onset of damage to cells and proteins. Horizontal arrows indicate that T_p , T_c , and T_d -thresholds of an individual can shift, within limits, between summer and winter (seasonal acclimatization) or when the species adapts to a cooler or warmer climate over generations (evolutionary adaptation). Under elevated CO_2 levels (ocean acidification) and in hypoxic waters performance levels can decrease and thermal windows narrow (dashed grey curves). (B) The width of the thermal range (horizontal arrows) also changes over time when an individual develops from egg to larva to adult and gains weight and size. Blue to red colour gradients illustrate the range between cold to warm temperatures (after Pörtner, 2002a, 2012; Pörtner and Farrell, 2008).]

[INSERT FIGURE 6-6 HERE

Figure 6-6: Maximal values of temperature covered by various domains and groups of free-living marine organisms (bacteria to animals, domains, and groups modified after Woese *et al.*, 1990). High organizational complexity is hypothesized to be associated with decreasing tolerance to heat and to enable an increase in body size which in turn, decreases heat tolerance further (Alker *et al.*, 2001; Baumgartner *et al.*, 2002; Campbell *et al.*, 2006; Chevaldonné *et al.*, 2000; De Jonckheere *et al.*, 2009, 2011; Pörtner, 2002a,b; Sorokin and Kraus, 1962). In the domain Bacteria, the Thermotogales are less complex and most tolerant to high temperatures (Abed *et al.*, 2002; Huber *et al.*, 1986; Takai *et al.*, 1999; Tenreiro *et al.*, 1997; Ventura *et al.*, 2000). The highest temperature, at which growth can occur is 122°C for hydrothermal vent archaea, seen under elevated hydrostatic pressure in laboratory experiments (Kashefi and Lovley, 2003; Takai *et al.*, 2008.)]

6.3.1.2. *Microbes*

Temperature effects on growth, abundance, distribution, phenology, and community structure of highly diverse microbes might have large implications for ecosystem functioning (6.3, Box CC-PP). A warming ocean may initially enhance the metabolic rates of microbes (Banse, 1991) and stimulate their overall growth (Bissinger *et al.*, 2008). Data from the Continuous Plankton Recorder (6.1.2) in the Northeast Atlantic confirm that warming from 1960 to 1995 enhanced phytoplankton growth (Edwards *et al.*, 2001). Eventually, with warming, the thermal tolerance of some

groups will be challenged (Chevin *et al.*, 2010), leading to the replacement of species. This is reflected in increasing fractions of smaller phytoplankton in warmer relative to colder waters (Morán *et al.*, 2010; Flombaum *et al.*, 2013).

In response to transient warming, phytoplankton distribution in the North Atlantic shifted poleward by hundreds of km per decade since the 1950s. Phenology of plankton in the North Atlantic was also affected, with differences in sensitivity between groups (6.3.1.5; Box 6-1, *high confidence*). Coccolithophore blooms (*E. huxleyi*) in the Bering Sea were reported for the first time during the period 1997-2000, probably in response to a 4°C warming, combined with a shallower mixed layer depth, higher light levels and low zooplankton grazing (Merico *et al.*, 2004). Loss of multi-year Arctic sea ice has had a profound effect on the diversity, structure, and function of the epipelagic microbial assemblage (i.e., found in the layer into which enough light penetrates for photosynthesis) (Comeau *et al.*, 2011), and further warming is likely to have even greater impacts on the food web and on ecosystem services (*medium confidence*). Warming may also have caused the southward range extension of coccolithophores in the Southern Ocean in the 2000s (Cubillos *et al.*, 2007). However, further experimental and field observations (Giovannoni and Vergin, 2012) are required to validate model projections (Taucher and Oschlies, 2011) of differential responses to warming by different microorganisms.

6.3.1.3. Macroalgae and Seagrasses

Macrophytes in coastal waters (WGII, Ch. 5) cover 0.6 % of the world's marine areas and supply about 2-5 % of total oceanic production (Smith, 1981; Charpy-Roubaud and Sournia, 1990; Field *et al.*, 1998). They have limited temperature ranges and are sensitive to temperature extremes (*high confidence*) resulting in changes of photosynthesis, growth, reproduction and survival (following the principles of Figures 6-5, 6-6, Harley *et al.*, 2012), with consequences for their abundance, distribution, and productivity. Ice retreat in polar areas leads to an expansion of macroalgal distribution, e.g. in the Antarctic (Quartino *et al.*, 2013).

Warm- versus cold- water adapted species may have different sensitivities to warming and show a range of responses in distribution shifts (Lima *et al.*, 2007). Temperate macroalgae with wide windows of thermal tolerance acclimatize by shifting these windows following seasonal temperature changes (Kübler and Davison, 1995). Antarctic and tropical macroalgae are exposed to permanently low or high temperatures, respectively, and have consequently specialized in a limited temperature range, paralleled by a low acclimatization potential (Pakker *et al.*, 1995; Eggert *et al.*, 2006; Gómez *et al.*, 2011). Thus, Antarctic and tropical macroalgae appear to be most vulnerable to warming (*high confidence*, Short and Neckles, 1999). While observations in the tropics indicate that seagrasses tolerate higher temperatures than seaweeds (Campbell *et al.*, 2006), an increase in maximum temperature by over 1°C from 1988-1999 to 2002-2006 (30.5.3.1.5) led to increased seagrass shoot mortality in the Mediterranean Sea (Marbà and Duarte, 2010). The molecular basis of acclimatization and evolutionary adaptation, as well as their limitation in relation to the climate regime require further study in the macrophytes.

6.3.1.4. Animals

The mechanisms shaping the thermal performance curve and, thereby, an animal's thermal niche have been explained by the concept of "oxygen and capacity limited thermal tolerance" (OCLTT), applicable to marine invertebrates and fishes (Pörtner *et al.*, 2010; Figure 6-5A; FAQ 6.2). The temperature range at which animals can function best, results from optimal oxygen supply at minimal oxygen usage. At temperature extremes, oxygen supply capacity becomes constrained in relation to demand, and metabolism becomes thermally limited. Beyond upper and lower temperature thresholds (T_p , Figure 6-5A), growth, reproduction, and other key functions decrease. These thresholds change during the individual life cycle, and with body size. At large body size, limitations in oxygen supply are exacerbated and heat tolerance limits shift to lower temperatures.

Surpassing species-specific heat tolerance limits (Figure 6-5, T_p) during warming causes a reduction of abundance (Pörtner and Knust, 2007; Katsikatsou *et al.*, 2012), coral losses (Donner *et al.*, 2005), shifts in the seasonal timing of (zooplankton) biomass formation (Mackas *et al.*, 1998; Schlüter *et al.*, 2010), and changes in growth (Lloret and Rätz, 2000; Brunel and Dickey-Collas, 2010). During early life, due to incomplete development, or as adult

spawners, due to large body size, animals may become more sensitive to warming because of narrower thermal windows (Pörtner *et al.*, 2008). This may cause high vulnerability of winter-spawning Atlantic cod to warming winter to spring temperatures (Table 6-2). In contrast, adult bigeye, bluefin, and skipjack tuna spawn at high temperature. They need to prevent overheating by moving to cooler (deeper) waters (Lehodey *et al.*, 2011).

Although temperature means are still most commonly used when attributing responses of marine organisms to climate effects, temperature extremes rather than means are most often mediators of effects (e.g., Easterling *et al.*, 2000; Wethey *et al.*, 2011; Wernberg *et al.*, 2013; Figure 6-5). During heat exposure near the borders of the distribution range (including the high intertidal or warming surface waters), reductions in growth, activity, and abundance accompany even small (<0.5°C) shifts in ambient temperature extremes (e.g., Takasuka and Aoki, 2006; Pörtner and Knust, 2007; Nilsson *et al.*, 2009; Neuheimer *et al.*, 2011). Local extinction events follow due to mortality or behavioral avoidance of unfavorable thermal environments (Breau *et al.*, 2011). Shifted species distribution ranges follow temperature clines from high to low, usually along latitudes, a lateral gradient at basin scale (Perry *et al.*, 2005; Poloczanska *et al.* 2013), or a vertical temperature gradient to deeper waters (Dulvy *et al.*, 2008; Figure 6-5B, Box CC-MB, *high confidence*).

Adopting OCLTT principles has enabled modeling studies to project climate effects (6.5), and paleo-studies to explain climate-induced mass extinction events and evolutionary patterns in Earth history (Pörtner *et al.*, 2005; Knoll *et al.*, 2007). For example, long-term observations show that warming affects the body size of marine fishes (*medium confidence*). Assessing effects of warming on body size may be complicated by effects on the animal's energy budget, the changing availability and body size of prey species, community structure, species interactions, or effects of fishing (Genner *et al.*, 2010; Cheung *et al.*, 2013a). Below the thermal optimum, warming causes growth and weight-at-age of some juvenile or younger fish populations to increase (e.g., Brunel and Dickey-Collas, 2010; Neuheimer and Grønkjær, 2012). However, OCLTT predicts that small individuals are more heat tolerant than large ones, in line with observations of falling animal body sizes in warming oceans ((Box 6-1, e.g., Daufresne *et al.*, 2009). This trend is projected to continue into the 21st century (*medium to high confidence*; Cheung *et al.*, 2013a).

Thermal windows of fishes and invertebrates roughly match ambient temperature variability (Figure 6-1) according to climate regime and seasonality (Pörtner and Peck, 2010; Sunday *et al.*, 2012). Sub-Arctic, small or highly mobile species are eurytherms. They function across a wide temperature range, i.e., they have wide thermal windows and distribution ranges, at the expense of higher energetic costs and associated lifestyles (Pörtner, 2002a, 2006). Conversely, high polar species are stenotherms, i.e., they have narrow thermal windows and low energy demand lifestyles, making them sensitive to temperature change. In a warming world, polar stenotherms will be marginalized, with no possibility to escape to colder regions (*high confidence*). However, extinction of polar species has not yet been reported. As marine fishes and invertebrates in the Southern hemisphere are adapted to less variable ocean temperatures than those in the Northern hemisphere (Jones *et al.*, 1999; Figure 6-1), they may generally be more vulnerable to warming extremes than northern ones. Tropical species (with thermal windows of intermediate width) live close to the highest temperatures tolerated by marine animals (Figure 6-6). Vulnerability is, therefore, highest for polar stenotherms, similar or lower for tropical and lowest for temperate species (*high confidence*).

Short-term shifts in thermal thresholds of an individual organism may happen over days and weeks, such as during seasonal acclimatization. Long-term shifts occur over many generations during evolutionary adaptation of a population to cooler or warmer climates (Figure 6-5A, Pörtner, 2006; Pörtner *et al.*, 2008; Eliason *et al.*, 2011). Both acclimatization and adaptation involve adjustments in biochemical characters (membranes, enzymes); however, the capacity to shift those boundaries is limited and depends on the species and the prevailing climate regime (Pörtner *et al.*, 2008, 2012). Ocean acidification, hypoxia, food availability, and stress affect those limits (6.3.5, Figure 6-5A).

Local adaptation may reduce climate vulnerability at the species level, by causing functional and genetic differentiation between populations, thereby enabling the species to cover wider temperature ranges and live in heterogeneous environments. Local adaptation on small spatial scales is particularly strong in intertidal organisms (Kelly *et al.*, 2012). On larger scales, the widening biogeographic and roaming ranges of Northern hemisphere eurytherms into Arctic waters (Pörtner *et al.*, 2008) are supported by the differentiation into populations with diverse thermal ranges, combined with high acclimatization capacity. By contrast, such capacity is small in high polar, e.g. Antarctic species (Peck *et al.*, 2010). Tropical reef fishes undergo rapid warm acclimation across generations

(Donelson *et al.*, 2012) but some may approach animal heat limits. The rates, mechanisms and limits of thermal acclimatization and evolutionary adaptation are poorly understood (*low confidence*).

[INSERT TABLE 6-2 HERE]

Table 6-2: Selected examples of species responses and underlying mechanisms to changing temperature, oxygen level and ocean acidification (OA). References are indicated by superscript numbers and in the footnote.]

Warm- and cold-water coral communities

Tropical corals live in shallow water and differ from most other animals by hosting dinoflagellates (*Symbiodinium sp.*) in their tissues, which provide the host with organic carbon from photosynthesis and with nitrogen and enable the corals to build and sustain carbonate reefs (Box CC-CR). High light, rapid salinity changes and small increases in temperature can trigger ‘coral bleaching’, the loss of symbionts and tissue color. In case of warming, early steps involve shifts in the photosynthetic processing of light, generating Reactive O₂ Species (ROS) that may in turn damage the symbionts (Glynn and D’Croze, 1990; Hoegh-Guldberg and Smith, 1989; Hoegh-Guldberg, 1999; Jones *et al.*, 1998). Mass bleaching is correlated with small temperature anomalies (+1-2 °C of the long-term summer maximum, satellite observations), causing mortalities (Goreau and Hayes, 1994; Strong *et al.*, 2011) and decreasing coral abundance, on average by 1-2 % per year (Bruno and Selig, 2007; Box CC-CR, 30.5.6, *high confidence*).

The degree of impact will depend on the coral reefs’ adaptability to thermal stress and the interaction of multiple drivers (Meissner *et al.*, 2012; Teneva *et al.*, 2012; Box CC-CR). Such capacity is suggested by different heat tolerances among coral genera (Hoegh-Guldberg and Salvat, 1995; Loya *et al.*, 2001), the exchange of genetic clades of *Symbiodinium* with more tolerant varieties (Baker, 2001; Jones *et al.*, 2008), as well as acclimatization phenomena (Howells *et al.*, 2012).

Studies of the thermal sensitivity of deeper-living cold-water corals (without endosymbionts) are scarce. One species, *Lophelia pertusa* responds to about 3°C warming with a three-fold increase in metabolic rate (Dodds *et al.*, 2007), indicating a narrow thermal window in the cold (cf. Pörtner, 2006).

6.3.1.5. Ecosystems

Heat exposure of ecosystem engineers may threaten the existence of a whole ecosystem. During the last warm interglacial period equatorial coral reefs deteriorated and retreated (Kiessling *et al.*, 2012), a finding emphasizing their thermal sensitivity (Veron *et al.*, 2009) and showing that warming oceans can reach temperatures well beyond the upper heat limits of distinct animal groups and marine animals overall (Figure 6-6). In present-day Great Barrier Reef, a large-scale survey found diverse coral types along a climatic gradient, with no consistent response to climatic drivers (Hughes *et al.*, 2012). However, warm-induced bleaching has contributed to the progressive decrease in live coral cover observed over the last decades (Box CC-CR, 30.5.6; De’ath *et al.*, 2012).

Within ecosystems, shifting competitive or trophic interactions, differential risks for species extinctions and, thereby, scenarios of community-level responses to temperature change (Urban *et al.*, 2012; Milazzo *et al.*, 2013) can be traced back to changing differences in the performance of participating animal species (Figure 6-7, e.g., Cairns *et al.*, 2008; Harley, 2011; Pörtner, 2012). Knowledge is insufficient to assess interactions of species from different domains, impeding a deeper understanding of shifting distributions, abundances, community assemblages, and food webs in space and time (Parmesan and Matthews, 2005, *low confidence* in current understanding).

For example, in a coastal microcosm (small-scale, simplified experimental ecosystem) resident heterotrophic bacteria were stimulated by warming more than a laboratory-reared phytoplankton (Wohlers-Zöllner *et al.*, 2011). Also, high- to low-latitude transects in both the North and South Atlantic revealed a shift between cold and warm waters, from photoautotrophs (gaining energy from photosynthesis) to chemo-heterotrophs (Hoppe *et al.*, 2002). Thermal stimulation of bacteria over phytoplankton has biogeochemical implications, for example, microbially-

mediated carbon dioxide flow to the atmosphere might increase (Sarmiento *et al.*, 2010). The principles and wider applicability of these findings require further investigation (*limited evidence, low agreement*, Kirchman *et al.*, 2009).

[INSERT FIGURE 6-7 HERE

Figure 6-7: Role of thermal tolerance and performance of organisms at ecosystem level. (A) Thermal tolerance ranges (Figure 6-5) differ between species across polar, temperate and tropical climate zones, then overlapping between coexisting species. Shifting temperatures and specific effects of additional drivers on the respective performance curves (dashed lines) change the fitness of coexisting species relative to each other as well as their temperature range of coexistence (after Pörtner and Farrell, 2008). Warming alters the timing of seasonal activities (e.g., elicited by spring warming cues) earlier, or can benefit only one of two interacting species (e.g., in predator-prey dynamics or competition) causing shifts in predominance. (B) During climate warming a largely unchanged thermal range of a species causes it to follow its normal temperatures as it moves or is displaced, typically resulting in a poleward shift of the biogeographic range (exemplified for the Northern hemisphere, modified after Beaugrand, 2009). The polygon delineates the distribution range in space and seasonal time; the level of grey denotes abundance. The southern time window of tolerated temperatures shifts to earlier and contracts, while the northern one dilates (indicated by arrows). Species display maximum productivity in low latitude spring, wide seasonal coverage in the center, and a later productivity maximum in the North. The impact of photoperiod (length of daily exposure to light) increases with latitude (grey arrow). Water column characteristics or photoperiod may overrule temperature control in some organisms (e.g. diatoms), limiting northward displacement.]

Observations of shifting distributions and phenologies, reproduction, and range shifts of phytoplankton, zooplankton, other invertebrates, fishes, and seabirds in pelagic and coastal marine ecosystems have at least partly been attributed to temperature-mediated biological responses (*high confidence*, Figure 6-8; Box 6-1; Box CC-MB). In the North Atlantic as a key example, many biological events have been occurring earlier in the year (*robust evidence, high agreement*, Box 6-1, 30.5.1.1.1). Species richness has increased due to shifts in ranges and abundances. In the Norwegian and Barents Seas a time series (1959 to 2006) of four commercial fish species and their zooplankton prey showed that climate shapes population growth rates through complex influences early in life, including direct temperature effects on growth, indirect effects via the biomass of zooplankton prey, and delayed feedback effects through predators (Stige *et al.*, 2010). Differential species responses to temperature and trophic amplification were demonstrated to modify species interactions at five trophic levels, primary producers (phytoplankton), primary, secondary, and tertiary consumers (zooplankton, fishes and jellyfishes), and benthic detritivores (echinoderms and bivalves) (Kirby and Beaugrand, 2009). Also, the responses of various plankton functional groups, such as diatoms, dinoflagellates, and copepods, to warming are not synchronous, resulting in predator-prey mismatches that carry over to higher trophic levels (Figure 6-7A; Edwards and Richardson, 2004; Costello *et al.*, 2006; 6.3.6, *high confidence*). In the intertidal, warming-induced changes in relative species ranges lead to shifts in dominance through competitive interactions and to modifications in predator pressure (Poloczanska *et al.*, 2008; Harley, 2011). Trans-Arctic interchange of species between Atlantic and Pacific has happened repeatedly in warm periods of the Pleistocene (Dodson *et al.*, 2007) and may occur again, now facilitated by ballast transport by enhanced trans-Arctic shipping (*low to medium confidence*).

Warming may increase the risk of disease outbreaks or parasite infections, in marine organisms and ecosystems, and ultimately, humans (Burge *et al.*, 2013; Altizer *et al.*, 2013 *medium confidence*). Some marine pathogens and protist diseases are shifting their distribution poleward as oceans warm (e.g., Baker-Austin *et al.*, 2013; Burge *et al.*, 2013). Climate change may weaken the immune response of hosts, particularly fishes and invertebrates and increase their susceptibility to disease, as observed during warming in coral reefs of the Pacific and Caribbean (Harvell *et al.*, 2009). Global outbreak frequencies of jellyfish aggregations may follow rising sea surface temperatures (SSTs) (*low confidence*, Mills, 2001; Purcell and Decker, 2005), but evidence is inconclusive. Some studies report an increasing trend (Brotz *et al.*, 2012) and others do not support this view (Condon *et al.*, 2013).

In conclusion, organisms live in limited temperature ranges and are sensitive to temperature extremes (*very high confidence*). Temperature governs the biogeography, diversity, development, reproduction, behavior and phenology of marine species as well as the composition of communities in both pelagic and benthic systems and the seasonal timing of relevant processes (phenology) (*very high confidence*). Ecosystems functioning at the coldest temperatures and warm adapted ones existing at their upper thermal limits are more sensitive (*medium confidence*).

[INSERT FIGURE 6-8 HERE

Figure 6-8: Multi-decadal changes in ecosystem structure in the NE Atlantic driven by warming from both anthropogenic climate change and natural climate variability. **A.** Index of temperature change over the North Atlantic (31°N-65°N and 99°W-11°E) reflecting climate change. This index is the first principal component (i.e., explaining 30.5% of observed variability) based on a Principal Component Analysis (PCA) performed on sea surface temperature. **B.** Index of temperature change (17.5% of observed variability) reflecting the Atlantic Multidecadal Oscillation (AMO). The index is the second principal component. **C-D.** Observed mean annual sea surface temperature in the North Sea during 1960-1981 (**C**) and 1988-2005 (**D**). The location of the critical thermal boundary (9-10°C) is indicated by '+'. **E.** Long-term changes in the mean number of warm-temperate pseudo-oceanic species from 1958 to 2005. **F.** Long-term changes in the mean number of temperate pseudo-oceanic species from 1958 to 2005. The period 1958-1981 was a period of relative stability and the period 1982-1999 was a period of rapid northward shifts, indicating that the abrupt ecosystem shift observed in the North Sea was part of a large-scale response of the zooplankton biodiversity to warming temperatures (see **A-D**). Average values are below 1 because they are annual averages. Note that the color bar is 10-fold smaller for warm-temperate pseudo-oceanic species because these species are less frequently observed than their temperate counterparts. Panels A. and B. from Edwards *et al.* (2013), and C. to F. from Beaugrand *et al.* (2008), and Beaugrand *et al.* (2009).]

_____ START BOX 6-1 HERE _____

Box 6-1. An Atlantic Ocean Example: Long-Term Responses of Pelagic Organisms and Communities to Temperature

Long-term observations (6.1.2, 30.5.1.1.1) encompassing the pelagic Northeast Atlantic over a 50-year period and longer (Figures 6-8, 6-9) show changes in the seasonal abundance of phytoplankton, rapid northerly displacements of temperate and subtropical zooplankton (e.g. calanoid copepods), and phytoplankton (e.g. dinoflagellates and diatoms), and the resulting changes in the ecosystem functioning and productivity (Edwards *et al.*, 2001; Beaugrand *et al.*, 2002; Edwards and Richardson, 2004, *high confidence*). The range limit of warm water copepods shifted by 10° north since 1960 (Beaugrand *et al.*, 2009), with attendant mismatch in the seasonal timing of trophic levels (predators and prey) and functional groups (Edwards and Richardson, 2004). Modes of climate variability reflected in climate indices like the Northern Hemisphere Temperature (NHT) and the North Atlantic Oscillation (NAO) over multidecadal periods accompanied these changes (Figure 6-1). In cooler regions, increased phytoplankton activity caused by warming favored growth resulting in the observed increase in phytoplankton biomass, whereas a decrease in nutrient supply would have prevented growth in warmer regions and caused a decrease in biomass (6.3.4; Richardson and Schoeman, 2004). Hinder *et al.* (2012) attributed a recent decline in North Sea dinoflagellates relative to diatoms to warming, increased summer windiness and thus, water column turbulence. The ecosystem response to natural climate variability in the past provides a glimpse into the climate-induced changes of the near future (Figure 6-9).

In regions of high vulnerability to climate, mild warming can trigger rapid and substantial ecosystem shifts, offering a way to anticipate future changes (Figure 6-9). In line with the increased understanding of physiology (6.3.1.1), warming in the temperate to polar North Atlantic was paralleled by a reduction in the average body lengths of about 100 copepod species, from 3-4 mm to 2-3 mm (Beaugrand *et al.*, 2010). Warming was also correlated with an increase in species richness among copepods and within the dinoflagellate genus *Ceratium*. In diatoms, which are major contributors to carbon export (Armbrust, 2009), warming and decreasing annual variability in SST resulted in lower diversity, smaller size and reduced abundance (Beaugrand *et al.*, 2010). Morán *et al.* (2010) found that temperature alone explained 73% of the variance in the contribution of small cells (picophytoplankton) to total phytoplankton biomass in the eastern and western temperate North Atlantic from -0.6 to 22°C. More recently, Marañón *et al.* (2012) analysed data from polar, sub-polar and tropical regions and suggested that nutrient availability may influence cell size more than temperature.

The ecosystem regime shift observed in North Sea plankton in the late 1980s involved an increase in phytoplankton and changes in species composition and abundance among holozooplankton (animals that are planktonic for their entire lifecycle) (Reid *et al.*, 2001; Kirby and Beaugrand, 2009; Kirby *et al.*, 2009; Lindley *et al.*, 2010). This shift

was paralleled by the northward propagation of a critical thermal boundary (CTB, i.e., the boundary of the subpolar gyre) between the temperate and the polar biomes (Box CC-PP, Figure 1; Beaugrand *et al.*, 2008). Warming to above the CTB coincided with pronounced and large-scale variations in phytoplankton productivity, an increase in calanoid copepod diversity (Beaugrand *et al.*, 2008) and herring abundance (Schlüter *et al.*, 2008), a reduction in the mean size of calanoids, and a decrease in the abundance of southern Atlantic cod populations in the North Atlantic Ocean (e.g., the North Sea, Pörtner *et al.*, 2008; Beaugrand *et al.*, 2010). These patterns also extend to the southern North Sea, where elevated salinities and average warming by 1.6°C, both in summer and winter between 1962 and 2007, expanded the time window for growth of microalgae and possibly supported the invasion and increase in numbers of warm-adapted silicified diatoms (Wiltshire *et al.*, 2010). Recent findings indicate a regime shift in the Bay of Biscay, the Celtic and the North Seas, in the mid to end 1990s (Luczak *et al.*, 2011). Changing plankton composition and changing abundances of both sardine and anchovies (Raab *et al.*, 2013) paralleled stepwise warming.

Northward range extensions or redistributions in fishes were largest along the European Continental shelf and attributed to regional warming, e.g. by 1.0°C from 1977 to 2001 in the North Sea, with winter warming being closely correlated with the shift of Atlantic cod (Perry *et al.*, 2005; 6.3.1). Similar trends were observed due to warming by 1–2 °C in the waters south and west of Iceland during the past 15 years (Valdimarsson *et al.*, 2012). In the Northwest Atlantic Arctic and sub-Arctic, winter and spring warming caused expansion of the area matching the thermal optimum of Atlantic salmon at 4–8°C and caused greater growth (Friedland and Todd, 2012). Pelagic sardines and anchovies entered the North Sea in the early to mid-1990s, after about 40 years of absence, in response to intensified NAO and AMO (Alheit *et al.*, 2012). Red mullet and bass extended into western Norway; Mediterranean and north-west African species extended to the south coast of Portugal (Brander *et al.*, 2003; Beare *et al.*, 2004; Genner *et al.*, 2004; 30.5.1.1.4).

In the Northwest Atlantic cooling and freshening occurred during the late 1980s-early 1990s and seemed to have the opposite effect, as capelin and their predator, Atlantic cod, shifted farther south (Rose and O'Driscoll, 2002). Between the early 1990s and mid-2000s in the Northwest Atlantic sub-polar gyre, phytoplankton biomass increased, due to warming. At the same time, Arctic copepod species became more abundant, due to increased influx of Arctic water (Head and Pepin, 2010). Although temperatures have risen on the Newfoundland Shelf (Colbourne *et al.*, 2011), capelin and cod remain scarce for reasons probably unrelated to climate (DFO 2011a, b). Farther south, Arctic freshwater inflows caused freshening and increased stratification of the area around the Gulf of Maine throughout the 1990s, resulting in enhanced phytoplankton abundance, a larger and later fall bloom, increased abundance of small copepods, and a decrease in the large copepod *Calanus finmarchicus* (deYoung *et al.*, 2004; Pershing *et al.*, 2005, 2010). Various fish species showed poleward shifts in distribution (Table 6-2) which were associated with reduced survival of larval cod (Mountain and Kane, 2010) and fewer right whale calves (Greene *et al.*, 2003), but increased herring abundance (Greene and Pershing, 2007).

[INSERT FIGURE 6-9 HERE

Figure 6-9: Schematic depiction of observed effects of ~1°C ocean warming in the northern North Atlantic driven by climate variability (A, C) versus effects expected from anthropogenic climate change (B, D). A) Transient warming and cooling associated with AMO variability (Drinkwater, 2006), based on the Kola Section temperatures (0-200 m; Stations 3-7, 71.5°-72.5°N, 33.5°E) in the Barents Sea obtained from <http://www.pinro.ru> and filtered using a 20-year running mean. Similar trends occurred across most of the northern North Atlantic although the amplitude and timing of the peaks and troughs varied spatially. B) Warming driven by climate change for the same region (RCP 8.5 simulations averaged from CMIP5 models, computed as the mean over the upper 200 m in the grid box (2.5x2.5 degrees) centered at 71.25N and 33.75E). C) Warming and subsequent cooling in the northern North Atlantic during the period shown in A resulted in complex multi-faceted changes (shown schematically) in NPP (green curve), zooplankton biomass (red curve) and fish stock abundances (blue curve). There was a general poleward shift and range expansion of many commercial (e.g., Atlantic herring, Atlantic cod, haddock) and non-commercial species, reversed during the subsequent cooling period (denoted by black dashed curve). Polewards shifts in spawning areas (e.g., Atlantic cod) were also reversed as the waters cooled. The purple and grey dashed curve represents shifts in seasonal timing (phenology) and community composition, respectively, that were influenced by earlier arrivals and later retreat of migratory fish (not shown). For more details see Drinkwater (2006). D) Projected effects of climate mediated warming on northern sub-polar and polar biota based on model projections of altered NPP (Bopp *et al.*,

2013), and of the range shift of exploited fishes and invertebrates (Cheung *et al.*, 2009, 2013a). The projected trends in panel D will differ with latitude, e.g., decreased NPP at lower latitudes and no significant change to NPP in temperate waters (Bopp *et al.*, 2013). Higher NPP supported and is projected to support higher trophic levels at high latitudes (C, D, 6.3.4). Note that climate variability will be superimposed on anthropogenic warming (B, see Figure 6-1, 6-8A,B). Dotted lines indicate projected changes to continue beyond the range of historical observations.]

_____ END BOX 6-1 HERE _____

6.3.2. CO₂ Effects

Evidence for biological effects of ocean acidification (OA) stems from paleo-observations (6.1.2), few observations in the field (6.3.2.5), studies at volcanic CO₂ seeps as natural analogues, and mostly from short- to medium-term (hours to months) experiments in the laboratory or field, exposing organisms to projected future CO₂ levels (6.3.2.1-4). A surging number of studies is providing evidence that rising CO₂ levels will increasingly affect marine biota and interfere with ecological and biogeochemical processes in the oceans (*high confidence*, FAQs 6.2, 6.3).

6.3.2.1. Principles

The absorption of rising atmospheric CO₂ by oceans and organisms changes carbonate system variables in the water and in organism internal fluids, i.e., the relative proportions of CO₂, carbonate, bicarbonate, and hydrogen ions (pH). Internal pH must be tightly controlled, as some processes, such as calcification, release protons thereby affecting pH and as other biochemical processes are pH-sensitive. Accumulation of CO₂ and the resulting acidification can also affect a wide range of organismal functions, such as membrane transport, calcification, photosynthesis in plants, neuronal processes in animals, growth, reproductive success, and survival. Effects translate from organism to ecosystem.

The capacity of organisms to resist and compensate for the CO₂ induced acidification of internal fluids depends on acid-base regulation, i.e., the capacity of ion exchange to accumulate bicarbonate internally, an aspect unexplored in many phyla (Figure 6-10A, *low to medium confidence*; e.g., animals: Heisler, 1986; Pörtner, 2008; Claiborne *et al.*, 2002; phytoplankton: Taylor *et al.*, 2011) (FAQ 6.3).

In unicellular microbes the regulation of intracellular pH may play a key role in modulating CO₂ responses (Taylor *et al.*, 2011). Findings in invertebrates and fish indicate an additional role for extracellular pH (Figure 6-10A); effective pH values may vary between species. Organisms pre-adapted to elevated CO₂ may minimize the decrease in pH (acidosis). They may also modify their sensitivity such that they respond less or not at all to the acidosis. Recent evidence, however, emphasizes a role for acid-base regulation in a natural low-pH setting. Between two urchin species, only the one successful in maintaining its setpoints of extracellular pH is able to settle close to volcanic CO₂ seeps (Calosi *et al.*, 2013). Compensating for the acidosis may cause increased energy demand and respiration rates. In general, such capacity rises with metabolic energy turnover, e.g., it is higher in more active marine animals, such as fishes, cephalopods and pelagic copepods, and in mobile coastal crabs compared to sessile species (Table 6-3, Pörtner *et al.*, 2005, 2011; Ishimatsu *et al.*, 2008; Melzner *et al.*, 2009; Ishimatsu and Dissanayake, 2010). This matches the sensitivity distribution seen among animals at the phylum level (Figure 6-9B, *medium confidence*).

Some species have lower metabolic rates in response to acidosis (Pörtner *et al.*, 1998; Michaelidis *et al.*, 2005; Pörtner, 2008; Liu and He, 2012; Navarro *et al.*, 2013); others display increased energy turnover and food ingestion rates, possibly indicating a capacity to resist acidification effects (Parker *et al.*, 2011; Saba *et al.*, 2012). The effects of the acidosis on various processes relevant to fitness may explain changes in whole-organism energy demand, probably paralleled by modified ion exchange, protein synthesis, growth and feeding rates. The magnitude of effect depends on the CO₂ concentrations reached (Figure 6-10B).

The internal formation of carbonate from bicarbonate is essential to calcification, which is the formation of solid CaCO_3 in internal or external calcified structures, used for defense and structural support. Calcification usually occurs in separate body or cell compartments, where pH and thus, CO_3^{2-} concentration and saturation Ω (6.1.1) are maintained at values higher than in other body fluids or ambient water (Trotter *et al.*, 2011; Taylor *et al.*, 2011; McCullough *et al.*, 2012; Venn *et al.*, 2013). CO_2 impedes the formation of carbonate such that calcification rate decreases. It may be maintained by enhanced transport of ions, incurring elevated energetic costs (Figure 6-10).

External carbonate structures like shells rely on ambient seawater being supersaturated with carbonates. Decreasing oceanic carbonate levels reduce the saturation levels (Ω) of calcite or aragonite in the water. Reduction to below unity may lead to the corrosion of carbonate shells (FAQ 6.3). However, many species protect their shells from direct contact with seawater by various types of organic coating (e.g., a periostracum in mollusks and brachiopods, an epicuticle covering the carapace of crustaceans; an epidermis covering the tests of urchins, epithelial tissue covering aragonite in corals, coralline algae precipitating calcium carbonate (mostly Mg-calcite) within their cell wall). A meta-analysis of the effects of ocean acidification on biological processes indicates that reductions in the rate of net calcification (calcification minus dissolution) and survival are the most uniform responses across organisms studied, relative to other, more variable impacts such as reduced growth, development, and abundance (Box CC-OA, Kroeker *et al.*, 2013).

Some organisms benefit from elevated CO_2 partial pressures ($p\text{CO}_2$). Photosynthesis and/or nitrogen fixation in selected microorganisms are impacted by OA, but effects are species- or taxon-specific, possibly depending on how they acquire carbon (i.e., the presence and in particular the type, capacity, and energetic costs of carbon-concentrating mechanisms (CCM's, Giordano *et al.*, 2005; Kranz *et al.*, 2011).

A comprehensive picture of responses to CO_2 requires consideration of variable sensitivities between species and life stages and taxon-specific sensitivity distributions, as shown by a meta-analysis of animal data (Figure 6-10B, Wittmann and Pörtner, 2013). Echinoderms, bivalves, gastropods, and corals begin to respond negatively at lower CO_2 levels than crustaceans or cephalopods (Figure 6-10B). This sensitivity pattern resembles the one seen in the Permian mass extinction (Knoll *et al.*, 2007; Knoll and Fischer, 2011). The picture for fishes is less clear, as the present findings of high vulnerability are not met by similar observations in the fossil record. Evolutionary adaptation may thus eliminate or minimize reported effects.

The capacity for pH and ion regulation and other relevant processes can be upregulated by gene expression, as seen in acclimation studies in echinoderm larvae (O'Donnell *et al.*, 2010; Martin *et al.*, 2011) and fishes (Deigweier *et al.*, 2008; Tseng *et al.*, 2013), in warm-water coral branches (Kaniewska *et al.*, 2012) but not in a study of warm-water coral larvae (Moya *et al.*, 2012). Few studies address whether and to what extent species undergo evolutionary adaptation to high $p\text{CO}_2$, as seen in the coccolithophore *Emiliania huxleyi* over 500 asexual generations (Lohbeck *et al.*, 2012). In organisms with longer generation times, perturbation studies in the laboratory measure tolerance and acclimation, but not adaptation or natural selection. Animal adaptation is accelerated by high functional variability among larvae, enabling selection of resistant genotypes (*low to medium confidence*, Parker *et al.*, 2012; Sunday *et al.*, 2011; Pespeni *et al.*, 2013). This may explain the selective mortality seen in Atlantic cod larvae under elevated CO_2 (Frommel *et al.*, 2012). Both acclimatization and adaptation will shift sensitivity thresholds but the capacity and limits of species to acclimatize or adapt remain largely unknown and hence impacts of acute exposures cannot easily be scaled up to effects on the longer, evolutionary timescales of ocean acidification (Wittmann and Pörtner, 2013). Observations in ecosystems characterized by permanently elevated or fluctuating CO_2 levels, like upwelling areas, OMZs (6.1.1), or seeps, reflect the existence of sensitivity thresholds (*high confidence*, 6.3.2.5) but organisms may have evolved a higher resistance to increased CO_2 levels than elsewhere (*low confidence*).

[INSERT FIGURE 6-10 HERE

Figure 6-10: A: Responses of a schematized marine animal (left) and a phytoplankter (right) to ocean acidification (OA). Effects are mediated via diffusive CO_2 entry (black arrows) into body and cell compartments, resulting in a rise in $p\text{CO}_2$ (highlighted in red), a drop in compartmental pH (highlighted in blue) and their effects (red arrows) on various processes (red text) in tissues and cellular compartments, as well as on calcium carbonate saturation state (Ω) at calcification sites (after Pörtner, 2008; Taylor *et al.*, 2011). Variable sensitivity relates to the degree of pH decline and compensation, depending on the capacity of pH and ion regulation. B: Distribution of sensitivities across

species within animal phyla, under progressively rising water CO₂ levels, as percent of studied scleractinian coral, echinoderm, molluscan, crustacean and fish species affected negatively, positively or not at all (for effects considered see text). As not all life stages, variables and *p*CO₂ ranges were covered in all species two assumptions partially compensate for missing data: 1) Negative effects at low *p*CO₂ will remain negative at high *p*CO₂. 2) A positive or neutral outcome at both low and high *p*CO₂ will be the same at intermediate *p*CO₂. As responses reported for each species vary for each CO₂ range, variable species numbers result (on top of columns). The total number of species studied in a group is shown as the number above the control column. Horizontal lines above columns represent frequency distributions significantly different from controls (Wittmann and Pörtner, 2013). C: Areas with reported annual catches of marine calcifiers (crustaceans and mollusks) >0.001 t km⁻², depicted on a global map showing the distribution of ocean acidification in 2100 according to RCP8.5 (WGI AR5 SPM) as well as the distribution of warm-water (green dots) and cold-water coral communities (pink dots).]

Table 6-3 compiles effects of ocean acidification observed across taxa in laboratory and field experiments. The latter include studies in mesocosms and at natural analogues, submarine CO₂ venting areas at locales like Ischia, Italy (Hall-Spencer *et al.*, 2008), Papua New Guinea (Fabricius *et al.*, 2011), and Puerto Morelos, Mexico (Crook *et al.*, 2012). It should be noted that anthropogenic CO₂ accumulation according to RCPs adds to the natural variability of CO₂ concentrations in marine environments. Many groups, especially sessile or non-photosynthetic calcifiers have sensitive species with vulnerability thresholds surpassed under RCP6.0 by 2100 (*low to medium confidence*).

Recent meta-analyses also summarize OA effects, two for biogeochemical processes and relative effect sizes (Kroeker *et al.*, 2013; Harvey *et al.*, 2013), one for the distribution of sensitivity between species within major animal phyla and its change depending on ambient *p*CO₂ (Figure 6-10, Wittmann and Pörtner, 2013). All of these analyses consider the interaction of warming and CO₂ accumulation (6.3.5). Present limitations in understanding the mechanisms of effect and their long-term persistence compounds accurate projections of the long-term effects of OA (*medium confidence*, Wittmann and Pörtner, 2013).

[INSERT TABLE 6-3 HERE]

Table 6-3: Tolerances to ocean acidification in marine taxa, assessed from laboratory and field studies of species in the *p*CO₂ range from <650 to >10 000 μatm, compared to present day atmospheric levels of 400 μatm. (It should be noted that anthropogenic CO₂ emissions add to the natural variability of CO₂ concentrations in marine environments, which can reach much higher than atmospheric levels). Variables studied include growth, survival, calcification, metabolic rate, immune response, development, abundance, behavior and others. Neither all life stages, nor all variables, including the entire range of CO₂ concentrations, were studied in all species. *Confidence* is based on the number of studies, the number of species studied and the agreement of results within one group. +: denotes that possibly more species or strains (genetically distinct populations of the same species) were studied, as only genus or family were specified; beneficial: most species were positively affected; vulnerable: more than 5% of species in a group will be negatively affected by 2100; tolerant: more than 95% of species will not be affected by 2100. RCP 6.0: representative concentration pathway with projected atmospheric *p*CO₂ = 670 μatm; RCP 8.5: *p*CO₂ = 936 μatm in 2100 (Meinshausen *et al.*, 2011). *Confidence* is limited by the short- to medium-term nature of various studies and the lack of sensitivity estimates on evolutionary timescales, i.e., across generations (see separate reference list, online supplementary material). Note that the assessment of variability between species from the same animal phylum has revealed an increase in the fraction of sensitive species with rising CO₂ levels, see Figure 6-10. *Rather than a sensitivity threshold the entire range of investigated *p*CO₂ values is given for groups of photosynthetic organisms. In all studies photosynthetic rates are stimulated to different, species-specific degrees by elevated *p*CO₂, indicating low vulnerability. Coccolithophores and calcifying algae are assessed as being more sensitive than other photosynthetic organisms due to reduced calcification and shell dissolution. NA, not available, ⁺confidence levels for fishes were converted from medium to low, in light of uncertainty on the long-term implications of behavioral disturbances.]

6.3.2.2. Microbes

The physiology of both calcifying (coccolithophores) and non-calcifying phytoplankton can be influenced by changes in carbonate system variables caused by ocean acidification (Figure 6-10A). Growth and photosynthetic rates of

diatoms in laboratory cultures are considered relatively insensitive to elevated CO₂ (Rost *et al.*, 2003; Trimborn *et al.*, 2008). Dinoflagellate sensitivity to elevated CO₂ is poorly studied (Hansen *et al.*, 2007), but in one species carbon fixation rates were enhanced at 750 μatm CO₂ while growth remained unaffected (Fu *et al.*, 2008). Indirect effects of ocean acidification on phytoplankton physiology include altered availability of trace metals needed for many biochemical cycles (Hoffmann *et al.*, 2012).

Harmful algal blooms are a growing problem in coastal waters worldwide (6.4.2.3), and many of the various phytoplankton species that produce bio-accumulated toxins are sensitive to changes in the seawater carbonate buffer system (Hallegraeff, 2010, Fu *et al.*, 2012). For example, the dominance and community structure of harmful bloom dinoflagellates can be profoundly altered by changing pCO₂ (Tatters *et al.*, 2013), and both toxic dinoflagellates and diatoms have been shown to produce higher toxin levels under near-future levels of ocean acidification (Fu *et al.*, 2010, Sun *et al.*, 2011).

Some planktonic N₂-fixing cyanobacteria (diazotrophs), e.g., strains (genetically distinct populations of the same species) of offshore cyanobacteria of the genera *Trichodesmium* and *Crocospaera*, respond to rising CO₂ with increased rates of both carbon and N₂ fixation (Fu *et al.*, 2008; Lomas *et al.*, 2012). In contrast, laboratory studies using the bloom-forming cyanobacteria *Nodularia* (an organism largely found in coastal stratified, eutrophic waters) revealed decreased growth and N₂ fixation under elevated CO₂ conditions (Czerny *et al.*, 2009). The wide range of responses in N₂ fixation (e.g., Hutchins *et al.*, 2007; Levitan *et al.*, 2007; Kranz *et al.*, 2010) may be explained by different CO₂ affinities (i.e., dependences of growth rates on CO₂ concentration) of a range of N₂-fixing cyanobacteria (*Trichodesmium* and *Crocospaera*) from different oceanic biomes. Some species/strains operate at close to maximum growth rates at present-day oceanic CO₂ levels, whereas others had sub-optimal growth rates under these conditions (Hutchins *et al.*, 2013). To date, the physiological mechanisms underlying these responses remain unknown, especially in open-ocean nitrogen fixers. Cyanobacteria may reallocate energy from their energetically expensive CCMs toward N₂ fixation and the acquisition of growth limiting nutrients (Kranz *et al.*, 2010; Levitan *et al.*, 2010), but evidence for such diversion of energy is lacking. Whether nitrogen fixation will increase with progressive ocean acidification remains to be explored (*low confidence, limited in situ evidence, medium agreement*).

The responses of coccolithophore calcification to OA are species-specific and highly variable. The function(s) of calcification are not well understood, making it difficult to evaluate the consequences of lowered calcification (e.g., Rost *et al.*, 2008). Reductions, increases, and unchanged calcification rates (and shell structure) have all been found in different coccolithophore species for RCP 8.5 CO₂ conditions projected around 2100 (Riebesell *et al.*, 2000; Zondervan *et al.*, 2001; Langer *et al.*, 2006; Iglesias-Rodriguez *et al.*, 2008). Calcification in coccolithophores is species- (Langer *et al.*, 2006) and in *Emiliana huxleyi* even strain-specific (Langer *et al.*, 2009, 2011; Hoppe *et al.*, 2011). It thus remains unclear whether OA will result in exoskeletons that are insufficiently calcified for sustained structural support and protection in coccolithophores (*medium evidence, low agreement*).

Foraminifera display decreasing calcification and shell weight under elevated CO₂ (Lombard *et al.*, 2010). Changes in historical specimens (Moy *et al.*, 2009, see below) and during glacial-interglacial cycles (Barker and Elderfield, 2002) support projections of future reductions in net calcification by foraminifera (*medium to high confidence*).

6.3.2.3. Macroalgae and Seagrasses

Primary production, shoot density, reproductive output, and below-ground biomass of seagrasses generally respond positively to elevated pCO₂, indicating CO₂ limitation of their productivity. Such effects were identified in both laboratory and field above 720 to 1800 μatm (*high confidence*, e.g., Palacios and Zimmerman, 2007; Hall-Spencer *et al.*, 2008; Andersson *et al.*, 2011, cf. 5.4.2.3). Production, growth, and recruitment of most but not all non-calcifying seaweeds also increased at CO₂ levels from 700 to 900 μatm (RCP8.5; Porzio *et al.*, 2011; Kroeker *et al.*, 2013). Some non-calcifying seaweeds and seagrasses will thus benefit from future ocean acidification (*high confidence*) but OA exposes them to higher than usual grazing as a consequence of losing deterrent phenolic substances (*low confidence*, Arnold *et al.*, 2012).

Calcifying algae (corallines) show complex and species-specific responses of photosynthesis to elevated CO₂, but calcification is impacted once species-specific *p*CO₂ thresholds are surpassed (*medium confidence*, Anthony *et al.*, 2008; Martin and Gattuso, 2009). At habitat temperature calcification by temperate coralline red and calcareous green algae increased at CO₂ levels up to 900 μatm and only decreased at the highest concentration applied (2850 μatm), but did not fall below rates found at present-day *p*CO₂ (Ries *et al.*, 2009). During 3 months of exposure, growth of *Lithothamnion glaciale*, a cold-water calcareous red alga, decreased progressively with rising CO₂ levels, and its structural integrity was weakened beyond 590 μatm (Ragazzola *et al.*, 2012), potentially influencing ecosystem function. Some calcifying algae may thus be impacted by future ocean acidification (*medium confidence*).

6.3.2.4. Animals

Studies of marine animals and their life stages show a high diversity and variability of processes affected by ocean acidification. Many variables studied reflect physiological performance (O₂ consumption, exercise, behavior, calcification, growth, immune response, acid-base balance, gene expression, fertilization, sperm motility, developmental time, production of viable offspring, and morphology; Table 6-3, Figure 6-10). In some species growth may be stimulated by OA, in others depressed, or unaffected (cf. Gooding *et al.*, 2009; Munday *et al.*, 2009a, 2011a; Dupont *et al.*, 2010). The degree of CO₂-induced acidosis and its compensation by ion exchange may shape sensitivity (6.3.2.1). Full exploitation of the ability to resist *p*CO₂ increases depends on the availability and high quality of food and the strengthening of fitness (Gooding *et al.*, 2009; Melzner *et al.*, 2011). However, food quality of prey organisms may decrease under elevated *p*CO₂. For example, slower reproduction and growth of the copepod *Acartia tonsa* under 760 μatm *p*CO₂ was related to the decreasing quality of its diatom food (Rossoll *et al.*, 2012).

Changes in calcification rates reported from CO₂ manipulation experiments vary widely. Reduced calcification and weakened calcified structures were seen under elevated *p*CO₂ in corals (see below), echinoderms (Kurihara and Shirayama, 2004), mollusks (Gazeau *et al.*, 2013), and larval crustaceans (Arnold *et al.*, 2009; Walther *et al.*, 2011). Some adult limpets and urchins increased calcification rates at *p*CO₂ from 600 to 900 μatm, before it fell at even higher *p*CO₂. In some adult crabs, lobsters, and shrimps calcification rates increased further with rising *p*CO₂ (Ries *et al.*, 2009). Stronger internal structures such as cuttlebones and otoliths resulted from enhanced calcification under elevated *p*CO₂ in juvenile cuttlefish (cephalopods, Gutowska *et al.*, 2008) and fishes (Checkley Jr *et al.*, 2009; Munday *et al.*, 2011b), with unclear impacts on fitness. Energy costs in epithelia or calcification compartments may be enhanced by elevated *p*CO₂ causing a stimulation of metabolism (6.3.2.1). In some cases, this may indicate imbalances in energy budget rather than increased CO₂ resistance, for example, if costs are down-regulated in muscle or liver. Enhanced calcification can then occur at the expense of growth (*medium confidence*, Wood *et al.*, 2008; Beniash *et al.*, 2010; Thomsen and Melzner, 2010; Parker *et al.*, 2011).

Studies on calcifying zooplankton focused on pteropods (planktonic mollusks with aragonite shells). These form an integral part of the food web, both as grazers and prey, e.g., for pink salmon (Armstrong *et al.*, 2005; Hunt *et al.*, 2008). In the Subarctic, the Arctic, and the Southern Ocean, pteropods will reduce calcification in response to OA until at least the end of the century (*medium confidence*, Orr *et al.*, 2005; Comeau *et al.*, 2009; Lischka *et al.*, 2011).

Elevated CO₂ causes behavioral disturbances in fishes (studied mostly in larvae and juveniles, Munday *et al.*, 2010; Ferrari *et al.*, 2011; Domenici *et al.*, 2012; Jutfeld *et al.*, 2013) through neural mechanisms (Nilsson *et al.*, 2012). The long-term persistence and evolutionary relevance of these behavioral effects need further study before general conclusions can be drawn (*low confidence*, Wittmann and Pörtner, 2013, Table 6-3).

6.3.2.4.1. Animal life cycles

It is generally held that early lifestages are always more sensitive to environmental stress than adults. In the context of ocean acidification this statement is supported by findings like larval oyster fatalities in aquaculture caused by upwelled CO₂-rich waters (*high confidence*; Barton *et al.*, 2012). A key aspect may also be that larvae growing or developing more slowly under elevated CO₂ as in various groups including fishes (Baumann *et al.*, 2012, 6.3.2.1) may encounter enhanced mortalities due to prolonged predator exposure. Comparative studies of animal sensitivities

to OA over a complete life cycle or during critical transition phases (e.g. fertilization, egg development and hatching, metamorphosis, moulting) are scarce and do not support generalized conclusions (*low confidence*).

Effects of elevated CO₂ on one life stage or transition phase may affect or carry over to the next one. Moulting success into the final larval stage was reduced in a crab species (Walther *et al.*, 2010). In a sea urchin species, negative impact was found to accumulate during 4 months acclimation of adults reducing reproductive success. This impact was, however, compensated for during extended acclimation of female urchins for 16 months (Dupont *et al.*, 2012). Negative impact was still transferred from urchin larvae to juveniles under elevated *p*CO₂. Conversely, adult oysters acclimated to high CO₂ acquired resistance which was carried over to their offspring (Parker *et al.*, 2012). More long-term acclimation studies to realistic emission scenarios are needed for generalized conclusions. Furthermore, the proposition that juvenile lifestages are always more sensitive than adults needs thorough re-investigation in the context of ocean acidification, especially in the context of the notion that larvae may provide a selection pool for survival of the most suitable phenotypes (6.3.2.1, *low confidence*).

6.3.2.4.2. Warm- and cold-water coral communities

In warm-water reef-building corals, OA causes genus-specific reductions in calcification (Leclercq *et al.*, 2002; Langdon and Atkinson, 2005; Kleypas and Langdon, 2006). Nutrient availability to symbionts may sustain calcification. Heterotrophic feeding by the corals also supports energy-dependent calcification and acid-base regulation and thus, resilience (Edmunds, 2011; Figure 6-10). Females may sacrifice calcification more than males due to energetic tradeoffs with reproduction (Holcomb *et al.*, 2012). Warm-water corals are sensitive to future OA (*high confidence*, Table 6-3)

The cold-water coral, *L. pertusa* shows resilience to ocean acidification. In short-term ship-board incubations pH reductions between 0.15 and 0.3 units (540 and 790 μ atm) led to calcification rates reduced by 30-56 % (Maier *et al.*, 2009), especially in young, fast growing polyps. However, net calcification was maintained at seawater aragonite saturation <1. Exposure to a *p*CO₂-induced pH reduction by 0.1 units or even to the projected end of century *p*CO₂ of 930 μ atm led to calcification rates being maintained over six to nine months (Form and Riebesell, 2012; Maier *et al.*, 2013). This ability is probably due to a regulated upward shift of pH and carbonate saturation at organismal calcification sites (Figure 6-10; McCulloch *et al.*, 2012). Natural distribution of other cold-water species covers wide natural pH gradients in Chilean fjords (*Desmophyllum dianthus*; Jantzen *et al.*, 2013) and ranges into waters with undersaturated carbonates as in Australian waters (4 scleractinian corals, Thresher *et al.*, 2011). Pre-adaption to elevated *p*CO₂ apparently exists, however species vulnerabilities to further increases in *p*CO₂ have not been investigated. Again, vulnerability is species specific, colonial scleractinians may be limited to water saturated or near-saturated with aragonite, whereas others are not (Thresher *et al.*, 2011). Conclusions on the relative vulnerability of the group appear premature (Table 6-3). To what extent a further lowering of carbonate saturation values will influence the future distribution of various calcite or aragonite forming cold-water corals is not clear (*low confidence*, Guinotte *et al.*, 2006).

6.3.2.5. Ecosystems

For insight into ecosystem level processes, laboratory studies have been supplemented with experimental studies in large volume mesocosms (i.e., > 1000 L) and in the field, and with long- term field observations. Together they inform the debate over the attribution of field observations to ocean acidification.

6.3.2.5.1. Evidence from field observations

Contributions of anthropogenic ocean acidification to climate-induced alterations in the field have rarely been established and are limited to observations in individual species (see also 30.5.1.1.3). Shell thinning in modern planktonic foraminifera (collected 1997 to 2004) in the Southern Ocean compared to those from the Holocene and before was attributed to anthropogenic ocean acidification (Moy *et al.*, 2009). Both anthropogenic OA and the

upwelling of CO₂-rich deep waters (30.5.4.1.4) were held responsible for shell thinning in planktonic foraminifera in the Arabian Sea over the last century (de Moel *et al.*, 2009) or in live pteropods collected in 2008 in the Southern ocean (Bednaršek *et al.*, 2012; *medium evidence, medium agreement*). However, no changes were observed in a 57-year record of the composition and abundance of calcifying zooplankton in the increasingly acidified California Current System (Ohman *et al.*, 2009). Possible explanations for the absence of significant responses in some studies include insufficient lengths of time series (6.1.2), organisms being preadapted to naturally high CO₂ in upwelling or other systems, linked to a low signal-to-noise ratio, or the difficulty of detecting small OA effects in comparison with larger ecosystem effects of other drivers such as temperature, e.g., in calcifying plankton, Beaugrand *et al.* (2013). Similarly, declines in coral calcification and performance in the field (De'ath *et al.*, 2009) were attributed to thermal extremes, but may also include an as-yet unclear contribution from OA.

6.3.2.5.2. *Microbial communities and nutrient cycles*

Laboratory experiments, coastal mesocosm studies (Weinbauer *et al.*, 2011) and field experiments (Beman *et al.*, 2011; Law *et al.*, 2012) have yielded various, sometimes conflicting results on the effects of CO₂ on microbial processes. From a meta-analysis of available data, Liu *et al.* (2010) conclude that the rates of several microbial processes will be affected by OA, some positively, others negatively. The potential of the microbial community to adapt to ocean acidification and maintain functionality, either by genetic change at the species level or through the replacement of sensitive species or groups at the community level, remains to be explored further. At the present time there is insufficient field-based evidence to conclude that elevated CO₂ will affect natural assemblages of microorganisms (*limited evidence, low agreement*) with the possible exception of the negative impact on calcification (Joint *et al.* 2011).

Experimental studies on OA effects (through reduced pH or increased CO₂) on autotrophic and heterotrophic microbial production have provided inconsistent results. Microbes are characterized by large diversity and broad environmental adaptation, and hence may respond to environmental challenges by exploiting such diversity via species replacements (Krause *et al.*, 2012). This makes it difficult to project the findings of laboratory experiments investigating the response of microbes to OA to the ecosystem level. Relevant variables include cellular elemental stoichiometry (C-N-P ratios, Riebesell, 2004; Fu *et al.*, 2007), rates of CO₂ and N₂ fixation (Riebesell, 2004; Hutchins *et al.*, 2007; Hutchins *et al.*, 2009), rates of nitrification (Beman *et al.*, 2011), changes in the proportion of dissolved (i.e., DOC) to particulate photosynthate produced during carbon fixation (Kim *et al.*, 2011), and the response of viruses (Danovaro *et al.*, 2011).

Field experiments led to the projection that nitrification rates (ammonia oxidation to nitrite and nitrite oxidation to nitrate) of bacteria and archaea will be reduced by 3–44% when pH is reduced by 0.05–0.14 (Beman *et al.*, 2011), corresponding to a mean rise in CO₂ by ~100 μatm. The reported decrease in nitrification occurred regardless of natural pH variability, providing no evidence for acclimation of the nitrifiers to reduced pH, e.g., in upwelling areas. Potential changes in microbial cell abundance, possibly as a result of lower cellular nitrification rates, could further decrease the total rate of nitrification.

It remains unclear whether OA has contributed to the systematic changes in phytoplankton abundance and community structure observed over recent decades, which have largely been attributed to warming (Chavez *et al.*, 2011). In natural assemblages from coastal and polar waters, NPP is stimulated by increased CO₂ (*medium confidence*; Riebesell *et al.*, 2008; Tortell *et al.*, 2008). Small differences in CO₂ sensitivity may lead to pronounced shifts in the dominance of species (Tortell *et al.*, 2008; Beaufort *et al.*, 2011). Quantification of the calcite mass of the coccolithophore community in the present ocean and over the last 40,000 years were in large part attributed to shifts between differently calcified species and morphotypes according to carbonate chemistry (Beaufort *et al.*, 2011). The same study, however, also observed heavily calcified *E. huxleyi* morphotypes in upwelling systems characterised by low pH, a finding which highlights the complexity of assemblage-level responses and may indicate pre-adaptation to elevated *p*CO₂. Due to the complex response patterns, it is not possible to project ecosystem-level effects from effects on coccolithophore calcification in monospecific culture experiments (*low confidence*). Projections of OA impacts on phytoplankton become even more complicated by synergistic interactions with other drivers (Boyd, 2011; 6.3.5).

6.3.2.5.3. *Macrophytes and macrofauna*

Macrofauna and macrophyte communities have been studied in mesocosms and in ecosystems exposed to shifted upwelling regimes or at natural volcanic CO₂ vents (Kroeker *et al.*, 2011; Fabricius *et al.*, 2011). The latter are considered as natural analogues of future ocean acidification. An eight-year trend of (variable) pH decline in upwelled waters along the Northeast Pacific coast was paralleled by shifts in community composition, where shelled species like mussels were replaced by fleshy algae and barnacles (Wootton *et al.*, 2008). Macrofaunal calcifiers at CO₂ vents (Hall-Spencer *et al.*, 2008; Fabricius *et al.*, 2011) and in mesocosms (Christen *et al.*, 2013) display a lowering of species richness. These findings suggest that non-calcifiers increasingly outcompete calcifiers once pH_T decreases to a mean of 7.8 to 7.7 (*medium confidence*). Finally, a loss of calcifiers from mesocosms occurred around 0.5 units below the pH values expected from OA under RCP 8.5 by 2100 (*medium confidence*, Christen *et al.*, 2013). At CO₂ seeps, calcitic bryozoans replace coralline algae, which have more soluble high-calcite skeletons (Martin *et al.*, 2008). Seagrasses and non-calcifying algae gain a competitive advantage (Fabricius *et al.*, 2011). Coral communities exposed to high pCO₂ waters (from upwelling or seeps) have lower growth, calcification and biodiversity (Manzello *et al.*, 2008; Fabricius *et al.*, 2011), resulting in a shift from net accretion to erosion (Box CC-CR). The use of seeps as analogues of future OA is limited as pH variability is high at these sites, such that effective values may be lower than indicated by the average change (Hall-Spencer *et al.*, 2008; Porzio *et al.*, 2011). During periods of high pH at the seeps, they are recolonized by invertebrates and fishes from neighboring areas with normal pH, compromising assessments of long-term sensitivity thresholds. Overall, findings available from mesocosms and natural analogues indicate losses in diversity, biomass and trophic complexity of benthic marine communities due to elevated CO₂ (*high confidence*) and support the projection of similar shifts in other systems with continued OA (*medium confidence*).

Enhanced freshwater input by poorly buffered rivers or by precipitation, into estuaries, brackish oceans like the Baltic (30.5.3.1.4), and into freshening polar oceans, reduces salinity and alkalinity at rising atmospheric pCO₂ and thereby, alters the carbonate system and enhances OA (6.1.1). Estuaries usually have OMZs, where background pCO₂ is elevated. Its reduction by dilution causes the acidification effect to be somewhat less. Enhanced pH reduction and variability in hyposaline waters may constrain the distribution of sensitive species further (*low confidence*, Miller *et al.*, 2009; Denman *et al.*, 2011).

6.3.2.5.4. *Conclusions*

Natural analogues, laboratory and mesocosm experiments provide evidence for differential effects of ocean acidification on species and communities. Sensitivity to OA is species-specific (*high confidence*); differential sensitivities and associated shifts in performance and distribution will change predator-prey relationships and competitive interactions (*low to medium confidence*). OA may stimulate global net primary production (*low confidence*) and nitrogen fixation (*medium confidence*). OA will increase the abundance and primary production of non-calcifying macrophytes, but will be harmful to calcifying algae and heterotrophs (*medium confidence*). Ecosystems relying on calcified structures and at risk of dissolution under OA include warm-water coral reefs (*high confidence*) and their cold-water equivalent (*medium confidence*). Further studies need to explore how OA may change the composition of communities, impact food webs and affect higher trophic levels.

6.3.3. *Life in Hypoxia and Anoxia*

6.3.3.1. *Principles*

Hypoxia constrains organisms which rely on aerobic metabolism (6.1.1, FAQ 6.2). Below O₂ concentrations of 60 μmoles kg⁻¹, commonly termed hypoxic (6.1.1.3), communities undergo species losses and replacements and are transformed into communities with species showing characteristic hypoxia adaptations. However, O₂ can limit animal life at even higher levels, just below air saturation (Gilly *et al.*, 2013). Organisms' tolerance thresholds have

been defined by either the critical O₂ partial pressure (P_c) or concentration (O₂crit). Thresholds vary across domains and are highest for large multicellular organisms. Among these, the P_c at rest varies depending on species, body size, and lifestage. In animals below the P_c aerobic metabolic rate fails to be maintained and anaerobic metabolism contributes to energy production (Pörtner and Grieshaber, 1993). The critical oxygen threshold is set by the capacity of ventilatory and circulatory systems to supply O₂ and cover demand. The threshold increases once metabolism is stimulated by muscular activity, temperature, or food uptake (Pörtner, 2002a; Ekau *et al.*, 2010; Seibel, 2011; Figure 6-11). At extreme temperatures, O₂crit approaches the oxygen content of air-saturated water (Pörtner, 2010; McBryan *et al.*, 2013), indicating high sensitivity to hypoxia in the warmth. Most animals can only sustain anaerobic metabolism temporarily, even if they are energy-efficient and survive long periods of anoxia (Grieshaber *et al.*, 1994). Such time-limited tolerance is higher in large than in small individuals or larvae, related to the higher capacity of anaerobic metabolism in large specimens (Gray *et al.*, 2002; Jessen *et al.*, 2009).

6.3.3.2. Microbes

Bacteria and protists consume ambient oxygen down to very low levels in ‘oxygen minimum zones’ and sustain OMZs by their metabolic diversity (Figure 6-11; WGI Box 6.5, Figure 1). OMZs form habitat for both anaerobic and aerobic microbes which can utilize very low (< 1 μmoles kg⁻¹) O₂ concentrations (Stolper *et al.*, 2010). Hypoxia is paralleled by elevated pCO₂ and enhanced acidification. Microbial life will benefit from expanding OMZs (*high confidence*).

6.3.3.3. Animals and Plants

In mesopelagic OMZs, zooplankton also contribute to the development of hypoxia (Robinson *et al.*, 2010, FAQ 6.2). During daytime zooplankton congregate at the upper margin of OMZs, where the degradation of organic material causes intensified respiration and oxygen depletion (Bianchi *et al.*, 2013). Animals living permanently in the OMZ cover virtually all energy demand by aerobic metabolism. This requires special adaptations leading to a reduction in O₂ and energy demand, and the improved ability to efficiently use available O₂. Enhanced hypoxia tolerance reflected in low O₂crit values is supported by small body size and by cold temperature (Vetter *et al.*, 1994; Pörtner, 2002b; Levin *et al.*, 2009). Accordingly, low O₂ levels support abundant meiofauna (very small fauna, <1mm) which benefit from abundant food and reduced predation by larger organisms (Levin, 2003). Under suboxia only specialists can survive (Vaquer-Sunyer and Duarte, 2008). Expansion of suboxic and anoxic centres of pelagic OMZs and benthic dead zones will lead to loss of habitat for animal life (*high confidence*).

Large, more active animals like fishes, crustaceans, and muscular (as opposed to ammoniacal) squids tend to have high O₂ demands associated with high O₂crit thresholds, and are therefore excluded from permanently hypoxic water bodies. However, even in high-activity animal groups some specialists such as Humboldt squid or bigeye tuna have adapted to enter hypoxic environments though only temporarily (Richards *et al.*, 2009; Seibel, 2011). The time-limited tolerance of animals to hypoxia below the O₂crit is maximized by the depression of energy demand, for example during periods of metabolic arrest (e.g. developmental arrest or diapause of copepods; Auel *et al.*, 2005). Hypoxia-adapted lifeforms will benefit from expanding OMZs (*high confidence*).

There is little information on the hypoxia sensitivity of macrophytes or their O₂crit values. In eelgrass (*Zostera marina*), warming causes the hypoxia threshold to rise due to a strong increase in tissue respiration. Concomitant water or sediment hypoxia can elicit tissue anoxia and sudden die-offs (Raun and Borum, 2013). By contrast, macroalgae attached to rocks rarely encounter anoxia (Raven and Scrimgeour, 1997). Expanding benthic OMZs will constrain the distribution of macrophytes (*medium confidence*).

6.3.3.4. Ecosystems

OMZs, shoaling and expanding vertically and laterally (Gilly *et al.*, 2013), will cause habitat and abundance losses for intolerant taxa such as mesopelagic (Koslow *et al.*, 2011) and epipelagic fishes with a high O₂ demand (*medium*

confidence, Prince *et al.*, 2010; Stramma *et al.*, 2012, FAQ 6.2). In line with the distribution of hypoxia sensitivities (Figure 6-11, 6.3.3.1, 6.3.3.3), expanding OMZs will further constrain the distribution of key zooplankton and nekton species and influence their diurnal and ontogenetic vertical migrations (*medium confidence*, Ekau *et al.*, 2010). The composition of microbial and faunal pelagic communities will shift from diverse mid-water assemblages to migrant biota that return to oxygenated surface waters at night (Seibel, 2011). Dissolved O₂, among other factors, plays an important role in shaping large alternating fluctuations of sardine and anchovy abundances, particularly off Peru. Anchovies are not strongly affected by a shallow oxycline (<10m), while sardines actively avoid such conditions (Bertrand *et al.*, 2010). Where OMZs intersect the continental shelves, groundfishes (McClatchie *et al.*, 2010) and large benthic invertebrates like crabs display high mortalities (Chan *et al.*, 2008). Susceptibility of early life stages to hypoxia in both pelagic and benthic ecosystems (Ekau *et al.*, 2010) threatens population survival. Effects of hypoxia propagate along the food chain, constraining fish stocks and top predators (Stramma *et al.*, 2010; *high confidence*). Hypoxia reduces biodiversity (Levin *et al.*, 2009; Gooday *et al.*, 2010) and causes the marginalization of calcifiers, due to low metabolic rates and high pCO₂ (Levin, 2003; Levin *et al.*, 2009; *high confidence*).

The expansion and enhanced variability of OMZs increases dissimilatory nitrate reduction and anaerobic ammonium oxidation (anammox), both releasing N₂ into the atmosphere, reducing the availability of fixed nitrogen and limiting oceanic primary productivity (*medium confidence*). Water column denitrification and N₂ fixation are spatially and temporally variable (*limited evidence, low confidence*), suggesting that climate effects on these processes are *unlikely* to operate uniformly (Brandes *et al.*, 2007; Fernandez *et al.*, 2011; Franz *et al.*, 2012).

If O₂ levels decline and OMZs expand, tolerant taxa, such as anaerobic bacteria (Ulloa *et al.*, 2012), gelatinous zooplankton (medusae, ctenophores), selected fishes (gobies, hake), and possibly selected cephalopods (Gilly *et al.*, 2006; Bazzino *et al.*, 2010), will respond with range expansions or population growth. Similar phenomena are expected with intensified upwelling causing extensive mortalities of coastal fishes and invertebrates (Box CC-UP). A community change toward hypoxia-tolerant fauna will occur in mid-water (*high confidence*). The diversity of macroorganisms will decrease and, finally, higher marine organisms will disappear and heterotrophic microorganisms will dominate (*high confidence*). In isolated water bodies like the Black Sea, warming will lead to the expansion of anoxia and H₂S poisoning, reduce pelagic and bottom faunal distributions, and shape trophic relations, energy flows, and productivity (Daskalov, 2003; Fashchuk, 2011).

[INSERT FIGURE 6-11 HERE

Figure 6-11: (A) Principal mechanisms underlying the formation of hypoxic conditions and their biological background (modified from Levin *et al.*, 2009; Levin and Sibuet, 2012). The buoyancy flux from fluvial discharges produces sharp density stratification at the base of the freshened layer (also valid for ice melt and high precipitation) near the surface and, hence, vertical mixing is greatly reduced. In consequence, the nutrient inputs from the river and the atmosphere accumulate in a narrow upper layer, leading to blooms of phytoplankton, possibly including harmful algae. The enhancement of oxygen consumption due to aerobic decomposition of sinking organic matter results in hypoxic conditions of benthic and midwater OMZs. Enrichment of nutrients (eutrophication) results in coastal dead zones. In the open oceans, heating of the upper layer increases stratification, while the wind-driven upwelling of hypoxic, nutrient-rich water from deeper layers adds to the formation of the OMZs (Box CC-UP) (B) Distribution of free-living marine organisms (microbes such as archaea, bacteria, protists, small and large multicellular animals, and plants) across the ranges of O₂ concentrations in various water layers. Hypoxia tolerance is enhanced in small compared to large organisms, allowing unicellular species and small animals to thrive in extremely hypoxic habitats. Species richness and body size of animals increases with rising O₂ levels.]

6.3.4. Mixed Layer Depth and Light Shaping Net Primary Production

The upper ocean is characterized by physical and chemical gradients in the surface mixed layer that influence the magnitude of photosynthetic carbon fixation, often termed Net Primary Production (NPP). The availability of light and nutrients to photoautotrophs sets daily rates of NPP and may be altered directly or indirectly, through changing mixed layer depths, shifts in the circulation regime at different spatial scales and the physical displacement of organisms (6.1.1.4, Box CC-PP, Figure 6-2). A changing climate will affect mixed layer depth, cloudiness, and/or

sea-ice areal extent and thickness and thereby, modulate NPP (*high confidence*). A stronger vertical density gradient will reduce the communication between the sunlit upper ocean where photosynthesis takes place and the underlying nutrient-rich waters (Figure 6-2). The supplies of plant nutrients (macro-nutrients) such as nitrate, and of micro-nutrients such as iron (Pitchford and Brindley, 1999) vary seasonally (Boyd, 2002) and regionally (Moore *et al.*, 2002), such that NPP may be simultaneously limited (co-limited) by more than one resource (6.3.5; Saito *et al.*, 2008).

The changing range and intensity of underwater light will lead to changes in NPP as well as in phytoplankton community composition (Doney, 2006; Boyd *et al.*, 2010). The response of phytoplankton to changing sunlight involves photo-physiological acclimation via changes in cellular chlorophyll, but such acclimation is constrained by unidentified limits (Falkowski and Raven, 1997). A longer growing season, with more sea-ice free days between 1998 and 2009, may have increased NPP in open Arctic waters (Arrigo and van Dijken, 2011; Box CC-PP), complemented by massive under-ice blooms as seen in 2011, favored by light that penetrates surface melt ponds and thinner, e.g., first-year ice (Arrigo *et al.*, 2012). There are also reports of increased incidences of high phytoplankton stocks, and hence of greater NPP, deeper in the water column (i.e., where it cannot be detected by satellite) during summer in the Arctic, which have implications to assessing changes in NPP from space (Hill *et al.*, 2013). Little is known about shifts from sea-ice algae to free-drifting phytoplankton expected with a decrease in sea-ice cover and effects of increased light in polar waters in the coming decades (*low confidence*). In the Arctic, summer ice melt led to a rapid export of sea-ice algae to the deep ocean (Boetius *et al.*, 2013). As some krill feed primarily on sea-ice algae, it is unclear (*low confidence*) whether they will adapt to feeding mainly on free-drifting phytoplankton (Smetacek and Nichol, 2005).

A range of time-series observations, from in situ phytoplankton abundances to satellite remotesensing, have been used to assess whether phytoplankton stocks and hence rates of NPP have altered over recent decades. Increases in phytoplankton stocks were found in regions where colder waters had warmed in the Northeast Atlantic, whereas the opposite trend was observed for warm water regions from a phytoplankton abundance time-series (Richardson and Schoeman, 2004). Lower chlorophyll concentrations at warmer SSTs in nutrient-poor low-latitude waters, based on satellite ocean color data, have been interpreted as an effect of increased stratification on phytoplankton stocks. It has thus been suggested that expanding, permanently stratified, low-chlorophyll, tropical regions (WGI, Ch. 3) indicate declining phytoplankton stocks in the warming oligotrophic waters of the N and S Pacific and N and S Atlantic (*limited evidence, low agreement* due to methodological uncertainties, Box CC-PP; Polovina *et al.*, 2008; Signorini and McClain, 2012; 30.5.1.1.2). Furthermore, a transition to conditions favoring increased frequency or even permanence of El Niño in a warmer future (Wara *et al.*, 2005) and further expansion of subtropical ocean gyres (Polovina *et al.*, 2008; 30.5.6), may lead to lower global ocean NPP (*low to medium confidence*).

However, these long-term ‘blended’ projections (i.e., constructing a biomass time-series using multiple proxies such as ocean transparency) of a global decrease in phytoplankton biomass (Boyce *et al.*, 2010) have been refuted (McQuatters-Gollop *et al.*, 2011; Mackas, 2011; Rykaczewski and Dunne, 2011). Time series shorter than 20 years do not resolve impacts of bidecadal variation such as the Pacific Decadal Oscillation or the lunar nodal cycle (e.g., Watanabe *et al.*, 2008; Henson *et al.*, 2010). Analysis of continental shelf ecosystems, including field data in the most productive upwelling areas covering the last 20 years (e.g. Chavez *et al.*, 2011), revealed a large variety of trends at scales of several decades but a general increase in NPP on most shelves (Sherman and Hempel, 2009; Bode *et al.*, 2011), possibly caused by natural climate variability, anthropogenic climate change, and/or anthropogenic eutrophication. Recent field measurements document an increasing rate of both anthropogenic fixed N (Duce *et al.*, 2008) and biological fixation of atmospheric nitrogen (Mouriño-Carballido *et al.*, 2011) entering the open ocean, which could lead to increased NPP especially in warm, stratified tropical and subtropical oceans provided sufficient phosphate and other growth requirements are present (*low confidence*, e.g., Sohm *et al.*, 2011).

For heterotrophs, from bacteria to fish, mammals and birds, the uptake of organic material as food, ultimately provided by NPP, is central not only to productivity but also for fueling energy consuming functions including the resistance of organisms to environmental change and pathogens (6.3.1-2). Any direct influence of climate on the abundance and quality of feed organisms will thus translate to indirect effects on the productivity and well-being of foraging animals (*high confidence*, Figure 6-5A, 6-7A, 6-12).

Overall, pelagic systems respond to climate change by region-specific changes in productivity with the projection of a small net reduction in global ocean NPP by 2100 (*medium confidence*, Box CC-PP). The spatial reorganization of NPP between latitudes affects higher trophic levels by alteration of the composition and functioning of pelagic communities (*medium confidence*).

6.3.5. Concurrent Responses to Multiple Drivers

Climate change alters oceanic properties globally, with concurrent changes in temperature, dissolved CO₂ and O₂, light, and nutrient concentrations (e.g., Sarmiento *et al.*, 1998; Matear and Hirst, 1999; Boyd and Doney, 2002; Ekau *et al.*, 2010; Figure 6-2). Additional direct human interventions at regional scale comprise the introduction of non-native species, overfishing, pollution, long-range atmospheric transport of nitrogen, point-source eutrophication, and habitat destruction (Carlton, 2000; Boyd and Hutchins, 2012). Worldwide alterations in marine ecosystems (Pauly *et al.*, 1998; Österblom *et al.*, 2007) have been linked to direct human activities, especially fishing (Frank *et al.*, 2005; deYoung *et al.*, 2008; Casini *et al.*, 2009), but may also be caused to some extent by climate variability and change (Cheung *et al.*, 2013a).

Alteration of each individual property has pronounced effects on organisms from microbes to animals, and hence on ecosystems (6.3.1-4). The cumulative effects of these factors will result in complex patterns of change, from organismal physiology to the areal extent and boundaries of biogeographic regions (Table 6-4). In many organisms, effects of ocean acidification interact with those of other key drivers like temperature and hypoxia (Pörtner, 2012; Boyd, 2011; Gruber, 2011) and translate from molecular to ecosystem level impacts. In phytoplankton, low light (Zondervan *et al.*, 2002) or nitrogen limitation (Sciandra *et al.*, 2003) limit beneficial OA effects on photosynthesis, and have a strong negative effect on plankton calcification (Rokitta and Rost, 2012). Nutrients and light support functional adjustments to OA through gene expression changes (Dyhrman *et al.*, 2006; Richier *et al.*, 2009).

Similar to today, paleo-events such as the Palaeocene-Eocene Boundary demonstrate concurrent warming, enhanced stratification of the oceans, deoxygenation of deeper waters, and OA, albeit at a rate more than 10 times slower than today's rate (6.1.2). Both the complexity of paleo-ecosystem changes and the complexity of present effects confound the clear attribution of biological trends to individual drivers (Parmesan *et al.*, 2011). For warming and hypoxia, changes are accelerated by effects of shifting seasonal or even diurnal extremes and their frequency on organisms and ecosystems (*medium evidence and agreement*) (e.g. Pörtner and Knust, 2007; Díaz and Rosenberg, 2008). This may also apply to effects of anthropogenic OA (*limited evidence and low agreement*).

6.3.5.1. Principles

Effects of various climate drivers on ocean ecosystems are intertwined and effects may be exacerbated by responses of biota. For example, warming reduces O₂ solubility and enhances biotic O₂ demand, which exacerbates hypoxia, produces CO₂ and causes acidification (Millero, 1995; Brewer and Peltzer, 2009). Drivers act with either additive, synergistic (i.e., amplification of) or antagonistic (i.e., diminution of) effects. A meta-analysis of 171 experimental studies that exposed marine systems to two or more drivers identified cumulative effects that were either additive (26%), synergistic (36%), or antagonistic (38%) (Crain *et al.*, 2008). Effects range from direct impacts of ocean warming on organismal physiology (Pörtner and Knust, 2007), to ocean acidification acting together with warming, e.g., on coccolithophore calcite production and abundances (Feng *et al.*, 2009), or with hypoxia and/or salinity changes (Table 6-4). Interactions of predominantly temperature, ocean acidification, and hypoxia have *likely* been involved in climate-driven evolutionary crises during Earth history (Pörtner *et al.*, 2005; 6.1.2).

[INSERT TABLE 6-4 HERE]

Table 6-4: Potential interactions between modes of anthropogenic forcing (environmental; foodwebs, harvesting) on different levels of biological organisation. These interactions, from simple to complex, are illustrated with examples from the literature. E, O, and M denote Experiments (lab or field), Observations, or Modeling approaches, respectively. Unknown denotes no published information is available for each of these categories. NA denotes Not Applicable for this category.]

Effects on individual organisms may also reflect intertwined impacts of ocean warming, acidification and hypoxia, which may operate through interrelated functional principles (Pörtner, 2012). Such knowledge helps to reconcile apparently contrasting findings. For example, warming toward the thermal optimum (Figure 6-5A) stimulates resistance to OA; CO₂-induced disturbances of growth and calcification were reversed by concomitant warming (Sheppard-Brennand *et al.*, 2010; Findlay *et al.*, 2010; Walther *et al.*, 2011). Warming to above optimum temperatures, however, constrains performance and exacerbates sensitivity to hypoxia (see below) and/or elevated CO₂ (Figure 6-5, e.g., via decreased calcification, Rodolfo-Metalpa *et al.*, 2011). Both hypoxia and/or elevated CO₂ in turn enhance heat sensitivity, as seen for CO₂ in crustaceans (via decreased heat limits, Walther *et al.*, 2009; Findlay *et al.*, 2010), coral reef fishes (via reduced performance, Munday *et al.*, 2009b) and corals (via decreased calcification and CO₂-enhanced bleaching; Reynaud *et al.*, 2003; Anthony *et al.*, 2008). This translates into a narrowing of the thermal niche (Walther *et al.*, 2009; Figure 6-5), which will shrink biogeographic ranges, affect species interactions and shift phenologies (Figure 6-7A). Hence, extreme warming and hypoxia exacerbate CO₂ effects and vice versa (*medium confidence*). Such principles need to be reconfirmed across organism taxa (Pörtner, 2012).

Differences in organism adaptation to a climate zone's characteristic temperatures, temperature variability, oxygen content, and ocean chemistry may shape vulnerability to climate change. In high polar species evolutionary cold adaptation enhances vulnerability to warming (*medium confidence*). In oxygen minimum zones, marine sediments, and in polar waters (due to high solubility in the cold), CO₂ levels are elevated and adaptation may reduce sensitivity and reliance on calcified structures (Clark *et al.*, 2009; Walther *et al.*, 2011; Maas *et al.*, 2012). The observed shift from 'overcalcified' to 'weakly calcified' coccolithophores *Emiliana huxleyi* in cold waters may reflect a related shift in ecotype dominance (Cubillos *et al.*, 2007, *limited evidence, medium agreement*).

Despite such potential adaptation, polar calcifiers exposed to higher CO₂ and lower carbonate saturation levels have been hypothesized to be highly sensitive to further CO₂ accumulation (*limited evidence, high agreement*; Orr *et al.*, 2005). Here it appears relevant that cold temperature reduces energy demand and thereby lowers resistance to ocean acidification. Both energy demand and resistance are higher in eurytherms than in high polar and deep sea stenotherms (Pörtner, 2006; e.g. crustaceans, Pane and Barry, 2007, cf. Whiteley, 2011; *limited evidence, medium agreement*). In turn, tropical species may be more sensitive than temperate zone species (Pörtner *et al.*, 2011). This rough differentiation of sensitivity is complicated by the local adaptation of populations from within-species genetic variability (*low confidence*).

Temperature influences hypoxia sensitivity (6.3.3). Warming causes the minimum tolerated O₂ level to rise, enhancing vulnerability (*high confidence*). Conversely, hypoxia enhances vulnerability to warming in animals. This may occur fastest in warm oceans, where metabolic rates are higher and animals live closer to upper thermal limits (*medium confidence*, Pörtner, 2010). However, evolutionary adaptation has led to high hypoxia tolerance (low P_c or O_{2,crit} values) in some warm-adapted coral reef fishes. Further warming then causes a rise in P_c which cannot be compensated for (Nilsson *et al.*, 2010). Limits to hypoxia adaptation coincide with upper thermal limits (*medium confidence*).

Complexity in responses rises with the number of drivers involved. Enhanced river runoff and increased precipitation cause a shift from marine to more brackish and even freshwater communities, with unclear consequences for effects of other drivers. Falling primary production reduces resilience of higher trophic levels (Kirby and Beaugrand, 2009; Stock *et al.*, 2011). The introduction of non-indigenous species, when supported by climate-induced shifts in interactions, may promote the displacement of ecotypes and shifts in ecosystem functioning, e.g., in the Mediterranean Sea (Occhipinti-Ambrogi, 2007; Coll *et al.*, 2010).

6.3.5.2. Microbes

Both synergistic and antagonistic effects of multiple drivers on microbial biota in the surface ocean have been observed in manipulation or modeling experiments (Boyd *et al.*, 2010; Folt *et al.*, 1999; Gruber, 2011). The productivity of many microbes was simultaneously limited by, for example, availability of nitrate and phosphate,

cobalt and iron (Saito *et al.*, 2002; Bertrand *et al.*, 2007), or iron and light (Boyd *et al.*, 2010; 6.2.2). Warming and high CO₂ synergistically enhanced photo-physiological rates of the cyanobacterium *Synechococcus*, whereas the cyanobacterial group *Prochlorococcus* showed no change (Fu *et al.*, 2007). The magnitude of CO₂ effects on growth, fixation rates or elemental ratios within single species is often strongly modulated by nutrient availability and light conditions (e.g., Zondervan *et al.*, 2002; Sciandra *et al.*, 2003; Kranz *et al.*, 2010). Such differences cause floristic shifts in phytoplankton with the potential to restructure predator-prey interactions (Table 6-4).

Co-limiting factors vary by group, such as nitrogen fixers (e.g., Hutchins *et al.*, 2007; Kranz *et al.*, 2010), diatoms (Boyd *et al.*, 2010), and coccolithophores (e.g., Feng *et al.*, 2009; Rokitta and Rost, 2012). This limits the ability to project climate change effects (Boyd *et al.*, 2010). The most reliable projections at ocean basin scale come from modeling, which mainly points to synergistic effects, such as those of elevated CO₂, hypoxia, and warming. For example, OA is projected to alter sinking particles (C:N ratio and/or reduced calcite content and slower sinking) with a consequent knock-on effect on water column O₂ demand already stimulated by warming, thereby causing expansion of OMZs (Gruber, 2011).

6.3.5.3. *Animals and Plants*

High oxygen availability alleviates thermal stress as seen in fish and mollusks (Mark *et al.*, 2002; Pörtner *et al.*, 2006). Conversely, hypoxia reduces heat tolerance (6.3.5.1), but acclimation to hypoxia compensates for this and increases thermal tolerance (Burlinson and Silva, 2011), for example by enhancing blood pigment content or reducing energy demand. Tolerances to hypoxia and to high temperature may be positively correlated in some fishes, indicating potential for adaptive evolution under climate change (*low confidence*; McBryan *et al.*, 2013).

As a consequence of hypoxia narrowing thermal ranges (6.3.5.1), combined warming and expanding hypoxia may cause mid-water mesopelagic and demersal fish stocks to decline at rates much quicker than anticipated in the California Current Ecosystem (Koslow *et al.*, 2011; McClatchie *et al.*, 2010). In benthic fauna, warming will also increase vulnerability to hypoxia. Experiments showed a rise in lethal oxygen concentrations by 25 % and thereby reducing survival by 36 % at 4 °C warmer temperatures (Vaquer-Sunyer and Duarte, 2011). Hence, warming is expected to expand the area of ecosystems affected by hypoxia even if oxygen concentrations remain unchanged (*high confidence*). Under combined hypoxia and warming, CO₂ can extend short-term passive tolerance (despite constraining long-term tolerance). It facilitates a reduction in energy demand (Reipschläger *et al.*, 1997; Pörtner *et al.*, 2000), thereby extending survival of transient extremes of temperatures or hypoxia (*medium confidence*).

In macroalgae (non-calcifying) light availability modulates the response to elevated *p*CO₂ and temperature levels (Russell *et al.*, 2011; Sarker *et al.*, 2013). In warm-water corals, warming acting synergistically with CO₂ reduces calcification and increases sensitivity to bleaching (*high confidence*; Anthony *et al.*, 2008). Combined warming and OA following B1 (~RCP4.5, reduced emission) and A1FI (~RCP8.5, business as usual) scenarios in mesocosms caused losses of symbionts and corals, and a nocturnal decalcification of the reef community in summer. Present day conditions already imply reduced resilience to episodic extreme events such as cyclones (Dove *et al.*, 2013, Box CC-CR).

6.3.5.4. *Ecosystems*

The cumulative impacts of climate change drivers underlie alterations of species interactions and ecosystem structure and functioning, including changes in trophodynamics and the physical and chemical characteristics of habitats (*high confidence*). These effects combine with more indirect effects, such as shifts in stratification and productivity, expanding oxygen minimum zones, and the changing composition and biomass of food (partly resulting from direct effects on prey organisms) (*high confidence*). These complexities reduce the precision and reliability of quantitative projections (6.5), including uncertainties concerning shifts in upwelling and their future role in global primary production and the development of fish stocks (Box CC-UP).

At the level of animal communities, effects of various drivers remain largely unexplored, some are highly complex. For example, the net eastward shift of Pacific skipjack tuna between 1980 and 2009 was linked to the shifting aggregation of macrozooplankton and micronekton, involving complex interactions of climate variability (due to ENSO, 30.5.2), warming ocean surface, shallowing mixed layer depth relative to the position of the warm pool, and the convergence of the pool with the Pacific Equatorial Divergence Province (30.5.6.1.1, Lehodey *et al.*, 2011). Interactive drivers will affect the relative performance of interacting species, thereby shifting species ranges, interactions and food webs (*medium confidence*, Figure 6-7A). Adaptation to various climate zones modifies the roles of light and temperature in seasonalities and species interactions (Bradshaw and Holzapfel, 2010). Moderate hypoxia expansion in warming seas, e.g. as the stratified central North Sea (Queste *et al.*, 2013) may well influence the degree of temperature-induced species displacements (Figure 6-7B).

Impacts of climate change on benthic ecosystem engineers can also profoundly alter ecosystems. Tropical corals respond to ocean warming and acidification by increased bleaching, impeded calcification rates, and increased incidence of disease (6.3.1-2; 30.5.6; Veron *et al.*, 2009; Veron, 2011, Box CC-CR, *high confidence*). In coral reefs under multiple stressors, differentiation of these large-scale phenomena into species-specific sensitivities is highly uncertain as trend data are virtually non-existent (Brainard *et al.*, 2011). Little is known about impacts on deep-water or cold-water corals and sponges, tropical calcified algae, bryozoans, sponges, and tube-forming serpulid worms (Wood, 1999). The reliance of all of these on surface productivity makes them vulnerable to any alteration in food supply. Projected severe stress from increased temperature, hypoxia and ocean acidification will cause reduced performance and increasing mortality in ecosystem engineers (*high confidence*), and a deterioration of habitat characteristics for other organisms (*medium to low confidence*).

As a corollary, shifts in the geographical distributions of marine species (e.g., to higher latitudes or deeper waters; Figures 6-7B, 6.5.2) cause changes in community composition and interactions (Simpson *et al.*, 2011; Harley, 2011; Hazen *et al.*, 2013). Some species may gain predominance and abundance from fitness benefits (Figure 6-7) while others become less competitive or easier prey (Occhipinti-Ambrogi, 2007). Thereby, climate change will reassemble communities and affect biodiversity, with differences over time and between biomes and latitudes (*high confidence*, Box CC-PP; 6.5, Sala and Knowlton, 2006; Cheung *et al.*, 2009; Parmesan and Matthews, 2005; Parmesan *et al.*, 2011).

6.3.6. Food Web Consequences

Community re-assembly under climate change involves a change in species composition and strongly alters food web structure, e.g., causing shifts in trophic pathways (Figure 6-12; Kirby and Beaugrand, 2009; Moloney *et al.*, 2011), some of which are irreversible (Jarre and Shannon, 2010). Through trophic cascades (Cury *et al.*, 2003; Luczak *et al.*, 2011), climate affects predation, competition, and food availability (e.g., via changes in NPP; Figure 6-12; Utne-Palm *et al.*, 2010), including fish resources (Parsons and Lear, 2001; Brown *et al.*, 2010). Trophic amplification then drives an ecosystem towards a new stable structure or regime, which may be difficult to reverse (Folke *et al.*, 2004). Warming may result in consumer control of food web structure because respiration of heterotrophic zooplankton and bacteria increases more strongly with warming than does photosynthesis of autotrophic phytoplankton (*medium confidence*, O'Connor *et al.*, 2009).

[INSERT FIGURE 6-12 HERE

Figure 6-12: Schematic diagram of expected responses to climate change in a marine food web. A coupled pelagic and benthic food web is structured by the body size spectrum of species. Combined warming, hypoxia and ocean acidification reduce body size, shift biogeographies, change species composition and abundance, and reconfigure trophic linkages and interaction dynamics. Fishing generally removes large-bodied species and truncates the body-size spectrum of the community. This confounds the detection and attribution of food web responses to climate change. Arrows represent species interactions (e.g., between predator and prey or competitors for food or space). Broken lines reflect the potential loss of populations and trophic linkages due to climate change.]

Many impacts of climate change on food webs resemble those caused by fishing, pollution, eutrophication, and associated hypoxia (6.3.3), and habitat change (Brander, 2007); unambiguous attribution to climate remains difficult

(*low to medium confidence*; Parmesan *et al.*, 2011). Some of these factors also affect food web responses to climate change. Fishing truncates the age and size structure of populations, making them more dependent on annual recruitment and reducing their ability to buffer environmental fluctuations (Genner *et al.*, 2010; Botsford *et al.*, 2011; Planque *et al.*, 2010, Figure 6-12). Both adult and larval fishes show greater variability in abundance in exploited compared to unexploited populations (Hsieh *et al.*, 2008). Warming, acidification, and removal of top or competing predators may all contribute to large fluctuations in gelatinous plankton (e.g., jellyfish) populations (*low confidence*, Molinero *et al.*, 2005; Richardson and Gibbons, 2008; Richardson *et al.*, 2009; Condon *et al.*, 2012).

Analyzing impacts on key species provides insight into how individual components of a food web will respond to perturbations. However, projections of future states must include the complex food web interactions that influence the species, and system-level responses, which affect stability and resilience of the overall ecosystem (Neutel *et al.*, 2007; Dunne and Williams, 2009; Romanuk *et al.*, 2009). There is no single approach currently available that includes the complex links within and among ecosystems, biogeochemistry, and climate as needed for projections of future states of marine food webs (Fulton, 2011; Moloney *et al.*, 2011). In conclusion, there is *low confidence* in the quantitative projections of such changes (for further discussion see 6.5).

6.3.7. Marine Reptiles, Mammals, and Birds

6.3.7.1. Principles

Marine reptiles (turtles, snakes, crocodiles), mammals, and seabirds breathe air but live mostly in water; some shift or expand their ranges as a result of climate warming. The body temperature of ectothermic reptiles is set by ambient conditions; only at large body size may their body store heat and temperature be higher than ambient. Reptiles are thus more responsive to temperature than homeothermic seabirds and marine mammals (McMahon and Hays, 2006), which regulate their body temperature by adjusting metabolic heat production and insulation from the environment, a trait beneficial especially in the cold. Various degrees of body core insulation in mammals and birds constrain their distribution to either warmer or colder waters (by poor or high insulation, respectively). However, large body sizes enable some aquatic air breathers to travel across the widest temperature ranges possible in some of the largest migrations on Earth.

Changes in water chemistry and hypoxia have minimal direct influences on the air-breathing vertebrates, reflecting their large independence from physical and chemical drivers in the oceans. There is evidence for increased sound propagation in a CO₂-enriched ocean, but no evidence yet for any effect on biota (Ilyina *et al.*, 2010). If habitat structures offering retreat or ambush disappear, this will increase the energetic costs of life. Warming waters increases the cost of pursuit-diving as prey fishes increase swimming velocity. The predation success of such mammals (e.g., sea lions) and seabirds (e.g., penguins, cormorants) is thus constrained to waters ≤ 20 °C (Cairns *et al.*, 2008), a trend that extrapolates into the future (*low to medium confidence*). As prey distributions shift, foragers tied to land between trips may be constrained by the physiological costs of finding prey (Hazen *et al.*, 2013; Péron *et al.*, 2012). If food items are only found in thermally restricted areas or move to greater depths, mammals and birds may become constrained to certain distribution ranges or to the physiological limits of their diving ability (McIntyre *et al.*, 2011). Conversely, hypoxic habitat compression for fishes may facilitate foraging opportunities for their air-breathing predators (Hazen *et al.*, 2009). Accordingly, many air-breathers encounter changing habitat and food availability with climate change (*high confidence*).

6.3.7.2. Field Observations

Some species of seabirds, marine mammals, and sea turtles have responded to the anomalous ocean climate of the 20th century (*high confidence*, Hughes, 2000). There is insufficient information to assess effects on sea snakes or crocodiles. Poleward distribution shifts of turtles consistent with recent warming have been recorded in almost all marine groups. Decadal-scale climate fluctuations affect their recruitment success and nesting abundance (Van Houtan and Halley, 2011), with an inverse correlation between warming and abundance in various species and regions (Balazs and Chaloupka, 2004; Chaloupka *et al.*, 2008; Mazaris *et al.*, 2009). Extreme weather causes nest

flooding, considerably reducing hatching success (Van Houtan and Bass, 2007), projected sea level rise (WGI, ch. 13) will exacerbate such impact. Those with high fidelity to nesting and foraging sites (Cuevas *et al.*, 2008) are impacted more than those capable of changing those sites (Fish *et al.*, 2009; Hawkes *et al.*, 2009). Continued warming, modulated by changing rainfall (Santidrián Tomillo *et al.*, 2012), may skew turtle sex ratios towards females, increase egg and hatchling mortality (Fuentes *et al.*, 2009), cause earlier onset of nesting (Pike *et al.*, 2006; Mazaris *et al.*, 2008), decrease nesting populations (Chaloupka *et al.*, 2008), and shift dietary breadths (Hawkes *et al.*, 2009), leading to projected recruitment declines (e.g., leatherback turtles; Saba *et al.*, 2012). Vulnerability due to shifting sex ratio alone remains unclear, as nesting beaches have persisted with low production of male hatchlings over decades or longer (*low confidence*, Broderick *et al.*, 2000; Godfrey *et al.*, 1999; Hays *et al.*, 2003). The absence of sea turtles in certain regions may be best explained by the temporal unavailability of food resources or strong thermoclines restricting their bottom foraging abilities (Braun-McNeill *et al.*, 2008; Gardner *et al.*, 2008).

Seabird range modifications probably caused by climate change were recorded in polar areas and the temperate zone of the North Atlantic (Grémillet and Boulinier, 2009). Temperate species have shifted their ranges to higher latitudes in both hemispheres (Robinson *et al.*, 2005; La Sorte and Jetz, 2010; Bunce *et al.*, 2002). Some species, like the king penguin, follow shifting foraging zones (Péron *et al.*, 2012); others, like the emperor penguin, are affected by changing habitat structure (sea ice, Jenouvrier *et al.*, 2012). Warming causes many bird species to breed earlier (Sydeman and Bograd, 2009). High-latitude, cool-water species undergo extended breeding seasons (Chambers *et al.*, 2011). There is often no agreement, whether changes reflect solely ocean warming, or a combination of factors, such as fishing pressure on seabirds' prey species, sea level rise, and pollution (Heath *et al.*, 2009; Galbraith *et al.*, 2005; Votier *et al.*, 2005). Most shifts in range and seasonal activity involve shifts in trophic relationships (*medium confidence*). Seabirds with narrow geographic domains are expected to be more susceptible to climate change (Chambers *et al.*, 2005; Grémillet and Boulinier, 2009), even leading to local extinctions (e.g., the Galápagos penguin, Vargas *et al.*, 2007; or the marbled murrelet, Becker *et al.*, 2007).

The distribution, phenology, and migratory timing of marine mammals is also shaped by predator-prey dynamics and climate impacts on specific habitats (Calambokidis *et al.*, 2009; Salvadeo *et al.*, 2011). Some marine mammals, i.e. dolphin, porpoise, and whale species shift their distribution poleward to follow the movement of their prey (*medium confidence*, Simmonds and Isaac, 2007; Salvadeo *et al.*, 2010; MacLeod *et al.*, 2005; Springer *et al.*, 1999). As in birds, vulnerability to climate change is high for marine mammals with narrow geographic ranges and high habitat dependence. For example, the critically endangered vaquita, endemic to the Northern Gulf of California, cannot move north because of the land barrier (MacLeod, 2009). The polar bear (Laidre *et al.*, 2008; Rode *et al.*, 2012) and the walrus depend on sea ice as a platform for hunting, resting, and giving birth. For polar bears, access to prey such as ringed seals has been disrupted by the later formation and earlier breakup of sea ice in the eastern Canadian Arctic. Seasonal migrants into the Arctic (fin, minke, gray, killer, humpback whales) may increasingly compete with species adapted to operate in habitat with sea ice (some seals, narwhal, bowhead whale, beluga). Both may benefit from the net loss of sea ice, which will offer them better access to foraging in a pelagic-dominated ecosystem (Moore and Huntington, 2008).

6.3.8. Summary and Conclusions

An organism's capacity to perform, but also its access to food energy fueling that performance, shape its sensitivity to climate change (*high confidence*). Extreme temperatures surpassing the fringes of the thermal envelope cause local abundance losses, extinction and shifts in temperature-dependent distribution ranges (6.3.1, *high confidence*).

Some climate change effects detected in the field can be attributed to temperature, but few allow clear attribution to other drivers (6.3.1-5, 6.6). In fishes and invertebrates, specialization in regional climate regimes co-defines sensitivity to warming, acidification and hypoxia (6.3.5, *high confidence*). In marine mammals, birds, and ectothermic reptiles changes in life history and population dynamics have often not been directly attributed to climate drivers (*low confidence*), but rather to the availability of habitat and food (6.3.7, *high confidence*).

Natural climatic variability (Figure 6-1) and anthropogenic change, with a strong role of warming, cause large-scale changes in biogeography, abundance, diversity, community composition and structure of marine species (6.3.1, *very*

high confidence). Warming reduces body size (6.3.1, *medium confidence*). Differential species responses modify their interactions across trophic levels through trophic amplification (6.3.6, *medium to high confidence*).

Some tropical species and ecosystems exist close to upper thermal limits placing them among the marine ecosystems most affected by climate change (6.3.1, *high confidence*). Corals and coral reefs are primary examples. However, other factors change concomitantly, such that quantifying the ecosystem change attributable to warming or other drivers has not always been possible (6.3.5).

Under future climate change ocean acidification will affect marine organisms and ecosystems for centuries (6.3.2, 6.3.5, *high confidence*). To date, very few ecosystem-level changes in the field have been attributed to anthropogenic or local ocean acidification (6.3.2, *medium confidence*). Concomitant trends of warming, O₂ depletion, OA, and other drivers prevent clear attribution to OA (6.3.5).

Elevated CO₂ levels stimulate primary production of some macroalgae and seagrass species (*high confidence*), causing them to be more competitive than calcifying organisms (6.3.2, *medium confidence*). High sensitivities to OA are associated with low capacities to maintain pH in internal fluids (*high confidence*). Calcification rates in sensitive invertebrates, including corals, echinoderms, and mollusks, decrease under OA, especially if combined with temperature extremes (6.3.5, *high confidence*). Thresholds beyond which effects occur can be quantified only with *low confidence*; there are differential sensitivities and thresholds between taxa and species (6.3.2, *high confidence*).

Expansion of OMZs leads to community shifts clearly attributable to extreme hypoxia (6.3.3, *high confidence*). Gradual effects of a progressive decline in ocean O₂ levels on communities have not been sufficiently explored.

In general, community re-assembly with new species coming in will occur in the transition to future climates (*medium confidence*) and lead to new ecosystem states (6.3.6, *low confidence*). Climate change interacts with top-down human interferences, like fisheries or other forms of harvesting, which accelerate impacts (*medium confidence*). Non-linearities challenge the projection of marine ecosystem trajectories (FAQ 6.4).

In microbes, a conceptual foundation suitable to support an integrated understanding of climate impacts on individual species and communities is lacking. Specific physiological responses, such as in primary production, N₂ fixation, or calcification, can be attributed to multiple environmental drivers associated with climate change (6.3.1-5, *high confidence*).

6.4. Human Activities in Marine Ecosystems: Adaptation Benefits and Threats

Human societies benefit from resources and processes supplied by marine ecosystems, so-called ecosystem services. Attributing and projecting ecosystem changes and their effects on human communities caused by climate change including ocean acidification (OA) is challenging. Insufficient observations compound an understanding of long-term changes and the definition of baseline conditions. Some of the challenges are related to the difficulty of projecting how human communities will adapt to changing marine ecosystem benefits.

6.4.1. Ecosystem Services

Marine ecosystem services (e.g., WGII, Ch. 5) include products (food, fuel, and biochemical resources), climate regulation and biogeochemical processes (CO₂ uptake, carbon storage, microbial water purification), coastal protection, provision of space and waterways for maritime transport, cultural services (recreational and spiritual opportunities, aesthetic enjoyment), and functions supporting all other ecosystem services (nutrient cycling, photosynthesis, and habitat creation). Most components of the marine environment contribute to more than one major category of ecosystem service: for example, ocean primary productivity is classified as a supporting service, but it affects provisioning services via changes in fisheries, generation of fossil fuel resources, regulating services via the global carbon cycle and climate regulation, and cultural services via the enjoyment of a healthy ecosystem. Rarely has economic damage of climate change to a whole ecosystem been evaluated and projected. The projected

loss of tropical reef cover due to ocean acidification under A1 and B2 SRES scenarios will cause damages of 870 and 528 billion USD (year 2000 value) by 2100, respectively (cost rising with parallel economic growth, Brander *et al.*, 2012; Box CC-OA). Such loss is felt most strongly in the respective regions.

6.4.1.1. Food from the Sea

Fisheries provide 3 billion people with almost 20 % of their average per capita intake of animal protein (FAO, 2012a), 400 million depend critically on fish for their food (Garcia and Rosenberg, 2010). Total world marine capture fisheries catches stabilized in the mid-1990s at about 90 million tons per year. Marine aquaculture of primarily mollusks and crustaceans contributes over 63 million tons annually to seafood production, mostly concentrated in coastal areas (FAO, 2012b). The growth of aquaculture has decelerated, but is still considered a development opportunity and a strong need in regions such as Africa and Latin America (WGII, Ch. 7.4.2.2).

Climate-induced shifts in ecosystems and fisheries production will create significant challenges to sustainability and management (7.5.1.1.3), particularly for countries with fewer resources and lower adaptive capacity, including many low-latitude and small island nations (*high confidence*, Allison *et al.*, 2009; Worm *et al.*, 2009; Cooley *et al.*, 2012; 7.2.1.2, 7.4.2.1, 30.6.2, WGIII, Ch. 2.1.4). Vulnerability will be exacerbated by increases in the frequency and severity of extreme events (e.g. floods or storms) damaging infrastructure, homes, health, livelihoods or non-marine food security (Kovats *et al.*, 2003; Rosegrant and Cline, 2003; Adger *et al.*, 2005; Haines *et al.*, 2006).

The projected trends in fish stocks will widen the disparity in food security between developing and developed nations. Fish migrations due to warming (6.3.1) have already shifted the composition of fisheries catches (Pinsky and Fogarty, 2012; Cheung *et al.*, 2013a) and altered stock distributions (Sabatés *et al.*, 2006). Further warming may be beneficial for fisheries productivity in some regions such as the North Atlantic, because of the poleward shift of exploited species and changes in primary productivity (Box 6-1, 30.5.1.1.1; Arnason, 2007; Stenevik and Sundby, 2007; Cheung *et al.*, 2010), or for some Pacific Islands due to the eastward redistribution of tuna stocks (Lehodey, 2000; Lehodey *et al.*, 2011). Resulting changes in accessibility and fishing operations costs are projected to straddle economic zones, perturb international fishery agreements and cause excessive exploitation (Hannesson, 2007; Sumaila *et al.*, 2011; 7.3.2.6; WGIII, Ch. 4.3.7).

Invertebrate fisheries and aquaculture appear very vulnerable to the impacts of ocean acidification (Barton *et al.*, 2012; Box CC-OA; Figure 6-10). This concerns especially shelled mollusks, with a substantial decline in their global production projected between 2020 and 2060 under the A2 business as usual scenario (Cooley and Doney, 2009; Cooley *et al.*, 2012). Effects on calcifying plankton will propagate through the food web, making estimates of economic impact on fish catch by OA difficult, also due to complex interactions with other stressors like warming and fisheries management (Branch *et al.*, 2013; Griffith *et al.*, 2012). Model projections suggest a potential loss of up to 13% (A1F1 scenario) to annual total fishery value in the US, or globally over 100 billion USD annually by 2100 (Cooley and Doney, 2009; Narita *et al.*, 2012). Vulnerability differs highly between nations according to the contribution of such fisheries to their economy (Cooley *et al.*, 2012; 7.3.2.6). These projections are sensitive to the projected vulnerabilities of the organisms to ocean acidification (6.3.2, *medium confidence*).

Fishing reduces abundances at high trophic levels, but increases abundances at mid trophic levels. It reduces species numbers, simplifies ecosystem structure, and increases ecosystem sensitivity to climate change (Perry *et al.*, 2010). Exploitation of fish stocks and the alteration of their demography, population dynamics, and life history traits (Planque *et al.*, 2010; Petitgas *et al.*, 2006; Perry *et al.*, 2010) can reduce the capacity of fish populations to buffer changes in climate variability (Ottersen *et al.*, 2006; Genner *et al.*, 2010), and increase variability in population size. Interactions between warming, OA, and human activities such as fishing may thus exacerbate climate impacts on a wide range of ocean processes and services, including marine fisheries (*medium confidence*, Table 6-4, 6-6; 30.6.2).

A 2°C global temperature increase by 2050 is estimated to cause global losses in landed value of USD 17-41 billion annually (in 2005 value), with an estimated cost of adaptation for the fisheries of \$7 - 30 billion annually over a 40-year timeframe between 2010 and 2050. The largest loss in landed value is projected to occur in East Asia and the Pacific (*low confidence*, Sumaila and Cheung, 2010). Overall impacts and the regional manifestations will partially

depend on the flexibility and response capacities of food production systems (Elmqvist *et al.*, 2003; Planque *et al.*, 2011a). Specific implications for the fishing industry are still poorly known, as future projections of shifts in primary production and knock-on effects through food webs and into fisheries remain uncertain (*low confidence* in effects of changing NPP; Planque *et al.*, 2011b; Stock *et al.*, 2011).

6.4.1.2. Other Provisioning Services

Reductions in marine biodiversity due to climate change and other anthropogenic stressors (Tittensor *et al.*, 2010), like OA (CBD, 2009) and pollution, might reduce the discovery of genetic resources from marine species useful in pharmaceutical, aquaculture, agriculture, and other industries (Arrieta *et al.*, 2010), leading to a loss of option value from marine ecosystems. Climate change increases the demand for marine renewable energy such as wind and wave power, though with potential ecosystem impacts of their infrastructure (6.4.2).

6.4.1.3. Climate Regulation and Extreme Events

The effect of climate change on marine biota will alter their contribution to climate regulation, i.e., the maintenance of the chemical composition and physical processes in the atmosphere and oceans (*high confidence*, Beaumont *et al.*, 2007). Regulatory mechanisms in which organisms (especially phytoplankton) play a key role, include control of the level of atmospheric CO₂ through the balance between photosynthesis and respiration (Johnson *et al.*, 2010), and through the biological and alkalinity pump (Feely *et al.*, 2008; Falkowski, 1997). They also include the modulation of further greenhouse gases such as nitrous oxide (N₂O; Jin and Gruber, 2003; Law, 2008; 6.1.1.3), and the modulation of other climatically reactive gases such as dimethylsulphide (DMS; Vogt *et al.*, 2008). A projected decrease in global ocean NPP (6.5.1) may result in decreased export of biogenic carbon to the deep ocean (Bopp *et al.*, 2002; Boyd and Doney, 2002; Hashioka and Yamanaka, 2007). A positive feedback on climate change may result, however, many of the factors controlling the pump are poorly understood (Figure 6-4; WGI, Ch. 6).

Coastal marine ecosystems reduce the effects of floods and storm surges which account for most of the natural disasters affecting people in coastal regions (IPCC, 2012a). Empirical and modeling studies show that coral reefs contribute to buffering the impact of tsunamis (Fernando *et al.*, 2005; Gravelle and Mimura, 2008; 5.4.2.4, 30.5; Box CC-CR). Experiments and models indicate that warming and OA slow coral growth by nearly 50% by 2050 (Box CC-CR, 5.4.2.4), making some islands and coastal areas more vulnerable to tsunamis, storm surges, wave energy and coastal erosion (*high confidence*). Wetlands and mangroves provide biologically diverse buffer zones (5.4.2.3). The combined impacts of climate change, pollution, deoxygenation and other overlapping stressors, on mangroves and wetlands have not been determined (Cooley *et al.*, 2009; Cooley, 2012). Some of these stressors enhance each other's effects in coastal systems (Cai *et al.*, 2011; Howarth *et al.*, 2011; Feely *et al.*, 2010).

6.4.1.4. Cultural Services

Cultural services encompass a wide array of services with marine biodiversity as a core component supporting recreation and tourism as the economically most relevant. Tropical coral reefs and their enormous biodiversity sustain substantial tourist industries, presently with global annual net benefits of about 9.6 billion USD (Cesar *et al.*, 2003; Box CC-CR; 30.6.2.2). If reef services degrade, coastal visitors might choose alternative attractions (UNWTO, 2008). Increased travel to see disappearing ecosystem types (e.g., Antarctica, Liggett *et al.*, 2011) or in previously inhospitable areas or seasons (Amelung *et al.*, 2007; Moore, 2010) create new pressures and are unsustainable as the locations of key attractors shift (e.g., cetaceans, Lambert *et al.*, 2010; Salvadeo *et al.*, 2013).

Climate change may endanger harvests of marine species with spiritual and aesthetic importance to indigenous cultures, raising ethical questions about cultural preservation (e.g., Nuttall, 1998). In coastal communities, losing the aesthetic values of marine ecosystems may harm local economies: better water quality and fewer harmful algal blooms are related to higher shellfish landings and real estate prices (Jin *et al.*, 2008).

Some heritage benefits of preserving marine ecosystems consist of the economic value of a healthy, diverse ecosystem to future generations. Any climate-related biodiversity loss or pollution of marine ecosystems would decrease the bank of resources for future opportunities. For example, the research and conservation value of coral reef biodiversity and its non-use value are estimated together at US\$ 5.5 billion annually (Cesar *et al.*, 2003). As with spiritual and aesthetic benefits, maintaining heritage benefits under climate change poses challenges for managers concerning equity and ethics as well as multigenerational (and possibly multi-cultural) ethical questions.

6.4.1.5. Supporting Services

Fully identifying the services supporting other ecosystem benefits is virtually impossible, as they are diverse in nature and scale. Ecosystem engineers play an important role in these services. Damage to calcifying algae and corals will reduce habitat for other species (6.3.5), biodiversity, cultural and leisure values and their climate regulation capacity.

Waterways for shipping are expected to change in the next several decades (*very high confidence*, 28, 30.6.2.3). Reductions in Arctic sea ice allow new trade routes such as the Northwest Passage (Wilson *et al.*, 2004; Granier *et al.*, 2006), enabling economically viable trans-Arctic shipping, and access to regional resources for exploitation and tourism. This development would increase emission of greenhouse gases and other pollutants (Lauer *et al.*, 2009; Corbett *et al.*, 2010), and facilitate the invasion of non-indigenous species carried on hulls and in ballast waters (Lewis *et al.*, 2004).

6.4.2. Management-Related Adaptations and Risks

6.4.2.1. Ecosystem Management

A changing climate will have both positive and negative consequences for managing ocean resources (*high confidence*) (6.4.1; Eide and Heen, 2002; Eide, 2007). Ecosystem-based management (EBM, an approach recognizing all, including human interactions within an ecosystem), or the ecosystem approach (EA, a strategy for the integrated management of living resources promoting both conservation and sustainable use), are increasingly adopted globally (FAO, 2003) to deal with the multitude of human pressures on marine ecosystems (Sherman *et al.*, 2005; Hoel, 2009). Extended EBM addresses changes driven by climate and human activities, considering that diverse drivers will interact and confound each other (6.3.5, Planque *et al.*, 2010; Eero *et al.*, 2011). Human activities will undermine resilience to other, including climate, impacts or undermine the effectiveness of mitigation and adaptation measures, by increasing variability (thereby reducing predictability), and limiting scope for adaptation (*high confidence*, e.g., Hughes, 2004; Eero *et al.*, 2011; Sissener and Bjørndal, 2005). Thus, managing ecosystems under climate change increases the resilience of ecosystems and adaptive capacity of management systems through reducing other human perturbations (e.g., overfishing) (Brander, 2008; 7.5.1.1.3). Managing ecosystems also reduces the consequences of ocean acidification until CO₂ emission reduction becomes effective (Box CC-OA; Rau *et al.*, 2012; Billé *et al.*, 2013; McLeod *et al.*, 2013). Ecosystem resilience is enhanced by reducing regional eutrophication (Falkenberg *et al.*, 2013), or in aquaculture by avoiding acidified water (Barton *et al.*, 2012) and by selecting and cultivating pre-adapted strains (Parker *et al.*, 2012).

However, effects of climate change cannot be reversed by reducing the impacts of non-climatic drivers, emphasizing the need for adaptive management. Increased variability of ecosystem responses to climate change and the low predictability of some biological responses undermine the effectiveness of management and conservation measures. A particular risk is that climate change may contribute to large-scale ecosystem regime shifts (6.3.1.5; Box 6-1). Detecting and forecasting such shifts from time-series of environmental and biological data (Carpenter and Brock, 2006; deYoung *et al.*, 2008), is constrained by an insufficient number of observations and limited quantitative understanding (6.1.2). Biogeographic shifts challenge spatial management (Box CC-MB; 6.3.1., 6.5), which is a fundamental part of EBM (Douvere, 2008), and demand that “fixed in law forever” site-attached zoning to protect specific species may need to become more flexible to maintain the original objectives as species move or community structures shift (*high confidence*, Soto, 2001; Hawkins, 2012).

6.4.2.2. Geoengineering Approaches

Geoengineering approaches to mitigate climate change and its effects, include Solar Radiation Management (SRM) and Carbon Dioxide Removal (CDR; see Table 6-5; IPCC, 2012b). SRM aims to reduce warming by increasing albedo, for example via stratospheric injection of sulphate aerosol (Crutzen, 2006). SRM may affect marine ecosystems through changes in precipitation. With continued CO₂ emissions it leaves ocean acidification largely unabated as it cannot mitigate rising atmospheric CO₂ concentrations (Vaughan and Lenton, 2011; Williamson and Turley, 2012). Termination of SRM after its implementation involves the risk of rapid climate change and more severe effects on ecosystems (Russell *et al.*, 2012).

Proposed CDR techniques include both ocean- and land-based approaches (Vaughan and Lenton, 2011; 30.6.4). CO₂ storage in geological reservoirs may occur beneath the seafloor, e.g., in porous marine aquifers, and includes the risk of CO₂ leakage to the marine environment. Proposals to directly or indirectly sequester CO₂ into the ocean (Table 6-5; Caldeira *et al.*, 2005; Boyd, 2008; Shepherd *et al.*, 2009; WGIII, Ch. 7.5.5) include, among others, the use of ocean fertilization techniques by nutrient addition, the direct storage of biomass in the deep ocean, the addition of alkalinity for build-up of DIC (dissolved inorganic carbon, i.e. carbonate), and the direct CO₂ injection into the deep ocean (Williamson *et al.*, 2012). All of these approaches have potentially negative consequences for marine ecosystems.

[INSERT TABLE 6-5 HERE

Table 6-5: Challenges for the oceans that will arise from the employment of a range of geoengineering methods (SRM, solar radiation management, CDR, carbon dioxide removal).]

Ocean fertilization by adding iron to high-nutrient low-chlorophyll (HNLC) oceanic waters could increase productivity and the net export of organic material to the deep ocean and its consecutive decomposition, causing deep-water accumulation of CO₂. Fertilization would affect all major marine biogeochemical cycles of the ocean with unclear side effects that could include the formation of methane (CH₄) and N₂O (Law, 2008) or the stimulation of harmful algal blooms (Trick *et al.*, 2010). The enhanced NPP would add more carbon to the base of food webs (de Baar *et al.*, 2005) and stimulate growth, e.g., of deep-sea benthos (Wolff *et al.*, 2011). Any regional increase in organic material (through fertilization or intentional storage of biomass) would cause enhanced O₂ demand and deep-water O₂ depletion (Sarmiento *et al.*, 2010; Table 6-5), increasing the level and extent of hypoxia and associated impacts on marine ecosystems (6.3.3., 6.3.5, 30.5.7). The synergistic effects CO₂-induced acidification will exacerbate the biological impacts (*high confidence*).

Neutralizing the acidifying water by the addition of alkalinity, e.g., calcium oxide, would require large-scale terrestrial mining with associated consequences (Caldeira *et al.*, 2005). The biological effects of increased concentrations of Ca²⁺ ions and dissolved inorganic carbon remain insufficiently explored. Direct injection of CO₂ or its localized disposal in the ocean (e.g. as a lake in a deep-sea valley) cause locally highly increased CO₂ and acidification effects on deep-sea organisms (6.3.3.4., *high confidence*, Caldeira *et al.*, 2005). In contrast to long-term ocean fertilization or storage of biomass, this technique leaves the oxygen inventory of the deep ocean untouched (*limited evidence, medium agreement*, Pörtner *et al.*, 2005).

The knowledge base on the implementation of SRM and CDR techniques and associated risks is presently insufficient. Comparative assessments suggest that the main ocean-related geoengineering approaches are very costly and have large environmental footprints (*high confidence*, Boyd, 2008; Vaughan and Lenton, 2011; Russell *et al.*, 2012).

6.4.2.3. Health Issues

Human health and near-shore ecosystems may be directly impacted by climate change effects on harmful algal blooms (HABs, Edwards *et al.*, 2006; 30.6.3) or disease vectors. Planktonic time-series archives and nearshore

sediment cores containing HAB cysts have revealed few examples of strong linkages between altered HABs and climate fluctuations (Dale *et al.*, 2006; 30.5.3.1.2). HABs can be stimulated by warming, nutrient fluctuations in upwelling areas, eutrophication in coastal areas, and enhanced surface stratification (*medium confidence*). Species-specific responses involve shifts in seasonal cycles and blooms (Johns *et al.*, 2003). Ocean acidification may exacerbate the toxicity of species in coastal oceans under nutrient-limited conditions (Tatters *et al.*, 2012; Sun *et al.*, 2011). Suitable adaptation measures include appropriate monitoring of biotoxin problems (Hallegraeff, 2010).

Continued warming of tropical and temperate coastal habitats, excessive nutrient loading leading to phytoplankton and zooplankton blooms, and sea water inundation due to sea level rise are all projected to exacerbate the expansion and threat of cholera (*medium confidence*, 11.5.2.1; 30.6.3), although attribution to climate change is confounded by climate variability and non-climate drivers (Lafferty, 2009; Dobson, 2009). Cholera and its pathogen, the marine bacterium, *Vibrio cholera*, have been widely studied. The pathogen associates with marine organisms, especially chitinized zooplankton (Vezzulli *et al.*, 2010). Where cholera is endemic (e.g. India, Bangladesh, Latin America), outbreaks are correlated with warming and high zooplankton abundance (Lobitz *et al.*, 2000; Lipp *et al.*, 2002). Based on an 18-year climate record for Bangladesh, Pascual *et al.* (2000) reported cholera outbreaks at ENSO events, and the recent reappearance of cholera in Peru has also been linked to the intense 1991-1992 ENSO (Lipp *et al.*, 2002). An increase in sustained maximum temperatures of the Baltic Sea (30.5.3.1.4) has been related to an increase in reported *Vibrio* infections; highest human mortality rates were associated with *V. vulnificus* infections (Baker-Austin *et al.*, 2013). Continued warming of tropical and temperate coastal habitats, excessive nutrient loading leading to phytoplankton and zooplankton blooms, and seawater inundation due to sea level rise are all projected to exacerbate the expansion and threat of cholera (*medium confidence*).

Ciguatera poisoning may occur when people consume fish, mainly from tropical reefs, that have ciguatoxins from the epiphytic dinoflagellate *Gambierdiscus* sp. Historical records show significant correlations between ciguatera poisoning and sea surface temperature in South Pacific nations (Hales *et al.*, 1999). However, the relationship is non-linear and dependent on the thermal window of the specific dinoflagellate (Llewellyn, 2010). This casts doubt on the accuracy of projected increases in ciguatera poisoning using linear extrapolations from observations (*low confidence*).

6.4.3. Conclusions

Human societies benefit from and depend on marine ecosystem services, including the provisioning of food and other goods, regulation of climate and extreme events, and cultural and supporting services (6.4.1). Attributing and projecting climate-change-mediated shifts in these services remains a challenge, due to the intrinsic difficulty of assessments, lack of baseline and long time-series data and confounding human impacts. However, empirical and modeling studies indicate that climate change impacts on marine ecosystems lead to changes in provisioning, regulating and supporting services (*high confidence*), as well as cultural services (*limited evidence, medium agreement*).

Food production from the sea is facing diverse stressors (6.4.1.1), such as overfishing and habitat degradation, which interact with climate change phenomena, including warming (6.3.1), ocean acidification (6.3.2), and hypoxia (6.3.3). Projections of impacts on capture fisheries are constrained by uncertainties in marine primary production (*medium evidence and agreement*, 6.5.1). Negative effects are projected to be most significant in developing nations in tropical regions (*high confidence*). Nations at higher latitudes may even benefit from climate change effects on ocean ecosystems, at least initially (6.5.3).

Climate change effects on biota will alter their climate regulation through mechanisms such as carbonate production, the biological pump, the balance between photosynthesis and respiration, and the modulation of greenhouse gases (6.4.1.3, *high confidence*). However, projections of the direction and magnitudes of feedbacks are at an early stage (*low confidence*).

Future management of ecosystems and fisheries might have to aim for increasing ecosystem resilience to climate change, e.g., through reductions of other human perturbations (6.4.2.1). Active ocean geoengineering strategies to

ameliorate climate change may prove detrimental to the functioning of ecosystems, which highlights the need for further research and careful governance (6.4.2.2). There is limited understanding of how harmful algal blooms and pathogens affecting human health will respond to climate change (6.4.2.3, *medium to low confidence*).

6.5. Projections of Future Climate Change Impacts through Modeling Approaches

A range of models explore climate change effects on marine biota, from primary producers to higher trophic levels, and test hypotheses about responses of marine species, food webs and ecosystems (Rose *et al.*, 2010; Stock *et al.*, 2011; Fulton *et al.*, 2011, FAQ 6.2). Both empirical and mechanistic approaches are used over a range of temporal and spatial scales (Barange *et al.*, 2010; Stock *et al.*, 2011). There is an increasing need for upscaling from molecular and physiological to ecosystem level (e.g., Le Quesne and Pinnegar, 2012). Uncertainty in projections of changes in marine ecosystems is partly contingent on the level of confidence in climatic and oceanographic projections (6.1.1; WGI, Ch. 9.8). Models are currently useful for developing scenarios of directional changes in net primary productivity, species distributions, community structure and trophic dynamics of marine ecosystems, as well as their implications for ecosystem goods and services under climate change. However, specific quantitative projections by these models remain imprecise (*low confidence*, Rose *et al.*, 2010; Hannah *et al.*, 2010; Stock *et al.*, 2011; FAQ 6.4).

Earth System Models couple atmosphere, cryosphere, and hydrosphere (including the oceans), as well as climate and carbon cycles and project changes in ocean biogeochemistry under a range of CO₂ emission scenarios (WGI, Ch. 6). Models focusing on population and species level responses comprise models of population dynamics, models of species distribution, and models which explicitly link effects of changes in ocean physics and chemistry to changes in interactions between species at different trophic levels, or human activities such as fishing and aquaculture (Rose *et al.*, 2010).

6.5.1. Oceanic Primary Production

Climate induced effects on global ocean NPP comprise changes in its long-term average, seasonal timing, and peak amplitude (Henson *et al.*, 2013). The magnitude, direction, and pattern of projected changes vary with differences in model structure and parameterization (Box CC-PP; Figure 6-13). Unknown accuracy of current NPP observations further increases the uncertainty of projections, as does the incomplete understanding of effects of multiple drivers on NPP (6.3.1-5, 6.4). Global coupled climate-ocean biogeochemical Earth System Models (WGI Ch. 6) project an increase in NPP at high latitudes but a decrease in permanently stratified oceans at mid-latitudes, in the tropics (west tropical Pacific, tropical Indian Ocean, tropical Atlantic) and in the North Atlantic (Steinacher *et al.*, 2010; Bopp *et al.*, 2013, *medium confidence*) (Figure 6-13). The overall result is a reduction in global mean NPP under all RCP scenarios (*medium confidence* in the direction of projected trends, *low confidence* in the magnitude of change).

[INSERT FIGURE 6-13 HERE

Figure 6-13: Multi-model annual mean changes of projected vertically-integrated net primary production (small and large phytoplankton) under the low emission scenario RCP2.6 (A) and the high emission scenario RCP8.5 (B) for the period 2090 to 2099 relative to 1990 to 1999 (after Bopp *et al.*, 2013). To indicate consistency in the sign of change, regions are stippled where 80% of the ten models from the Coupled Model Intercomparison Project Phase 5 (Bopp *et al.* 2013) agree on the sign of change.]

6.5.2. Higher Trophic Levels

Projected future changes in temperature and other physical and chemical oceanographic factors are expected to affect the distribution and abundance of marine fishes and invertebrates, as elaborated by species distribution models. Limits of distribution ranges of 1066 exploited species are projected to undergo shifts by a median of around 50 km per decade to higher latitudes by 2050 relative to 2000 under the SRES A1B (RCP6.0) scenario (Cheung *et al.*, 2009). Some species shift towards the equator following a regional temperature gradient (Burrows *et al.*, 2011;

Cheung *et al.*, 2013b; Pinsky *et al.*, 2013). The rate of range shifts is projected to be three times higher for pelagic than for demersal fishes (Cheung *et al.*, 2009), the latter shifting at a rate of around 27 – 36 km per decade (Cheung *et al.*, 2013b). However, the expansion of hypoxic waters may have a greater impact than warming on demersal fishes (Koslow *et al.*, 2011). As a result of distribution shifts, high-latitude regions (the Arctic, Southern Ocean) are projected to have high rates of species invasions. Intermediate latitudes are expected to undergo both invasions and local extinctions. High rates of local extinction are projected for the tropics and semi-enclosed seas (e.g. Mediterranean Sea, Persian Gulf). In addition, the future productivity and distribution of higher trophic level organisms are projected to change due to changes in primary productivity (6.3.6). For example, the migration route of Pacific sardine is projected to shift because of changes in primary productivity and food availability (Ito *et al.*, 2010). The global pattern of distribution shifts is generally consistent with regional-scale projections and past observations (e.g., Lenoir *et al.*, 2011; Cheung *et al.*, 2013a). However, detailed quantitative projections are sensitive to model structure and assumptions (Hare *et al.*, 2012; Jones *et al.*, 2013) and responses of specific populations may differ from average species responses (Hazen *et al.*, 2013).

[INSERT FIGURE 6-14 HERE

Figure 6-14: Climate change effects on the biogeography, body size and fisheries catch potential of marine fishes and invertebrates. (A) Shifts in distribution range and reduction in body size of exploited fish driven by projected warming, oxygen depletion and sea-ice retreat (cf. Figure 6-7). Whenever the shift in distribution does not fully compensate for warming and hypoxia, the result will be a decrease in body size. Shifts in (B) latitudinal and (C) depth distribution of 610 exploited demersal fishes are projected to have a median (central line of the box) of 31 km decade⁻¹ and 3.3 m decade⁻¹, respectively, with variation between species (box boundary: 25th and 75th percentiles) from 1991-2010 to 2041 - 2060 under the SRES A2 (between RCP6.0 - 8.5) scenario (Cheung *et al.*, 2011; Cheung *et al.*, 2013b). (D) Combining species' range shift with projected changes in NPP leads to a projected global redistribution of maximum catch potential (Analysis includes ~1,000 species of exploited fishes and invertebrates, under warming by 2°C according to SRES A1B (~RCP6.0), comparing the 10-year averages 2001-2010 and 2051-2060; redrawn from Cheung *et al.* 2010). (E) Changes in species distribution and individual growth are projected to lead to reduced maximum body size of fish communities at a certain site. The analysis includes 610 species of marine fishes, from 1991-2010 to 2041-2060 under the SRES A2 (~RCP6.0 to 8.5, Cheung *et al.*, 2013b). Key assumptions of the projections are that current distribution ranges reflect the preferences and tolerances of species for temperature and other environmental conditions and that these preferences and tolerances do not change over time. Catch potential is determined by species range and net primary production. Growth and maximum body size of fishes are a function of temperature and ambient oxygen level.]

Coral reefs are projected to undergo long-term degradation by 2020 to 2100 relative to the 2000s under RCP 2.6, 4.5 and 8.5 or their equivalent (30.5.6). Reefs projected to be threatened most by bleaching under the SRES A1B scenario by 2100 include the Central and Western Equatorial Pacific, Coral Triangle and parts of Micronesia and Melanesia (Teneva *et al.*, 2012). These projections assume that coral bleaching occurs when SST exceeds a certain threshold, and that there is limited potential to shift such threshold by adaptation. Reef degradation will impact ecosystem services (Hoegh-Guldberg, 2011; 6.4; Box CC-CR).

Some groups of marine air-breathing fauna are projected to shift in distribution and abundance (6.3.7). Cetacean richness will increase above 40° latitude in both hemispheres, while at lower latitudes both pinniped and cetacean richness are projected to decrease by 2040-2049 relative to 1990-1999 under the SRES A1B scenario (Kaschner *et al.*, 2011). Using SST as a predictor, the distribution of loggerhead turtles is projected to expand poleward in the Atlantic Ocean and to gain habitat in the Mediterranean Sea by 2070-2089 relative to 1970-1989 (Witt *et al.*, 2010). Leatherback turtle may decrease in abundance at a rate of 7% per decade because of reduced hatching success with warming following the SRES A2 scenario (Saba *et al.*, 2012). Abundances of some seabirds such as European breeding seabirds (Huntley *et al.*, 2007), Cassin's auklet in the California Current ecosystem or Emperor penguin in Antarctica, are projected to decline because of climate induced changes in oceanographic conditions, such as temperature and upwelling intensity (Box CC-UP; Wolf *et al.*, 2010), or summer sea ice conditions (Jenouvrier *et al.*, 2012). The diversity of megafaunal responses to climate change will have cascading ecosystem impacts, and will affect ecosystem services such as tourism (6.3.7, 6.4.1, *high confidence*).

6.5.3. Ecosystems and Fisheries

One of the most direct impacts of climate change on marine ecosystem services is through fisheries (6.4.1; 7.2.1.2, 7.3.2.4, 7.4.2). Projected climate impacts on fisheries are based on recruitment, growth, mortality, abundance and distribution of fish stocks as well as changes in ocean NPP (Cheung *et al.*, 2008), evaluated from chlorophyll concentration and other variables such as sea surface temperature (Campbell *et al.*, 2002). Friedland *et al.* (2012) suggested that chlorophyll concentration, indicating both phytoplankton production and biomass, is a better predictor of the fishery yield in large marine ecosystems than NPP. While the principle holds that catch potential is dependent on energy from primary production, quantitative projections of catch potential are limited by residual uncertainty on the best possible indicators of primary production and biomass.

Assuming that the potential fish catch is proportional to NPP, the fish catch in the North Pacific Ocean subtropical biome is projected to increase by 26% through expansion of the biome, while catches in the temperate and equatorial biomes may decrease by 38 and 15%, respectively, through contraction of the biomes by 2100 relative to 2000 under the SRES A2 (RCP6.0 to 8.5) scenario (Polovina *et al.*, 2011). Changes in phytoplankton size structure are projected to affect fisheries catch potential (Cheung *et al.*, 2011), resulting in a 0 up to 75.8% decrease in the potential catch of large fishes in the central North Pacific and increases of up to 43% in the California Current region over the 21st century under the SRES A2 scenario (Woodworth-Jefcoats *et al.*, 2013). Globally, climate change is projected to cause a large-scale redistribution of global catch potential, with an average 30 - 70 % increase in yield at high latitudes and up to 89% in some regions, after 2°C warming from pre-industrial periods following SRES A1B (RCP6.0) (Figure 6-14; Cheung *et al.*, 2010; Blanchard *et al.*, 2012). Redistribution between areas, with average catch potential remaining unchanged, will occur at mid latitudes. A 40 - 60% drop will occur in the tropics and in Antarctica by the 2050s relative to the 2000s (*medium confidence* for direction of trends in fisheries yields, *low confidence* for the magnitude of change). This highlights high vulnerabilities in the economies of tropical coastal countries (Allison *et al.*, 2009; 6.4).

Fisheries targeting specific species may show more complex responses to climate change. For example, driven by changes in temperature and primary production, catches of skipjack and bigeye tuna in the south Pacific are projected to increase by 2035 relative to 1980-2000 under the SRES B1 and A2 scenario, but for 2100, skipjack tuna catch is projected to decrease under the A2 scenario, while bigeye tuna catch decreases under both A2 and B1 scenarios (Lehodey *et al.*, 2011). Regionally, tuna catches in the western Pacific are projected to decrease, while those in the eastern Pacific will increase (Lehodey *et al.*, 2011). Mollusk fisheries under ocean acidification is discussed under 6.4.1.

Identifying responses to climate change is complicated by species interactions and multiple stressors. Major marine habitats and biodiversity hotspots are projected to encounter cumulative impact from changes in temperature, pH, oxygen and primary productivity by the end of the 21st century (RCP4.5 and 8.5) (Mora *et al.* 2013). Acidification and hypoxia will reduce maximum catch potential over 50 years from about 2000 onward in both the North Atlantic and Northeast Pacific (Ainsworth *et al.*, 2011; Cheung *et al.*, 2011). Changes in O₂ content as well as warming will drive a global decrease of community-averaged maximum body size of 14-24% of exploited demersal marine fishes by 2050 relative to the 2000s under the SRES A2 (RCP6.0 to 8.5) scenario (Cheung *et al.*, 2013b, Figure 6-14). The decrease in maximum body size may affect natural mortality rates, trophic interactions, and reduce yield-per-recruit and, thus, potential catch. Responses of exploited marine species and their fisheries may interact with other human stressors such as overfishing, exacerbating their impacts (e.g., Lindegren *et al.*, 2010; Ainsworth *et al.*, 2011). Through species shifts climate change may also cause overlap of habitats of species targeted by fishing with habitat of threatened species, potentially increasing the chances of the latter of being caught as bycatch (Jones *et al.* 2013). Moreover, differences in vulnerability and adaptive capacity of species to changing environmental and ecosystem conditions will affect the responses of fisheries to climate change (e.g., Griffith *et al.*, 2011; Le Borgne *et al.*, 2011).

The complex and non-linear interactions and responses of both biophysical and socio-economic systems to climate change may lead to changes that have a low probability of occurrence based on empirical data (Doak *et al.*, 2008). The risk of such low probability but potentially high impact events may be under-estimated in existing model projections (Williams and Jackson, 2007; Lindenmayer *et al.*, 2010). Projected changes in the distribution and production potential of fisheries resources are expected to affect economics, human livelihood and food security

(Allison *et al.*, 2009; Sumaila and Cheung, 2010; *low confidence* in the magnitude and direction of the projected socio-economic impacts).

6.5.4. Conclusions

Modeling projects that the distribution of invertebrates, fishes, and some marine mammals, birds and reptiles will shift further under most emission scenarios, with rates and directions of shifts consistent with those observed in the last century (*high confidence*, 6.3.1-7). These projections are valid for those species that adapt not at all or incompletely to warmer temperatures and the associated ecosystem changes, as indicated by present trends (6.3.1, Box CC-MB). For non-adapting species rates of shift will thus increase with increasing rates of warming and higher emission scenarios (*high confidence*), unless the shift is blocked by geographic or other barriers (e.g., light regime, Figure 6-7). The average shift in distribution will continue to be poleward at large spatial scales (6.5.2, Box CC-MB, *high confidence*). Species richness and the abundance of warm-water species will increase at high latitudes (*high confidence*) and decrease in the tropics (6.5.2, *medium confidence*). Projections for individual species and populations are more variable and sensitive to model parameters.

Maximum fisheries catch potential is projected to increase at high and decrease at low latitudes by 2050 under SRES B1 (RCP4.5) and A1B (RCP6.0) climate scenarios (6.5.3, *medium confidence*). Quantifying such projections is constrained by uncertainties in projected primary production rates (6.3.4, 6.5.1), biological responses such as species interactions (6.3.6), and in projected effects of multiple climate drivers and human activities (6.3.5, *low confidence*).

Models that integrate climate and ocean changes with biological responses and interactions, and with current human activities, have led to agreement on species and food web responses to climate change (6.5.3). However, most of these models do not include trophic interactions. They insufficiently consider physiological principles and none include evolutionary adaptation that affect responses of biota to physical and chemical changes.

Projections of ocean biogeochemistry represent the open oceans rather well, but coastal and shelf regions only poorly. From a global perspective, open ocean NPP will decrease moderately by 2100 under both low (SRES B1 or RCP4.5) and high emission scenarios (A2 or RCP6.0 - 8.5; 6.3.4, 6.5.1, *medium confidence*), paralleled by an increase in NPP at high latitudes and a decrease in the tropics (6.3.4, 6.5.1, Box CC-PP, *medium confidence*).

Overall, the projected responses of marine organisms and ecosystems to climate change include changes in primary productivity (*medium confidence*), species' life history (*medium confidence*), distribution, abundance and diversity across marine food webs (*high confidence*) in a time frame of 20 to 80 years from 2010, with substantially larger long-term (end of 21st century) responses under high emission scenarios (*high confidence*). These changes will be largest under business as usual scenarios (RCP8.5) and increase the vulnerability of human societies, by affecting income, employment and food security through their effects on fisheries, tourism, and regulatory services such as coastal protection (6.4.1.3, Box CC-CR, *medium confidence*).

6.6. Chapter Conclusions and Key Uncertainties

This section provides an overview of confidence levels in the detection and projection of climate change effects on ocean systems, and of confidence levels in their attribution to different forcings. It distinguishes between effects previously observed and those projected, and considers confidence in the knowledge of underlying principles as discussed in this chapter. While the anthropogenic signal is conspicuous in the oceans (6.1.1), clear attribution to anthropogenic influences on climate is not always possible in individual case studies, due to the inherent variability of the system (Figure 6-15. Acronyms of relevant processes, capitalized, link between text and figure).

Present day observations and those from the Geological Record (GR, Figure 6-15) show similar signs of response to environmental changes, e.g., warming at high CO₂ levels, and similar ecological consequences in the ocean (*robust evidence, medium agreement and confidence*). However, the ongoing rate of anthropogenic CO₂ release and hence ocean acidification is unprecedented in the last 65 Ma (*high confidence*) and probably the last 300 Ma (6.1.2).

[INSERT FIGURE 6-15 HERE]

Figure 6-15: Overview of the levels of confidence in detection (left), as well as in projection (right) of climate change effects on ocean systems, in relation to the levels of confidence in attributing these effects to the respective climate forcings. Case studies, processes, and concepts relevant in assessing the effects of climate change are represented by their acronyms in both text and figure. While confidence in the presence of effects is often high, the direct attribution to one driver in field experiments is difficult, as drivers are often highly correlated with each other (e.g., warming with changes in stratification and hence reduced nutrient supply). Some climate change impacts have been condensed into broad categories to avoid overpopulating the figures (e.g., Bio-Geochemical processes, BG). Note that the term “attribution” is used for both present-day detections in the field and future projections, the latter including qualitative and quantitative extrapolations and simulations from fundamental principles and models. Firm knowledge from experiments (field, laboratory and modeling) simulating future conditions enhances the respective confidence levels to those for detection or projection. The empirical observations resulting from those experiments are directly attributable to the respective drivers. Confidence in attribution is enhanced if these experiments identify the underlying mechanisms and their responses. See text for the discussion of depicted examples and categories. Confidence assignments focus on the nature and size of effects, not on model capacity to reliably quantify their magnitude.]

6.6.1. Key Risks Related to Climate Change: Constraints on Ecosystem Services

Empirical studies provide evidence that climate change has impacted marine ecosystems (*high confidence*, FAQ 6.4, Table 6-6) and has caused changes in provisioning, regulating, and supportive Ecosystem Services (ES, *medium confidence*). Climate change may also have affected cultural services (*limited evidence* and *medium agreement*) but attribution of impacts to these services remains a challenge (*low confidence*), due to the intrinsic difficulties of assessing these services, the lack of long time-series data and confounding human impacts. In light of available understanding of cause and effect of climate change impacts on marine ecosystems (*high confidence*), future climate change will affect some ecosystem services (*high confidence* in projection, *medium confidence* in attribution). Projected changes in the availability of marine resources and ecosystem services are expected to affect economics, human livelihood and food security. Vulnerability is highest for the national economies of tropical coastal countries (*high confidence*).

[INSERT TABLE 6-6 HERE]

Table 6-6: Coastal and oceanic key risks from climate change and the potential for risk reduction through mitigation and adaptation. Key risks are identified based on assessment of the literature and expert judgments made by authors of the various WGII AR5 chapters, with supporting evaluation of evidence and agreement in the referenced chapter sections. Each key risk is characterized as very low, low, medium, high, or very high. Risk levels are presented for the near-term era of committed climate change (here, for 2030-2040), in which projected levels of global mean temperature increase do not diverge substantially across emissions scenarios. Risk levels are also presented for the longer-term era of climate options (here, for 2080-2100), for global mean temperature increase of 2°C and 4°C above preindustrial levels. For each timeframe, risk levels are estimated for the current state of adaptation and for a hypothetical highly adapted state. As the assessment considers potential impacts on different physical, biological, and human systems, risk levels should not necessarily be used to evaluate relative risk across key risks. Relevant climate variables are indicated by symbols. Acronyms for oceans sub-regions are as follows: HLSBS: High-Latitude Spring Bloom Systems; EUS: Equatorial Upwelling Systems; SES: Semi-Enclosed Seas; CBS: Coastal Boundary Systems; EBUE: Eastern Boundary Upwelling Ecosystems; STG: Sub-Tropical Gyres, DS: Deep Sea (>1000 m).]

Redistribution and constraints on microbial functions and primary productivity

Laboratory and mesocosm studies have identified various microbially mediated processes responding to climate-induced changes in light, nutrient supply, temperature, CO₂, and hypoxia (*high confidence*). Such processes include nitrogen fixation and the nitrogen cycle, carbon sequestration and export production, calcification, respiration, O₂ production, climate-feedback by DMS (dimethylsulphide) production, and nutrient recycling. However, changes in these Bio-Geochemical processes (BG) in the field are difficult to detect, project and attribute to climate change (*low confidence* 6.3.1-5).

The trends in Net Primary Production (NPP) recently reported for much of the low-latitude ocean using satellite observations differ considerably from those few long-term direct estimates of NPP at oceanic time-series sites (6.1.2, 6.3.4). Increased NPP at high latitudes (hNPP, detected and attributable to climate change with *medium confidence*, 6.3.4, Box CC-PP) are indicated by satellite images (*medium confidence*) and due to reduction and thinning of sea ice. Trends in NPP will be strengthened with further warming (*medium confidence*). Modeling projects that global NPP (gNPP) will decrease by 2100 under RCP scenarios (6.5.1, Box CC-PP, *medium confidence*).

Warming-induced species redistribution, loss of biodiversity and fisheries catch potential

Long-term observations show variability in oceanographic conditions with a key role of temperature and changing oceanographic regimes causing observed changes in ecosystem structure and fish stocks (*very high confidence*, cf. 30.7.1.1). Temperature Effects (TE) reflect the differential specialization of all life forms in limited ambient temperature ranges (*very high confidence*). Temperature exerts strong Macroorganism Effects (MAE), i.e. on animals and plants. Warming is presently causing and will cause species displacements and largely poleward shifts in biogeographic distribution of zooplankton and fishes, paralleled by altered seasonal activity, species abundance, migration, and body size (6.3.1., *high to very high confidence*), and leading to shifts in Community Composition (CC, *high confidence*, Box 6-1). Causes and effects are understood for fishes and most invertebrates via their Oxygen and Capacity Limited Thermal Tolerance (OCLTT, 6.3.1, *robust evidence, medium agreement and high confidence*). Such knowledge supports projections into the future (6.5, *high confidence*), which are influenced by the limited potential of organisms to adapt. Alterations in species ABundance (AB) result when organisms encounter shifting mean and extreme temperatures (*high confidence* in detection and attribution). Such trends will be exacerbated during future warming (*high confidence*, 6.5.1).

Among prominent examples, warming has caused and will cause northward shift and expansion of the geographic distribution of North Atlantic Cod (AC, 6.3.1, *high confidence* in detection or projection, *medium confidence* in detection or projection and attribution) and shifting growth patterns in relation to the distribution of Banded Morwong around New Zealand (BM, *high confidence* in detection or projection, *medium confidence* in detection or projection and attribution). Warming has shifted dominant species from Sardines to Anchovies in the Sea of Japan (SAJ, 6.3.1, 6.3.6, *medium confidence* in detection, *medium confidence* in detection and attribution). Warming extremes have reduced and will further reduce the abundance of Eelpout in the Wadden Sea (EWS, 6.3.1, *high confidence* in detection or projection, *high confidence* in detection or projection and attribution). Extreme warming events increase mortalities of Pacific Salmon during spawning migrations (PS, 6.3.1, *high confidence* in detection, *high confidence* in detection and attribution) in Fraser River, Canada. At temperate and high latitudes, communities display increasing fish Species Richness (SR) resulting from latitudinal shifts of species and attributed to warming and loss of sea ice, although the relative contributions of regional climate variation and long-term global trends have not been quantified (6.3.1, 6.5.2, *high confidence* in detection, *medium confidence* in detection and attribution). Latitudinal species shifts are projected to continue in the 21st century under all IPCC emission scenarios (*high confidence*, 6.3.1, 6.3.5, 6.3.7, 6.4.1, 6.5.2).

Climate-induced regime shifts and regional changes in Plankton Phenology (PP, *medium confidence*) have caused and will cause changes in food composition and availability to animals. Species shifts and changing species composition lead to changes in Fishery Catch Potential (FCP, *high confidence*, 5a in Figure 6-15), partly attributable to climate change (*high confidence*) and to sustained fishing pressure (6.5.3). Fisheries Catch Potentials (FCP) will be redistributed, decrease at low latitudes and increase at high latitudes (*high confidence*, 5a in figure). These trends will possibly be strengthened by the projected decrease in NPP at low latitudes and increase in NPP at high latitudes (*medium confidence*, 6.5.2-3, 5b in Figure 6-15). Polar Organisms (PO) that are unable to migrate to cooler waters, and to acclimatize or to adapt to warming, will become marginalized, contributing to the projected high species turnover in polar areas (*high confidence*, 6.3.1, 6.5.2).

Detected effects on Marine Air Breathers (MAB: mammals, seabirds and reptiles) include changing abundances and phenology, shifts in species distribution, and in sea turtle sex ratios (*high confidence*), all of which are partly attributed to climate change (*high confidence*). However, few effects are directly linked to climate drivers (e.g., temperature-driven turtle sex ratio); most effects are due to shifts in habitat structure (e.g., loss of sea ice), changing

availability of prey organisms, or changes in foraging efficiency, in both mammals (polar bears, walruses) and birds (penguins, albatrosses). Such trends will be exacerbated by future warming (*high confidence*, 6.3.7, 6.5.2).

Expanding hypoxia affecting marine resources

Hypoxic Zones in marine sediments and pelagic OMZs will continue to expand in the future, due to climate-induced warming trends (6.1.1). Local and regional Hypoxia Effects (HE) have been observed (*medium confidence*) and will be exacerbated in the future (*high confidence*, 6.3.3) causing habitat loss for groundfishes and pelagic predators and affecting the distribution of key zooplankton and nekton species (*medium confidence*). Progressive hypoxia is causing shifts in community composition toward hypoxia-tolerant species, excluding calcifiers due to elevated $p\text{CO}_2$ (*high confidence*), benefiting specialized microbes, and leading to reduced biodiversity and the loss of higher life forms (*high confidence*) (6.3.3). Loss of deep habitat and biomass of Mid-Water Fishes (MWF, 6.3.3, *medium confidence* in detection) off California is also attributed to hypoxia (*medium confidence*). These trends will continue into the future (*medium confidence*).

Constraints on marine calcifiers and associated fisheries and aquaculture due to ocean acidification

Ocean acidification will exert negative effects on species and whole ecosystems and their services, especially those relying on carbonate structures such as warm-water coral reefs (*high confidence*, cf. 30.7.1.2). Presently, only a small number of field observations have detected Ocean Acidification Effects (OAE) (*medium confidence*), but experiments and natural analogues support reliable but qualitative projections and attribution (*high confidence*). A specific glimpse into the future of anthropogenic OA is provided by negative Effects of upwelled CO_2 -rich waters on Pacific Oysters (EO) introduced to aquaculture along the North American west coast (*high confidence* in detection, *low confidence* in attribution to anthropogenic causes). Findings in experimental laboratory and field studies as well as at natural analogues support attribution of projected effects to future CO_2 concentrations (*medium confidence*), with species-specific sensitivities across phyla (*high confidence*). Projected effects are most harmful to strong Calcifiers (CAL, *high confidence*), e.g., some echinoderms, bivalves, gastropods, warm-water corals, and crustose algae, and less harmful to some crustaceans and, possibly, fishes. Projections from experimental studies and observations at natural analogues indicate shifts in Community Composition (CC) to more active animals and from calcifiers (CAL) to non-calcifiers in all organism groups (*high confidence* in both projection and attribution to increased CO_2 , 6.3.2, Table 6-3).

Interactions of climate-related drivers exacerbating impacts on organisms, ecosystems, and their services

Climate change involves interactions of temperature with other climate related drivers and their effects (ocean acidification, hypoxia, freshening, nutrient supply, organism shifts resulting in changing interactions between species, changes in habitat structure, e.g., loss of sea ice). Strong interactions with other human impacts like eutrophication, fishing, and other forms of harvesting accelerate and amplify climate-induced changes (*high confidence*, 6.3.5; 30.7.1.1). Harmful algal blooms (HABs) will be stimulated by warming, nutrient fluctuations in upwelling areas, eutrophication in coastal areas (Table 6-6), ocean acidification, and enhanced surface stratification (*medium confidence*). Synergistic Effects (SE) will be exacerbated in the future (*medium confidence*), but have not yet been clearly detected and attributed in the field (*low confidence*). For projected future effects, attribution of observed impacts to such synergisms is supported by experimental evidence, especially in animals and plants (*medium confidence*).

Increased bleaching and decreased calcification displayed by several Reef-building Warm-water Corals (RWC, *very high confidence*) over the last three decades are attributed to the ongoing warming trend, and the associated rise in extreme temperature events and amplitudes (*high confidence*, 6.3.1; Box CC-CR; 30.5.6). Such trends will be exacerbated by future warming and synergistic effects (*high confidence*, cf. 30.5.4.2), with some amelioration by latitudinal shifts and evolutionary adaptation (6.3.1, *low confidence*). Ocean acidification will have an increasing influence on reefs (*high confidence*), as indicated by similar phenomena during mass extinctions in Earth history.

6.6.2. Key Uncertainties

Key uncertainties result from insufficient knowledge of ocean systems. International organizations (both inter- and non-governmental) have the opportunity to play a key role in coordinating research concepts and approaches,

working toward a coherent picture of climate change effects on the global ocean. Countries around the world have limited capacity and infrastructure to study the ocean's response to climate change. Long-term observational time series are especially lacking, in both quantity and quality. Research has provided valuable insights, but a unifying approach addressing principles across organism domains and ecosystems is still missing. Processes investigated so far differ largely by study organisms (plants, animals, phytoplankton, and bacteria) and by level of organization (ecosystem, whole organism, tissue, cell, molecular). Especially for microbes, available data are patchy and reported trends are often in different directions, partly due to different experimental protocols and/or over-reliance on species or strains of microbes that are readily culturable, and hence have been used for decades in laboratory research. The knowledge base of climate impacts on species, strains or communities in the field is insufficient. Scaling from physiological studies on individual species to ecosystem changes has been successful in individual cases but has not been widely implemented, e.g., to shifts in species interactions or food webs. An integrated framework of climate sensitivity at the ecosystem level that considers multiple drivers and their interactive effects needs to be developed further. This includes an in depth understanding of ecosystem structure (physical and biological) and functioning, of ecosystem complexity and species interactions, and of the resulting implications for biogeochemical processes. For all climate drivers, especially ocean warming, acidification, and hypoxia, studies integrating mechanistic knowledge and evolutionary adaptation over generations are needed. Research should also cover various climate zones and biomes. Laboratory and modeling experiments are needed to test hypotheses building on long-term field observations and observations at natural or paleo-analogues. Models should better integrate observations and mechanism-based understanding, and better project future interactions between human and natural systems in a changing climate.

Frequently Asked Questions

FAQ 6.1: Why are climate impacts on oceans and their ecosystems so important?

[to be placed in 6.1, before 6.1.1]

Oceans create half the oxygen (O₂) we use to breathe and burn fossil fuels. Oceans provide on average 20% of the animal protein consumed by more than 1.5 billion people. Oceans are home to species and ecosystems valued in tourism and for recreation. The rich biodiversity of the oceans offers resources for innovative drugs or biomechanics. Ocean ecosystems such as coral reefs and mangroves protect the coastlines from tsunamis and storms. About 90% of the goods the world uses are shipped across the oceans. All these activities are affected by climate change.

Oceans play a major role in global climate dynamics. Oceans absorb 93% of the heat accumulating in the atmosphere, and the resulting warming of oceans affects most ecosystems. About a quarter of all the carbon dioxide (CO₂) emitted from the burning of fossil fuels is absorbed by oceans. Plankton converts some of that CO₂ into organic matter, part of which is exported into the deeper ocean. The remaining CO₂ causes progressive acidification from chemical reactions between CO₂ and seawater, acidification being exacerbated by nutrient supply and with the spreading loss of oxygen content. These changes all pose risks for marine life and may affect the oceans' ability to perform the wide range of functions that are vitally important for environmental and human health.

The effects of climate change occur in an environment that also experiences natural variability in many of these variables. Other human activities also influence ocean conditions, such as overfishing, pollution, and nutrient runoff via rivers that causes eutrophication, a process that produces large areas of water with low oxygen levels (sometimes called 'Dead Zones'). The wide range of factors that affect ocean conditions and the complex ways these factors interact make it difficult to isolate the role any one factor plays in the context of climate change, or to identify with precision the combined effects of these multiple drivers.

FAQ 6.2: What is different about the effects of climate change on the oceans compared to the land, and can we predict the consequences? *[to be placed in 6.3, before 6.3.1]*

The ocean environment is unique in many ways. It offers large-scale aquatic habitats, diverse bottom topography, and a rich diversity of species and ecosystems in water in various climate zones that are found nowhere else.

One of the major differences in terms of the effect of climate change on the oceans compared to land is ocean acidification. Anthropogenic CO₂ enters the ocean and chemical reactions turn some of it to carbonic acid, which acidifies the water. This mirrors what is also happening inside organisms once they take up the additional CO₂. Marine species that are dependent on calcium carbonate, like shellfish, seastars and corals, may find it difficult to build their shells and skeletons under ocean acidification. In general, animals living and breathing in water like fish,

squid, and mussels, have between five and 20 times less CO₂ in their blood than terrestrial animals, so CO₂ enriched water will affect them in different and potentially more dramatic ways than species that breathe in air.

Consider also the unique impacts of climate change on ocean dynamics. The ocean has layers of warmer and colder water, saltier or less saline water, and hence less or more dense water. Warming of the ocean and the addition of more freshwater at the surface through ice melt and higher precipitation increases the formation of more stable layers stratified by density, which leads to less mixing of the deeper, denser, and colder nutrient-rich layers with the less dense nutrient-limited layers near the surface. With less mixing, respiration by organisms in the mid-water layers of stratified oceans will produce oxygen-poor waters, so-called oxygen minimum zones (OMZs). Large, more active fish can't live in these oxygen poor waters, while more simple specialized organisms with a lower need for oxygen will remain, and even thrive in the absence of predation from larger species. Therefore, the community of species living in hypoxic areas will shift.

State-of-the-art ecosystem models build on empirical observations of past climate changes and enable development of estimates of how ocean life may react in the future. One such projection is a large shift in the distribution of commercially important fish species to higher latitudes and reduced harvesting potential in their original areas. But producing detailed projections, e.g. what species and how far they will shift, is challenging because of the number and complexity of interactive feedbacks that are involved. At the moment, the uncertainties in modeling and complexities of the ocean system even prevent any quantification of how much of the present changes in the oceans is being caused by anthropogenic climate change or natural climate variability, and how much by other human activities such as fishing, pollution, etc.

It is known, however, that the resilience of marine ecosystems to adjust to climate change impacts is likely to be reduced by both the range of factors and their rate of change. The current rate of environmental change is much faster than most climate changes in the Earth's history, so predictions from longer term geological records may not be applicable if the changes occur within a few generations of a species. A species that had more time to adapt in the past may simply not have time to adapt under future climate change.

FAQ 6.3: Why are some marine organisms affected by ocean acidification? [to be placed in 6.3.2, before 6.3.2.1]

Many marine species, from microscopic plankton to shellfish and coral reef builders, are referred to as calcifiers, species that use solid calcium carbonate (CaCO₃) to construct their skeletons or shells. Seawater contains ample calcium but to use it and turn it into calcium carbonate, species have to bring it to specific sites in their bodies and raise the alkalinity (lower the acidity) at these sites to values higher than in other parts of the body or in ambient seawater. That takes energy. If high CO₂ levels from outside penetrate the organism and alter internal acidity levels, keeping the alkalinity high takes even more energy. The more energy is needed for calcification, the less is available for other biological processes like growth or reproduction, reducing the organisms' weight and overall competitiveness and viability.

Exposure of external shells to more acidic water can affect their stability by weakening or actually dissolving carbonate structures. Some of these shells are shielded from direct contact with seawater by a special coating that the animal makes (as is the case in mussels). The increased energy needed for making the shells to begin with impairs the ability of organisms to protect and repair their dissolving shells. Presently, more acidic waters brought up from the deeper ocean to the surface by wind and currents off the Northwest coast of the United States are having this effect on oysters grown in aquaculture.

Ocean acidification not only affects species producing calcified exoskeletons. It affects many more organisms either directly or indirectly and has the potential to disturb food webs and fisheries. Most organisms that have been investigated display greater sensitivity at extreme temperatures, so as ocean temperatures change, those species that are forced to exist at the edges of their thermal ranges will experience stronger effects of acidification.

FAQ 6.4: What changes in marine ecosystems are likely because of climate change? [to be placed after 6.3.8]

There is general consensus among scientists that climate change significantly affects marine ecosystems and may have profound impacts on future ocean biodiversity. Recent changes in the distribution of species as well as species richness within some marine communities and the structure of those communities have been attributed to ocean warming. Projected changes in physical and biogeochemical drivers such as temperature, CO₂ content and acidification, oxygen levels, the availability of nutrients, and the amount of ocean covered by ice, will affect marine life.

Overall, climate change will lead to large-scale shifts in the patterns of marine productivity, biodiversity, community composition and ecosystem structure. Regional extinction of species that are sensitive to climate change

will lead to a decrease in species richness. In particular, the impacts of climate change on vulnerable organisms such as warm water corals are expected to affect associated ecosystems, such as coral reef communities.

Ocean primary production of the phytoplankton at the base of the marine food chain is expected to change but the global patterns of these changes are difficult to project. Existing projections suggest an increase in primary production at high latitudes such as the Arctic and the Southern Ocean (because the amount of sunlight available for photosynthesis of phytoplankton goes up as the amount of water covered by ice decreases). Decreases are projected for ocean primary production in the tropics and at mid-latitudes because of reduced nutrient supply. Alteration of the biology, distribution, and seasonal activity of marine organisms will disturb food web interactions such as the grazing of copepods (tiny crustaceans) on planktonic algae, another important foundational level of the marine food chain. Increasing temperature, nutrient fluctuations, and human-induced eutrophication may support the development of harmful algal blooms in coastal areas. Similar effects are expected in upwelling areas where wind and currents bring colder and nutrient rich water to the surface. Climate change may also cause shifts in the distribution and abundance of pathogens such as those that cause cholera.

Most climate change scenarios foresee a shift or expansion of the ranges of many species of plankton, fish and invertebrates towards higher latitudes, by tens of kilometres per decade, contributing to changes in species richness and altered community composition. Organisms less likely to shift to higher latitudes because they are more tolerant of the direct effects of climate change or less mobile may also be affected because climate change will alter the existing food webs on which they depend.

In polar areas, populations of species of invertebrates and fish adapted to colder waters may decline as they have no place to go. Some of those species may face local extinction. Some species in semi-enclosed seas such as the Wadden Sea and the Mediterranean Sea, also face higher risk of local extinction because land boundaries around those bodies of water will make it difficult for those species to move laterally to escape waters that may be too warm.

Cross-Chapter Boxes

Box CC-CR. Coral Reefs

[Jean-Pierre Gattuso (France), Ove Hoegh-Guldberg (Australia), Hans-Otto Pörtner (Germany)]

Coral reefs are shallow-water ecosystems that consist of reefs made of calcium carbonate which is mostly secreted by reef-building corals and encrusting macroalgae. They occupy less than 0.1% of the ocean floor yet play multiple important roles throughout the tropics, housing high levels of biological diversity as well as providing key ecosystem goods and services such as habitat for fisheries, coastal protection and appealing environments for tourism (Wild *et al.*, 2011). About 275 million people live within 30 km of a coral reef (Burke *et al.*, 2011) and derive some benefits from the ecosystem services that coral reefs provide (Hoegh-Guldberg, 2011) including provisioning (food, livelihoods, construction material, medicine), regulating (shoreline protection, water quality), supporting (primary production, nutrient cycling) and cultural (religion, tourism) services. This is especially true for the many coastal and small island nations in the world's tropical regions (29.3.3.1).

Coral reefs are one of the most vulnerable marine ecosystems (*high confidence*; 5.4.2.4, 6.3.1, 6.3.2, 6.3.5, 25.6.2, and 30.5) and more than half of the world's reefs are under medium or high risk of degradation (Burke *et al.*, 2011). Most human-induced disturbances to coral reefs were local until the early 1980s (e.g., unsustainable coastal development, pollution, nutrient enrichment and overfishing) when disturbances from ocean warming (principally mass coral bleaching and mortality) began to become widespread (Glynn, 1984). Concern about the impact of ocean acidification on coral reefs developed over the same period, primarily over the implications of ocean acidification for the building and maintenance of the calcium carbonate reef framework (Box CC-OA).

[INSERT FIGURE CR-1 HERE]

Figure CR-1: A and B: the same coral community before and after a bleaching event in February 2002 at 5 m depth, Halfway Island, Great Barrier Reef. Coral cover at the time of bleaching was 95% bleached almost all of it severely bleached, resulting in mortality of 20.9% (Elvidge *et al.*, 2004). Mortality was comparatively low due in part because these coral communities were able to shuffle their symbiont to more thermo-tolerant types (Berkelmans and van Oppen, 2006; Jones *et al.*, 2008). C and D: three CO₂ seeps in Milne Bay Province, Papua New Guinea show that prolonged exposure to high CO₂ is related to fundamental changes in the ecology of coral reefs (Fabricius *et al.*,

2011), including reduced coral diversity (-39%), severely reduced structural complexity (-67%), lower density of young corals (-66%) and fewer crustose coralline algae (-85%). At high CO₂ sites (panel D; median pH_T ~7.8), reefs are dominated by massive corals while corals with high morphological complexity are underrepresented compared with control sites (D; median pH ~8.0). Reef development ceases at pH_T values below 7.7. pH_T: pH on the total scale. E: temporal trend in coral cover for the whole Great Barrier Reef over the period 1985–2012 (N, number of reefs, mean ± 2 standard errors; De'ath et al., 2012). F: composite bars indicate the estimated mean coral mortality for each year, and the sub-bars indicate the relative mortality due to crown-of-thorns starfish, cyclones, and bleaching for the whole Great Barrier Reef (De'ath et al., 2012). Photo credit: R. Berkelmans (A and B) and K. Fabricius (C and D).]

A wide range of climatic and non-climatic drivers affect corals and coral reefs and negative impacts have already been observed (5.4.2.4, 6.3.1, 6.3.2, 25.6.2.1, 30.5.3, 30.5.6). Bleaching involves the breakdown and loss of endosymbiotic algae, which live in the coral tissues and play a key role in supplying the coral host with energy (see 6.3.1. for physiological details and 30.5 for a regional analysis). Mass coral bleaching and mortality, triggered by positive temperature anomalies (*high confidence*), is the most widespread and conspicuous impact of climate change (Figure CR-1A and B, Figure 5-3; 5.4.2.4, 6.3.1, 6.3.5, 25.6.2.1, 30.5 and 30.8.2). For example, the level of thermal stress at most of the 47 reef sites where bleaching occurred during 1997–98 was unmatched in the period 1903 to 1999 (Lough, 2000). Ocean acidification reduces biodiversity (Figure CR-1C and D) and the calcification rate of corals (*high confidence*; 5.4.2.4, 6.3.2, 6.3.5) while at the same time increasing the rate of dissolution of the reef framework (*medium confidence*; 5.2.2.4) through stimulation of biological erosion and chemical dissolution. Taken together, these changes will tip the calcium carbonate balance of coral reefs towards net dissolution (*medium confidence*; 5.4.2.4). Ocean warming and acidification have synergistic effects in several reef-builders (5.2.4.2, 6.3.5). Taken together, these changes will erode habitats for reef-based fisheries, increase the exposure of coastlines to waves and storms, as well as degrading environmental features important to industries such as tourism (*high confidence*; 6.4.1.3, 25.6.2, 30.5).

A growing number of studies have reported regional scale changes in coral calcification and mortality that are consistent with the scale and impact of ocean warming and acidification when compared to local factors such as declining water quality and overfishing (Hoegh-Guldberg *et al.*, 2007). The abundance of reef building corals is in rapid decline in many Pacific and SE Asian regions (*very high confidence*, 1–2% per year for 1968–2004; Bruno and Selig, 2007). Similarly, the abundance of reef-building corals has decreased by over 80% on many Caribbean reefs (1977 to 2001; Gardner *et al.*, 2003), with a dramatic phase shift from corals to seaweeds occurring on Jamaican reefs (Hughes, 1994). Tropical cyclones, coral predators and thermal stress-related coral bleaching and mortality have led to a decline in coral cover on the Great Barrier Reef by about 51% between 1985 and 2012 (Figure CR-1E and F). Although less well documented, benthic invertebrates other than corals are also at risk (Przeslawski *et al.*, 2008). Fish biodiversity is threatened by the permanent degradation of coral reefs, including in a marine reserve (Jones *et al.*, 2004).

Future impacts of climate-related drivers (ocean warming, acidification, sea level rise as well as more intense tropical cyclones and rainfall events) will exacerbate the impacts of non-climate related drivers (*high confidence*). Even under optimistic assumptions regarding corals being able to rapidly adapt to thermal stress, one-third (9 to 60%, 68% uncertainty range) of the world's coral reefs are projected to be subject to long-term degradation (next few decades) under the RCP3-PD scenario (Frieler *et al.*, 2013). Under the RCP4.5 scenario, this fraction increases to two-thirds (30 to 88%, 68% uncertainty range). If present day corals have residual capacity to acclimate and/or adapt, half of the coral reefs may avoid high frequency bleaching through 2100 (*limited evidence, limited agreement*; Logan *et al.*, 2013). Evidence of corals adapting rapidly, however, to climate change is missing or equivocal (Hoegh-Guldberg, 2012).

Damage to coral reefs has implications for several key regional services:

- *Resources*: Coral reefs account for 10 to 12% of the fish caught in tropical countries, and 20 to 25% of the fish caught by developing nations (Garcia and Moreno, 2003). Over half (55%) of the 49 island countries considered by Newton *et al.* (2007) are already exploiting their coral reef fisheries in an unsustainable way and the production of coral reef fish in the Pacific is projected to decrease 20% by 2050 under the SRES A2 emissions scenario (Bell *et al.*, 2013).

- *Coastal protection*: Coral reefs contribute to protecting the shoreline from the destructive action of storm surges and cyclones (Sheppard *et al.*, 2005), sheltering the only habitable land for several island nations, habitats suitable for the establishment and maintenance of mangroves and wetlands, as well as areas for recreational activities. This role is threatened by future sea level rise, the decrease in coral cover, reduced rates of calcification and higher rates of dissolution and bioerosion due to ocean warming and acidification (5.4.2.4, 6.4.1, 30.5).
- *Tourism*: More than 100 countries benefit from the recreational value provided by their coral reefs (Burke *et al.*, 2011). For example, the Great Barrier Reef Marine Park attracts about 1.9 million visits each year and generates A\$ 5.4 billion to the Australian economy and 54,000 jobs (90% in the tourism sector; Biggs, 2011).

Coral reefs make a modest contribution to the Global Domestic Product but their economic importance can be high at the country and regional scales (Pratchett *et al.*, 2008). For example, tourism and fisheries represent 5% of the GDP of South Pacific islands (average for 2001–2011; Laurans *et al.*, 2013). At the local scale, these two services provided in 2009–2011 at least 25% of the annual income of villages in Vanuatu and Fiji (Pascal, 2011; Laurans *et al.*, 2013).

Isolated reefs can recover from major disturbance, and the benefits of their isolation from chronic anthropogenic pressures can outweigh the costs of limited connectivity (Gilmour *et al.*, 2013). Marine protected areas (MPAs) and fisheries management have the potential to increase ecosystem resilience and increase the recovery of coral reefs after climate change impacts such as mass coral bleaching (McLeod *et al.*, 2009). Although they are key conservation and management tools, they are unable to protect corals directly from thermal stress (Selig *et al.*, 2012) suggesting that they need to be complemented with additional and alternative strategies (Rau *et al.*, 2012; Billé *et al.*, 2013). While MPA networks are a critical management tool, they should be established considering other forms of resource management (e.g., fishery catch limits and gear restrictions) and integrated ocean and coastal management to control land-based threats such as pollution and sedimentation. There is *medium confidence* that networks of highly protected areas nested within a broader management framework can contribute to preserving coral reefs under increasing human pressure at local and global scales (Salm *et al.* 2006). Locally, controlling the input of nutrients and sediment from land is an important complementary management strategy (McLeod *et al.*, 2009) because nutrient enrichment can increase the susceptibility of corals to bleaching (Wiedenmann *et al.*, 2012) and coastal pollutants enriched with fertilizers can increase acidification (Kelly *et al.*, 2011). In the long term, limiting the amount of ocean warming and acidification is central to ensuring the viability of coral reefs and dependent communities (*high confidence*; 5.2.4.4, 30.5).

CC-CR References

- Bell J. D., A. Ganachaud, P.C. Gehrke, S.P. Griffiths, A.J. Hobday, O. Hoegh-Guldberg, J.E. Johnson, R. Le Borgne, P. Lehodey, J.M. Lough, R.J. Matear, T.D. Pickering, M.S. Pratchett, A. Sen Gupta, I. Senina I. and M. Waycott . 2013: Mixed responses of tropical Pacific fisheries and aquaculture to climate change. *Nature Climate Change* **3**, 591-591.
- Berkelmans R. and M.J.H. van Oppen, 2006: The role of zooxanthellae in the thermal tolerance of corals: a ‘nugget of hope’ for coral reefs in an era of climate change. In: *Proceedings of the Royal Society of London. Series B: Biological Sciences*, **273**, 2305-2312.
- Biggs D., 2011. Case study: the resilience of the nature-based tourism system on Australia’s Great Barrier Reef. Report prepared for the Australian Department of Sustainability, Environment, Water, Population and Communities on behalf of the State of the Environment 2011 Committee Government. 32 p.
- Billé R., R. Kelly, E. Harrould-Kolieb, D. Herr, F. Joos, K.J. Kroeker, D. Laffoley, A. Oschlies and J.P. Gattuso , 2013: Taking action against ocean acidification: a review of management and policy options. *Environmental Management*, **52**, 761-779.
- Bruno J. F. and E.R. Selig, 2007: Regional decline of coral cover in the Indo-Pacific: timing, extent, and subregional comparisons. *PLoS ONE*, **2**(8), e711.
- Burke L. M., K. Reytar, M. Spalding and A. Perry, 2011: Reefs at risk revisited. World Resources Institute, Washington, DC.: p.114.
- De’ath G., K.E. Fabricius, H. Sweatman and M. Puotinen, 2012: The 27-year decline of coral cover on the Great Barrier Reef and its causes. In: *Proceedings of the National Academy of Science U.S.A.* **109**, 17995-17999.
- Elvidge C., J. Dietz, R. Berkelmans, S. Andréfouët, W. Skirving, A. Strong and B. Tuttle, 2004: Satellite observation of Keppel Islands (Great Barrier Reef) 2002 coral bleaching using IKONOS data. *Coral Reefs* **23**, 123-132.

- Fabricius K. E.**, C. Langdon, S. Uthicke, C. Humphrey, S. Noonan, G. De'ath, R. Okazak, N. Muehlehner, M.S. Glas and J.M. Lough, 2011: Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations. *Nature Climate Change* **1**, 165-169.
- Frieler K.**, M. Meinshausen, A. Golly, M. Mengel, K. Lebek, S.D. Donner and O. Hoegh-Guldberg, 2013: Limiting global warming to 2 °C is unlikely to save most coral reefs. *Nature Climate Change* **3**, 165-170.
- Garcia S. M.** and I. de Leiva Moreno, 2003: Global overview of marine fisheries. In: *Responsible fisheries in the marine ecosystem*, [Sinclair M. and Valdimarsson G. (eds.)], Wallingford: CABI pp. 1-24.
- Gardner T. A.**, I.M. Cote, J.A. Gill, A. Grant and A.R. Watkinson, 2003: Long-term region-wide declines in Caribbean corals. *Science* **301(5635)**, 958-960.
- Gilmour J. P.**, LD. Smith, A.J. Heyward, A.H. Baird and M.S. Pratchett M. S., 2013: Recovery of an isolated coral reef system following severe disturbance. *Science* **340**, 69-71.
- Glynn P. W.**, 1984: Widespread coral mortality and the 1982-83 El Niño warming event. *Environmental Conservation* **11**, 133-146.
- Hoegh-Guldberg O.**, 2011: Coral reef ecosystems and anthropogenic climate change. *Regional Environmental Change* **11**, 215-227.
- Hoegh-Guldberg O.**, 2012: The adaptation of coral reefs to climate change: is the Red Queen being outpaced? *Scientia Marina* **76**, 403-408.
- Hoegh-Guldberg, O.**, P. J. Mumby, A. J. Hooten, R. S. Steneck, P. Greenfield, E. Gomez, C. D. Harvell, P. F. Sale, A. J. Edwards, K. Caldeira, N. Knowlton, C. M. Eakin, R. Iglesias-Prieto, N. Muthiga, R. H. Bradbury, A. Dubi, and M. E. Hatzios, 2007: Coral reefs under rapid climate change and ocean acidification. *Science* **318**, 1737-1742.
- Hughes T. P.**, 1994. Catastrophes, phase shifts, and large-scale degradation of a Caribbean coral reef. *Science* **265(5178)**, 1547-1551.
- Jones A. M.**, R. Berkelmans, M.J. van Oppen, J.C. Mieog and W. Sinclair, 2008: A community change in the algal endosymbionts of a scleractinian coral following a natural bleaching event: field evidence of acclimatization. In: *Proceedings of the Royal Society of London. Series B: Biological Sciences* **275**, 1359-1365.
- Jones G. P.**, M.I. McCormick, M. Srinivasan and J.V. Eagle, 2004: Coral decline threatens fish biodiversity in marine reserves. In: *Proceedings of the National Academy of Science U.S.A.* **101**, 8251-8253.
- Kelly R. P.**, M.M. Foley, W.S. Fisher, R.A. Feely, B.S. Halpern, G.G. Waldbusser and M.R. Caldwell, 2011: Mitigating local causes of ocean acidification with existing laws. *Science* **332**, 1036-1037.
- Laurans Y.**, N. Pascal, T. Binet, L. Brander, E. Clua, G. David, D. Rojat and A. Seidl, 2013: Economic valuation of ecosystem services from coral reefs in the South Pacific: taking stock of recent experience. *Journal of Environmental Management* **116C**, 135-144.
- Logan C. A.**, J.P. Dunne, C.M. Eakin and S.D. Donner, 2013: Incorporating adaptation and acclimatization into future projections of coral bleaching. *Global Change Biology*, doi:10.1111/gcb.12390.
- Lough J. M.**, 2000: 1997-98: Unprecedented thermal stress to coral reefs? *Geophysical Research Letters*. **27(23)**, 3901-3904.
- McLeod E.**, R. Salm, A. Green and J. Almany, 2009: Designing marine protected area networks to address the impacts of climate change. *Frontiers in Ecology and the Environment* **7**, 362-370.
- Newton K.**, I. M. Côté, G.M. Pilling G, S. Jennings and N.K. Dulvy, 2007: Current and future sustainability of island coral reef fisheries. *Current Biology* **17**. 655-658.
- Pascal N.**, 2011. Cost-benefit analysis of community-based marine protected areas: 5 case studies in Vanuatu. Moorea, French Polynesia: CRISP-CRIOBE, 107p.
- Pratchett M. S.**, P.L. Munday and S.K. Wilson, 2008: Effects of climate-induced coral bleaching on coral-reef fishes- Ecological and economic consequences. *Oceanography and Marine Biology: an Annual Review* **46**, 251-296.
- Przeslawski R.**, A. Ahyong, M. Byrne, G. Worheide and P. Hutchings, 2008: Beyond corals and fish: the effects of climate change on noncoral benthic invertebrates of tropical reefs. *Global Change Biology* **14**, 2773-2795.
- Rau G. H.**, E.L. McLeod and O. Hoegh-Guldberg, 2012: The need for new ocean conservation strategies in a high-carbon dioxide world. *Nature Climate Change* **2**, 720-724.
- Salm RV**, T. Done and E. Mcleod, 2006: Marine protected area planning in a changing climate. In: *Coral Reefs and Climate Change: Science and Management*. [Phinney, J.T., Hoegh- Guldberg O, J. Kleypas, et al. (eds)]. Washington, DC: American Geophysical Union 244 pp..
- Selig E. R.**, K.S. Casey and J.F. Bruno, 2012: Temperature-driven coral decline: the role of marine protected areas. *Global Change Biology* **18**, 1561-1570.
- Sheppard C.**, D.J. Dixon, M. Gourlay, A. Sheppard and R. Payet, 2005: Coral mortality increases wave energy reaching shores protected by reef flats: examples from the Seychelles. *Estuarine, Coastal and Shelf Science* **64**, 223-234.
- Wiedenmann J.**, C. D'Angelo, E.G. Smith, A.N. Hunt, F.E. Legiret, A.D. Postle and E.P. Achterberg, 2013: Nutrient enrichment can increase the susceptibility of reef corals to bleaching. *Nature Climate Change* **3**, 160-164.
- Wild C.**, O. Hoegh-Guldberg, M.S. Naumann, M. Florencia Colombo-Pallotta, M. Ateweberhan, W.K. Fitt , R. Iglesias-Prieto, C. Palmer, J.C. Bythell, J.-C.Ortiz, Y. Loya and R. van Woesik, 2011: Climate change impedes scleractinian corals as primary reef ecosystem engineers. *Marine and Freshwater Research* **62**, 205-215.

Box CC-MB. Observed Global Responses of Marine Biogeography, Abundance, and Phenology to Climate Change

[Elvira Poloczanska (Australia), Ove Hoegh-Guldberg (Australia), William Cheung (Canada), Hans O. Pörtner (Germany), Michael Burrows (UK)]

WGII AR4 presented detection and attribution of a global climate change fingerprint on natural systems (AR4, Ch 1, SPM Figure 1), but studies from marine systems were mostly absent. Since AR4, there has been a rapid increase in studies that focus on climate change impacts on marine species, which represents an opportunity to move from more anecdotal evidence to examining and potentially attributing detected changes within the Ocean to climate change (6.3, Figure MB-1). Recent changes in populations of marine species and the associated shifts in diversity patterns are resulting, at least partly, from climate change-mediated biological responses across ocean regions (6.2, Table 6.7, 30.5) (*robust evidence, high agreement, high confidence*).

Poloczanska *et al.* (2013) assess a potential pattern in responses of ocean life to recent climate change using a global database of 208 peer-reviewed papers. Observed responses (n=1735) were recorded from 857 species or assemblages across regions and taxonomic groups, from phytoplankton to marine reptiles and mammals (Figure MB-1). Observations were defined as those where the authors of a particular paper assessed the occurrence change in a biological parameter (including distribution, phenology, abundance, demography or community composition) and, if change occurs, the consistency of the change with that expected under climate change. Studies from the peer-reviewed literature were selected using three criteria: (1) authors inferred or directly tested for trends in biological and climatic variables; (2) included data after 1990; and (3) observations spanned at least 19 years, to reduce bias resulting from biological responses to short-term-climate variability.

[INSERT FIGURE MB-1 HERE

Figure MB-1: 1735 observed responses to climate change from 208 single- and multi-species studies. Changes attributed to climate change (blue), inconsistent with climate change (red) and are equivocal (yellow). Each circle represents the centre of a study area. Where points fall on land, it is because they are centroids of distribution that surround an island or peninsula. Pie charts show the proportions within regions bounded by red squares and in the Mediterranean; numbers indicate the total (consistent, opposite or equivocal) observations within each region. Note: 57% of the studies included were published since AR4 (from Poloczanska *et al.*, 2013).]

The results of this meta-analysis show that climate change has already had widespread impacts on species' distribution, abundance, phenology, and subsequently, species richness and community composition across a broad range of taxonomic groups (plankton to top predators). Of the observations that showed a response in either direction, changes in phenology, distribution and abundance were overwhelmingly (81%) in a direction that was consistent with theoretical responses to climate change (6.2). Knowledge gaps exist, especially in equatorial sub-regions and the Southern Hemisphere (Figure MB-1).

The timing of many biological events (phenology) had an earlier onset. For example, over the last 50 years, spring events shifted earlier for many species with an average advancement of 4.4 ± 0.7 days decade⁻¹ (mean \pm SE) and summer events by 4.4 ± 1.1 days decade⁻¹ (*robust evidence, high agreement, high confidence*) (Figure MB-2). Phenological observations included in the study, range from shifts in peak abundance of phytoplankton and zooplankton, to reproduction and migration of invertebrates, fishes and seabirds (6.3.2, 30.5).

The distributions of benthic, pelagic and demersal species and communities have shifted by 10s to 1000s of km, although the range shifts have not been uniform across taxonomic groups or ocean regions (6.3.2, 30.5) (*robust evidence, high agreement, high confidence*). Overall, leading range edges expanded in a poleward direction at 72.0 ± 13.5 km decade⁻¹ and trailing edges contracted in a poleward direction at 15.8 ± 8.7 km decade⁻¹ (Figure MB-2) revealing much higher current rates of migration than the potential maximum rates reported for terrestrial species (Figure 4.6) despite slower warming of the Ocean than land surface (WG1 3.2).

[INSERT FIGURE MB-2 HERE

Figure MB-2. Rates of change in distribution (km decade⁻¹) for marine taxonomic groups, measured at the leading edges (red) and trailing edges (brown). Average distribution shifts calculated using all data, regardless of range

location, are in black. Distribution rates have been square-root transformed; standard errors may be asymmetric as a result. Positive distribution changes are consistent with warming (into previously cooler waters, generally poleward). Means \pm standard error are shown, along with number of observations (from Poloczanska *et al.*, 2013).]

Poleward distribution shifts have resulted in increased species richness in mid to high latitude regions (Hiddink and ter Hofstede, 2008) and changing community structure (Simpson *et al.*, 2011) (28.2.2). Increases in warm-water components of communities concurrent with regional warming have been observed in mid to high latitude ocean regions including the Bering Sea, Barents Sea, Nordic Sea, North Sea, and Tasman Sea (Box 6.1, 30.5). Observed changes in species composition of catches from 1970–2006 that is partly attributed to long-term ocean warming suggest increasing dominance of warmer water species in sub-tropical and higher latitude regions, and reduction in abundance of sub-tropical species in equatorial waters (Cheung *et al.*, 2013), with implications for fisheries (6.5, 7.4.2, 30.6.2.1)

The magnitude and direction of distribution shifts can be related to temperature velocities (i.e., the speed and direction at which isotherms propagate across the Ocean’s surface (30.3.1.1, Burrows *et al.* 2011). Pinsky *et al.* (2013) showed that shifts in both latitude and depth of benthic fish and crustaceans could be explained by climate velocity with remarkable accuracy, using a database of 128 million individuals across 360 marine taxa from surveys of North American coastal waters conducted over 1968 to 2011. Poloczanska *et al.* (2013) found that faster distribution shifts generally occur in regions of highest surface temperature velocity, such as the North Sea and sub-Arctic Pacific Ocean. Observed marine species shifts, since approximately 1950s, have generally been able to track observed velocities (Fig MB-3), with phyto- and zooplankton distribution shifts vastly exceeding climate velocities, but with considerable variability within and among taxonomic groups (Poloczanska *et al.* 2013).

Biogeographic shifts are also be influenced by other factors such as nutrient and stratification changes, species’ interactions, habitat availability and fishing (6.3). Rate and pattern of biogeographic shifts in sedentary organisms and benthic macroalgae are complicated by the influence of local dynamics and topographic features (islands, channels, coastal lagoons, e.g., of the Mediterranean (Bianchi, 2007), coastal upwelling e.g., Lima *et al.* (2007)). Geographical barriers constrain range shifts and may cause a loss of endemic species (Ben Rais Lasram *et al.*, 2010), with associated niches filled by alien species, either naturally migrating or artificially introduced (Philippart *et al.*, 2011).

Whether marine species can continue to keep pace as warming rates, hence climate velocities, increase (Fig MB-3b) is a key uncertainty. Climate velocities on land are expected to outpace the ability of many terrestrial species to track climate velocities this century (4.3.2.5, Figure 4.6) For marine species, the observed rates of shift are generally much faster than those land for land species, particularly for primary producers and lower trophic levels (Poloczanska *et al.* 2013). Phyto- and zooplankton communities (excluding larval fish) have extended distributions at remarkable rates (Figure MB-3b), such as in the North-east Atlantic (30.5.1) with implications for marine food webs.

Geographical range shifts and depth distribution vary between coexisting marine species (Genner *et al.*, 2004; Perry *et al.*, 2005; Simpson *et al.*, 2011) as a consequence of species-specific thermal window widths and associated vulnerabilities (Figure 6.5). Warming therefore causes differential changes in growth, reproductive success, larval output, early juvenile survival, and recruitment, implying shifts in the relative performance of animal species and, thus, their competitiveness (Pörtner and Farrell, 2008; Figure 6.7A). Such effects may underlie abundance losses or local extinctions, “regime shifts” between coexisting species, or critical mismatches between predator and prey organisms. Changes in local and regional species richness, abundance, community composition, productivity, energy flows and invasion resistance result. Even among Antarctic stenotherms such differences related to mode of life, phylogeny and associated metabolic capacities exist (6.3.1.4). As a consequence, marine ecosystem functions may be substantially reorganized at the regional scale, potentially triggering a range of cascading effects (Hoegh-Guldberg and Bruno, 2010). A focus on understanding the mechanisms underpinning the nature and magnitude of responses of marine organisms to climate change can help forecast impacts and the associated costs to society and facilitate adaptive management strategies effective in mitigating these impacts (6.3, 6.4).

[INSERT FIGURE MB-3 HERE

Figure MB-3. A. Rate of climate change for the Ocean (sea surface temperature (SST) °C); B. corresponding climate velocities for the Ocean and median velocity from land (adapted from Burrows et al., 2011); and C. observed rates of displacement of marine taxonomic groups over several decades until 2010. The thin dotted red arrows give an example of interpretation. Rates of climate change of 0.008 °C yr⁻¹ correspond to ca. 2.4 km yr⁻¹ median climate velocity in the Ocean. When compared to observed rates of displacement, many marine taxonomic groups have been able to track these velocities, except phyto- and zooplankton where rates of displacement greatly exceed climate velocity. All values are calculated for ocean surface with the exclusion of polar seas (Figure 30-1a). (A) Observed rates of climate change for Ocean SST (Black dotted line) are derived from HadISST1.1 data set, all other rates are calculated based on the average of the CMIP5 climate model ensembles (Table S30-3) for the historical period and for the future based on the four RCP emissions scenarios. Data were smoothed using a 20-year sliding window. (B) Median climate velocity calculated from HadISST1.1 dataset over 1960–2010 using the methods of Burrows et al., 2011. The three axes represent estimated median climate velocities are representative of areas of slow velocities such as Pacific subtropical gyre (STG) system (Purple line), the global Ocean surface (excluding polar seas, Blue line), and areas of high velocities such as the Coral Triangle and North Sea (Orange line). Figure 30-3 shows climate velocities over the ocean surface calculated over 1960–2010. The Red line corresponds to the median rate over global land surface calculated using historical surface temperatures from the CMIP5 model ensemble (Table S30-3). (C) Rates of displacement for marine taxonomic groups estimated by Poloczanska et al. 2013 using published studies (Figure MB-2 Black data set). Note the displacement rates for phytoplankton exceed the axis, so values are given.]

Box CC-MB References

- Ben Rais Lasram**, F.B., F. Guilhaumon, C. Albouy, S. Somot, W. Thuiller and D. Mouillot, 2010: The Mediterranean Sea as a ‘cul-de-sac’ for endemic fishes facing climate change, *Global Change Biology*, **16**, 3233-3245.
- Bianchi**, C.N., 2007: Biodiversity issues for the forthcoming Mediterranean Sea, *Hydrobiologia*, **580**, 7-21.
- Burrows**, M.T., D. S. Schoeman, L.B. Buckley, P.J. Moore, E.S. Poloczanska, K. Brander, K. C.J. Brown, J.F. Bruno, C.M. Duarte, B.S. Halpern, J. Holding, C.V. Kappel, W. Kiessling, M.I. O’Connor, J.M. Pandolfi, C. Parmesan, F. Schwing, W.J. Sydeman and A.J. Richardson, 2011: The pace of shifting climate in marine and terrestrial ecosystems, *Science*, **334**, 652-655.
- Cheung**, W.W.L., J.L. Sarmiento, J. Dunne, T.L. Frölicher, V. Lam, M.L.D. Palomares, R. Watson and D. Pauly, 2013: Shrinking of fishes exacerbates impacts of global ocean changes on marine ecosystems. *Nature Climate Change* **3**, 254-258.
- Genner**, M.J., D.W. Sims, V.J. Wearmouth, E.J. Southall, A.J. Southward, P.A. Henderson and S.J. Hawkins, 2004: Regional climatic warming drives long-term community changes of British marine fish. *Proceedings of the Royal Society of London B: Biological Sciences*, **271**(1539), 655-661.
- Hiddink**, J. G., and R. ter Hofstede (2008), Climate induced increases in species richness of marine fishes, *Global Change Biology*, **14**, 453-460.
- Hoegh-Guldberg**, O. and J.F. Bruno, 2010: The impact of climate change on the World’s marine ecosystems, *Science*, **328**, 1523-1528.
- Lima**, F.P., P.A. Ribeiro, N. Queiroz, S.J. Hawkins and A.M. Santos, 2007: Do distributional shifts of northern and southern species of algae match the warming pattern? *Global Change Biology*, **13**, 2592-2604.
- Perry**, A.L., P.J. Low, J.R. Ellis and J.D. Reynolds, 2005: Climate change and distribution shifts in marine fishes. *Science*, **308**(5730), 1912-1915.
- Philippart**, C.J.M., R. Anadon, R. Danovaro, J.W. Dippner, K.F. Drinkwater, S.J. Hawkins, T. Oguz, G. O’Sullivan and P.C. Reid, 2011: Impacts of climate change on European marine ecosystems: observations, expectations and indicators, *Journal of Experimental Marine Biology and Ecology*, **400**, 52-69.
- Pinsky**, M.L., B. Worm, M.J. Fogarty, J.L. Sarmiento, and S.A. Levin, 2013 Marine taxa track local climate velocities. *Science* **341**, 1239-1242.
- Pörtner**, H.O. and A.P. Farrell, 2008: Ecology: Physiology and climate change. *Science*, **322**(5902), 690-692.
- Poloczanska**, E.S., C.J. Brown, W.J. Sydeman, W. Kiessling, D.S. Schoeman, P.J. Moore, K. Brander, J.F. Bruno, L.B. Buckley, M.T. Burrows, C.M. Duarte, B.S. Halpern, J. Holding, C.V. Kappel, M.I. O’Connor, J.M. Pandolfi, C. Parmesan, F. Schwing, S.A. Thompson and A.J. Richardson, 2013: Global imprint of climate change on marine life, *Nature Climate Change*, published online 4 August 2013, doi: 10.1038/NCLIMATE1958, 7 pp.
- Simpson**, S.D., S. Jennings, M.P. Johnson, J.L. Blanchard, P.J. Schon, D.W. Sims and M.J. Genner, 2011: Continental shelf-wide response of a fish assemblage to rapid warming of the sea, *Current Biology*, **21**, 1565-1570.

Box CC-OA. Ocean Acidification

[Jean-Pierre Gattuso (France), Peter Brewer (USA), Ove Hoegh-Guldberg (Australia), Joan A. Kleypas (USA), Hans-Otto Pörtner (Germany), Daniela Schmidt (UK)]

Anthropogenic ocean acidification and global warming share the same primary cause, which is the increase of atmospheric CO₂ (Figure OA-1A; WGI, 2.2.1). Eutrophication, loss of sea ice, upwelling and deposition of atmospheric nitrogen and sulphur all exacerbate ocean acidification locally (5.3.3.6, 6.1.1, 30.3.2.2).

[INSERT FIGURE OA-1 HERE

Figure OA-1: A: Overview of the chemical, biological, socio-economic impacts of ocean acidification and of policy options (adapted from Turley and Gattuso, 2012). B: Multi-model simulated time series of global mean ocean surface pH (on the total scale) from CMIP5 climate model simulations from 1850 to 2100. Projections are shown for emission scenarios RCP2.6 (blue) and RCP8.5 (red) for the multi-model mean (solid lines) and range across the distribution of individual model simulations (shading). Black (grey shading) is the modelled historical evolution using historical reconstructed forcings. The models that are included are those from CMIP5 that simulate the global carbon cycle while being driven by prescribed atmospheric CO₂ concentrations. The number of CMIP5 models to calculate the multi-model mean is indicated for each time period/scenario (WGI AR5 Figure 6.28). C: Effect of near future acidification (seawater pH reduction of 0.5 unit or less) on major response variables estimated using weighted random effects meta-analyses, with the exception of survival which is not weighted (Kroeker et al., 2013). The log-transformed response ratio (LnRR) is the ratio of the mean effect in the acidification treatment to the mean effect in a control group. It indicates which process is most uniformly affected by ocean acidification but large variability exists between species. Significance is determined when the 95% bootstrapped confidence interval does not cross zero. The number of experiments used in the analyses is shown in parentheses. * denotes a statistically significant effect.]

Chemistry and Projections

The fundamental chemistry of ocean acidification is well understood (*robust evidence, high agreement*). Increasing atmospheric concentrations of CO₂ result in an increased flux of CO₂ into a mildly alkaline ocean, resulting in a reduction in pH, carbonate ion concentration, and the capacity of seawater to buffer changes in its chemistry (*very high confidence*). The changing chemistry of the surface layers of the open ocean can be projected at the global scale with high accuracy using projections of atmospheric CO₂ levels (Fig. CC-OA-1B). Observations of changing upper ocean CO₂ chemistry over time support this linkage (WGI Table 3.2 and Figure 3.18; Figures 30.8, 30.9). Projected changes in open ocean, surface water chemistry for year 2100 based on representative concentration pathways (WGI, Figure 6.28) compared to preindustrial values range from a pH change of -0.14 unit with RCP 2.6 (421 ppm CO₂, +1 °C, 22% reduction of carbonate ion concentration) to a pH change of -0.43 unit with RCP 8.5 (936 ppm CO₂, +3.7 °C, 56% reduction of carbonate ion concentration). Projections of regional changes, especially in the highly complex coastal systems (5.3.3.6, 30.3.2.2), in polar regions (WGI 6.4.4), and at depth are more difficult but generally follow similar trends.

Biological, Ecological, and Biogeochemical Impacts

Investigations of the effect of ocean acidification on marine organisms and ecosystems have a relatively short history, recently analyzed in several metaanalyses (6.3.2.1, 6.3.5.1). A wide range of sensitivities to projected rates of ocean acidification exists within and across diverse groups of organisms, with a trend for greater sensitivity in early life stages (*high confidence*; 5.4.2.2, 5.4.2.4, 6.3.2). A pattern of positive and negative impacts emerges (*high confidence*; Fig. OA-1C) but key uncertainties remain in our understanding of the impacts on organisms, life histories and ecosystems. Responses can be influenced, often exacerbated by other drivers, such as warming, hypoxia, nutrient concentration, and light availability (*high confidence*; 5.4.2.4, 6.3.5).

Growth and primary production are stimulated in seagrass and some phytoplankton (*high confidence*; 5.4.2.3, 6.3.2.2-3, 30.5.6). Harmful algal blooms could become more frequent (*limited evidence, medium agreement*). Ocean acidification may stimulate nitrogen fixation (*limited evidence, low agreement*; 6.3.2.2). It decreases the rate of calcification of most, but not all, sea-floor calcifiers (*medium agreement, robust evidence*) such as reef-building corals (Box CC-CR), coralline algae, bivalves and gastropods reducing the competitiveness with non-calcifiers

(5.4.2.2, 5.4.2.4, 6.3.2.5). Ocean warming and acidification promote higher rates of calcium carbonate dissolution resulting in the net dissolution of carbonate sediments and frameworks and loss of associated habitat (*medium confidence*; 5.4.2.4, 6.3.2.5, 6.3.5.4-5). Some corals and temperate fishes experience disturbances to behavior, navigation and their ability to tell conspecifics from predators (6.3.2.4). However, there is no evidence for these effects to persist on evolutionary timescales in the few groups analyzed (6.3.2).

Some phytoplankton and mollusks displayed adaptation to ocean acidification in long-term experiments (*limited evidence, medium agreement*; 6.3.2.1), indicating that the long-term responses could be less than responses obtained in short-term experiments. However, mass extinctions in Earth history occurred during much slower rates of ocean acidification, combined with other drivers changing, suggesting that evolutionary rates are not fast enough for sensitive animals and plants to adapt to the projected rate of future change (*medium confidence*; 6.1.2).

Projections of ocean acidification effects at ecosystem level are made difficult by the diversity of species-level responses. Differential sensitivities and associated shifts in performance and distribution will change predator-prey relationships and competitive interactions (6.3.2.5, 6.3.5-6), which could impact food webs and higher trophic levels (*limited evidence, high agreement*). Natural analogues at CO₂ vents indicate decreased species diversity, biomass and trophic complexity of communities (Box CC-CR; 5.4.2.3, 6.3.2.5, 30.3.2.2, 30.5). Shifts in community structure have also been documented in regions with rapidly declining pH (5.4.2.2).

Due to an incomplete understanding of species-specific responses and trophic interactions the effect of ocean acidification on global biogeochemical cycles is not well understood (*limited evidence, low agreement*) and represents an important knowledge gap. The additive, synergistic or antagonistic interactions of factors such as temperature, concentrations of oxygen and nutrients, and light are not sufficiently investigated yet.

Risks, Socioeconomic Impacts and Costs

The risks of ocean acidification to marine organisms, ecosystems, and ultimately to human societies, include both the probability that ocean acidification will affect fundamental physiological and ecological processes of organisms (6.3.2.1), and the magnitude of the resulting impacts on ecosystems and the ecosystem services they provide to society (Box 19-2). For example, ocean acidification under RCP4.5 to RCP8.5 will impact formation and maintenance of coral reefs (*high confidence*; Box CC-CR, 5.4.2.4) and the goods and services that they provide such as fisheries, tourism and coastal protection (*limited evidence, high agreement*; Box CC-CR, 6.4.1.1, 19.5.2, 27.3.3, 30.5, 30.6). Ocean acidification poses many other potential risks, but these cannot yet be quantitatively assessed due to the small number of studies available, particularly on the magnitude of the ecological and socioeconomic impacts (19.5.2).

Global estimates of observed or projected economic costs of ocean acidification do not exist. The largest uncertainty is how the impacts on lower trophic levels will propagate through the food webs and to top predators. However, there are a number of instructive examples that illustrate the magnitude of potential impacts of ocean acidification. A decrease of the production of commercially-exploited shelled mollusks (6.4.1.1) would result in a reduction of US production of 3 to 13% according to the SRES A1FI emission scenario (*low confidence*). The global cost of production loss of mollusks could be over 100 billion USD by 2100 (*limited evidence, medium agreement*). Models suggest that ocean acidification will generally reduce fish biomass and catch (*low confidence*) and that complex additive, antagonistic and/or synergistic interactions will occur with other environmental (warming) and human (fisheries management) factors (6.4.1.1). The annual economic damage of ocean-acidification-induced coral reef loss by 2100 has been estimated, in 2009, to be 870 and 528 billion USD, respectively for the A1 and B2 SRES emission scenarios (*low confidence*; 6.4.1). Although this number is small compared to global GDP, it can represent a very large GDP loss for the economies of many coastal regions or small islands that rely on the ecological goods and services of coral reefs (25.7.5, 29.3.1.2).

Mitigation and Adaptation

Successful management of the impacts of ocean acidification includes two approaches: mitigation of the source of the problem (i.e. reduce anthropogenic emissions of CO₂), and/or adaptation by reducing the consequences of past and future ocean acidification (6.4.2.1). Mitigation of ocean acidification through reduction of atmospheric CO₂ is

the most effective and the least risky method to limit ocean acidification and its impacts (6.4.2.1). Climate geoengineering techniques based on solar radiation management will not abate ocean acidification and could increase it under some circumstances (6.4.2.2). Geoengineering techniques to remove carbon dioxide from the atmosphere could directly address the problem but are very costly and may be limited by the lack of CO₂ storage capacity (6.4.2.2). Additionally, some ocean-based approaches, such as iron fertilization, would only re-locate ocean acidification from the upper ocean to the ocean interior, with potential ramifications on deep-water oxygen levels (6.4.2.2; 30.3.2.3 and 30.5.7). A low-regret approach, with relatively limited effectiveness, is to limit the number and the magnitude of drivers other than CO₂, such as nutrient pollution (6.4.2.1). Mitigation of ocean acidification at the local level could involve the reduction of anthropogenic inputs of nutrients and organic matter in the coastal ocean (5.3.4.2). Some adaptation strategies include drawing water for aquaculture from local watersheds only when pH is in the right range, selecting for less sensitive species or strains, or relocating industries elsewhere (6.4.2.1).

CC-OA References

- Kroeker K., R.C. Kordas, A. Ryan, I. Hendriks, L.Ramajo, G. Singh, C. Duarte and J.-P. Gattuso, 2013: Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. *Global Change Biology* **19**, 1884-1896.
- Turley C. and J.-P. Gattuso, 2012. Future biological and ecosystem impacts of ocean acidification and their socioeconomic-policy implications. *Current Opinion In Environmental Sustainability* **4**, 278-286.

Box CC-PP. Net Primary Production in the Ocean

[Philip W. Boyd (New Zealand), Svein Sundby (Norway), Hans-Otto Pörtner (Germany)]

Net Primary Production (NPP) is the rate of photosynthetic carbon fixation minus the fraction of fixed carbon used for cellular respiration and maintenance by autotrophic planktonic microbes and benthic plants (6.2.1, 6.3.1). Environmental drivers of NPP include light, nutrients, micronutrients, carbon dioxide, and temperature (Panel A). These drivers in turn, are influenced by oceanic and atmospheric processes, including cloud cover, sea-ice extent, mixing by winds, waves and currents, convection, density stratification, and various forms of upwelling induced by eddies, frontal activity and boundary currents. Temperature has multiple roles as it influences rates of phytoplankton physiology and heterotrophic bacterial recycling of nutrients, in addition to stratification of the water column and sea-ice extent (Panel A). Climate change is projected to strongly impact NPP through a multitude of ways that depend on the regional and local physical settings (WGI, Ch. 3), and on ecosystem structure and functioning (*medium confidence*, 6.3.4, 6.5.1). The influence of environmental drivers on NPP causes as much as a 10-fold variation in regional productivity: from <50 g C m⁻² year⁻¹ in nutrient-poor subtropical waters and light-limited Arctic waters to >> 300 g C m⁻² year⁻¹ in productive upwelling regions and highly eutrophic coastal regions (Panel B).

The oceans currently provide ~50 x 10¹⁵ g C year⁻¹, or about half of global NPP (Field *et al.* 1998). Global estimates of NPP are mainly obtained from satellite remote-sensing (6.1.2), which provides unprecedented spatial and temporal coverage, and may be validated regionally against oceanic measurements. Observations reveal significant changes in rates of NPP when environmental controls are altered by episodic natural perturbations, such as volcanic eruptions enhancing iron supply, as observed in high-nitrate low-chlorophyll waters of the NE Pacific (Hamme *et al.*, 2010). Climate variability can drive pronounced changes in NPP (Chavez *et al.*, 2011), such as during El Niño to La Niña transitions in Equatorial Pacific, when vertical nutrient and trace element supply are enhanced (Chavez *et al.*, 1999).

Multi-year time-series records of NPP have been used to assess spatial trends in NPP in recent decades. Behrenfeld *et al.* (2006) using satellite data, reported a prolonged and sustained global NPP decrease of 190 x 10¹² g C year⁻¹, for the period 1999 to 2005 - an annual reduction of ~0.4 % of global NPP. In contrast, a time-series of directly measured NPP between 1988 to 2007 by Saba *et al.* (2010) (i.e. *in situ* incubations using the radiotracer ¹⁴C-bicarbonate) revealed an increase (2 % year⁻¹) in NPP for two low latitude open ocean sites. This discrepancy between *in situ* and remotely-sensed NPP trends points to uncertainties in either the methodology used and/or the extent to which discrete sites are representative of oceanic provinces (Saba *et al.*, 2010, 2011). Modeling studies

have subsequently revealed that the <15 year archive of satellite-derived NPP is insufficient to distinguish climate-change mediated shifts in NPP from those driven by natural climate variability (Henson *et al.*, 2010; Beaulieu *et al.*, 2013). Although multidecadal, the available time-series of oceanic NPP measurements are also not of sufficient duration relative to the timescales of climate variability modes (up to 60-70 years for AMO, for example, Figure 6-1). Recent attempts to synthesize longer (i.e. centennial) records of chlorophyll as a proxy for phytoplankton stocks (e.g., Boyce *et al.*, 2010) have been criticized for relying on questionable linkages between different proxies for chlorophyll over a century of records (e.g., Rykaczewski and Dunne, 2011).

Models in which projected climate-change alters the environmental drivers of NPP provide estimates of spatial changes and of the rate of change of NPP. For example, four global coupled climate-ocean biogeochemical Earth System Models (WGI Ch. 6) projected an increase in NPP at high latitudes as a result of alleviation of light and temperature limitation of NPP particularly in Northern and Southern Hemisphere 'subpolar gyre' biomes (Steinacher *et al.*, 2010). However, this regional increase in NPP was more than offset by decreases in NPP at lower latitudes and at mid-latitudes due to the reduced input of macro-nutrients into the photic zone. The reduced mixed-layer depth and reduced rate of circulation may cause a decrease in the flux of macronutrients to the photic zone (Figure 6-2). These changes to oceanic conditions result in a reduction in global mean NPP by 2 to 13% by 2100 relative to 1860 under a high emission scenario (Polovina *et al.*, 2011; SRES A2, between RCP6.0 and RCP8.5). This is consistent with a more recent analysis based on 10 Earth System Models (Bopp *et al.*, 2013), which project decreases in global NPP by 8.6 (± 7.9), 3.9 (± 5.7), 3.6 (± 5.7), 2.0 (± 4.1) % in the 2090s relative to the 1990s, under the scenarios RCP8.5, RCP6.0, RCP4.5 and RCP2.6, respectively. However, the magnitude of projected changes varies widely between models (e.g. from 0 to 20% decrease in NPP globally under RCP 8.5). The various models show very large differences in NPP at regional (i.e. provinces, see panel B) scales.

Earlier model projections had predicted changes in global NPP from a decrease of > 10% (Field *et al.*, 1998; Boyd and Doney, 2002) to an increase of up to 8.1% under an intermediate scenario (SRES A1B, similar to RCP6.0) (Sarmiento *et al.*, 2004; Schmittner *et al.*, 2008). These projections did not consider the potential contribution of primary production derived from atmospheric nitrogen fixation in tropical and subtropical regions, favoured by increasing stratification and reduced nutrient inputs from mixing. This mechanism is potentially important, although such episodic increases in nitrogen fixation are not sustainable without the presence of excess phosphate (e.g. Moore *et al.*, 2009; Boyd *et al.*, 2010). This may lead to an underestimation of NPP (Mohr *et al.*, 2010; Mulholland *et al.*, 2012; Wilson *et al.*, 2012), however, the extent of such underestimation is unknown (Luo *et al.*, 2012).

Care must be taken when comparing global, provincial (e.g. low latitude waters, for example Behrenfeld *et al.*, 2006) and regional trends in NPP derived from observations, as some regions have additional local environmental influences such as enhanced density stratification of the upper ocean from melting sea ice. For example, a longer phytoplankton growing season, due to more sea-ice free days, may have increased NPP (based on a regionally validated time-series of satellite NPP) in Arctic waters (Arrigo and van Dijken, 2011) by an average of 8.1 Tg C year⁻¹ between 1998 and 2009. Other regional trends in NPP are reported in 30.5.1-6. In addition, although future model projections of global NPP from different models (Steinacher *et al.*, 2010; Bopp *et al.*, 2013) are comparable, regional projections from each of the models differ substantially. This raises concerns as to which aspect(s) of the different model NPP parameterizations are responsible for driving regional differences in NPP, and moreover, how accurate model projections are of global NPP.

From a global perspective, open ocean NPP will decrease moderately by 2100 under both low (SRES B1 or RCP4.5) and high emission scenarios (A2 or RCP6.0 - 8.5, 6.3.4, 6.5.1, *medium confidence*), paralleled by an increase in NPP at high latitudes and a decrease in the tropics (*medium confidence*). However, there is *limited evidence* and *low agreement* on the direction, magnitude and differences of a change of NPP in various ocean regions and coastal waters projected by 2100 (*low confidence*).

[INSERT FIGURE PP-1 HERE]

Figure PP-1: A) Environmental factors controlling Net Primary Production (NPP). NPP is mainly controlled by three basic processes: 1) Light conditions in the surface ocean, i.e. the photic zone where photosynthesis occurs, 2) upward flux of nutrients and micronutrients from underlying waters into the photic zone, 3) Regeneration of nutrients and micronutrients via the breakdown and recycling of organic material before it sinks out of the photic

zone. All three processes are influenced by physical, chemical and biological processes and vary across regional ecosystems. In addition, water temperature strongly influences the upper rate of photosynthesis for cells that are resource-replete. Predictions of alteration of primary productivity under climate change depend on correct parameterizations and simulations of each of these variables and processes for each region. B) Annual composite map of global areal NPP rates (derived from MODIS Aqua satellite climatology from 2003–2012; NPP was calculated with the Carbon-based Production Model (CbPM, Westberry *et al.*, 2008)). Overlaid is a grid of (thin black lines) that represent 51 distinct global ocean biogeographical provinces (after Longhurst, 1998 and based on Boyd and Doney, 2002). The characteristics and boundaries of each province are primarily set by the underlying regional ocean physics and chemistry. Figure courtesy of Toby Westberry (OSU) and Ivan Lima (WHOI), satellite data courtesy of NASA Ocean Biology Processing Group.]

Box CC-PP References

- Arrigo, K.R.** and G.L. van Dijken, 2011: Secular trends in Arctic Ocean net primary production. *Journal of Geophysical Research*, **116(C9)**, C09011.
- Beaulieu, C., S.A. Henson, J.L. Sarmiento, J.P. Dunne, S.C. Doney, R.R. Rykaczewski and L. Bopp**, 2013: Factors challenging our ability to detect long-term trends in ocean chlorophyll. *Biogeosciences*, **10(4)**, 2711-2724.
- Behrenfeld, M.J., R.T. O'Malley, D.A. Siegel, C.R. McClain, J.L. Sarmiento, G.C. Feldman, A.J. Milligan, P.G. Falkowski, R.M. Letelier and E.S. Boss**, 2006: Climate-driven trends in contemporary ocean productivity. *Nature*, **444(7120)**, 752-755.
- Bopp, L., L. Resplandy, J.D. Orr, D. S.C., J.P. Dunne, M. Gehlen, P. Halloran, C. Heinze, T. Ilyina, R. Séférian, J. Tjiputra and M. Vichi**, 2013: Multiple stressors of ocean ecosystems in the 21st century: projections with CMIP5 models. *Biogeosciences*, **10(10)**, 6225-6245.
- Boyce, D.G., M.R. Lewis and B. Worm**, 2010: Global phytoplankton decline over the past century. *Nature*, **466(7306)**, 591-596.
- Boyd, P.W.** and S.C. Doney, 2002: Modelling regional responses by marine pelagic ecosystems to global climate change. *Geophysical Research Letters*, **29(16)**, 1806.
- Boyd, P.W., R. Strzepek, F.X. Fu and D.A. Hutchins**, 2010: Environmental control of open-ocean phytoplankton groups: now and in the future. *Limnology and Oceanography*, **55(3)**, 1353-1376.
- Chavez, F.P., M. Messie and J.T. Pennington**, 2011: Marine primary production in relation to climate variability and change. *Annual Review of Marine Science*, **3(1)**, 227-260.
- Chavez, F.P., P.G. Strutton, C.E. Friederich, R.A. Feely, G.C. Feldman, D.C. Foley and M.J. McPhaden**, 1999: Biological and chemical response of the equatorial Pacific Ocean to the 1997-98 El Niño. *Science*, **286(5447)**, 2126-2131.
- Field, C.B., M.J. Behrenfeld, J.T. Randerson and P. Falkowski**, 1998: Primary production of the biosphere: integrating terrestrial and oceanic components. *Science*, **281(5374)**, 237-240.
- Hamme, R.C., P.W. Webley, W.R. Crawford, F.A. Whitney, M.D. DeGrandpre, S.R. Emerson, C.C. Eriksen, K.E. Giesbrecht, J.F.R. Gower, M.T. Kavanaugh, M.A. Peña, C.L. Sabine, S.D. Batten, L.A. Coogan, D.S. Grundle and D. Lockwood**, 2010: Volcanic ash fuels anomalous plankton bloom in subarctic northeast Pacific. *Geophysical Research Letters*, **37**, L19604.
- Henson, S.A., J.L. Sarmiento, J.P. Dunne, L. Bopp, I. Lima, S.C. Doney, J. John and C. Beaulieu**, 2010: Detection of anthropogenic climate change in satellite records of ocean chlorophyll and productivity. *Biogeosciences*, **7(2)**, 621-640.
- Longhurst, A.R.**, 1998: *Ecological Geography of the Sea*. Academic Press, San Diego, CA, USA, 560 pp.
- Luo, Y.-W., S.C. Doney, L.A. Anderson, M. Benavides, I. Berman-Frank, A. Bode, S. Bonnet, K.H. Boström, D. Böttjer, D.G. Capone, E.J. Carpenter, Y.L. Chen, M.J. Church, J.E. Dore, L.I. Falcón, A. Fernández, R.A. Foster, K. Furuya, F. Gómez, K. Gundersen, A.M. Hynes, D.M. Karl, S. Kitajima, R.J. Langlois, J. LaRoche, R.M. Letelier, E. Marañón, D.J. McGillicuddy Jr., P.H. Moisander, C.M. Moore, B. Mouriño-Carballido, M.R. Mulholland, J.A. Needoba, K.M. Orcutt, A.J. Poulton, E. Rahav, P. Raimbault, A.P. Rees, L. Riemann, T. Shiozaki, A. Subramaniam, T. Tyrrell, K.A. Turk-Kubo, M. Varela, T.A. Villareal, E.A. Webb, A.E. White, J. Wu and J.P. Zehr**, 2012: Database of diazotrophs in global ocean: abundances, biomass and nitrogen fixation rates. *Earth System Science Data*, **5**, 47-106.
- Mohr, W., T. Grosskopf, D.W.R. Wallace and J. LaRoche**, 2010: Methodological underestimation of oceanic nitrogen fixation rates. *PLoS ONE*, **5(9)**, e12583.
- Moore, C.M., M.M. Mills, E.P. Achterberg, R.J. Geider, J. LaRoche, M.I. Lucas, E.L. McDonagh, X. Pan, A.J. Poulton, M.J.A. Rijkenberg, D.J. Suggett, S.J. Ussher and E.M.S. Woodward**, 2009: Large-scale distribution of Atlantic nitrogen fixation controlled by iron availability. *Nature Geoscience*, **2(12)**, 867-871.
- Mulholland, M.R., P.W. Bernhardt, J.L. Blanco-Garcia, A. Mannino, K. Hyde, E. Mondragon, K. Turk, P.H. Moisander and J.P. Zehr**, 2012: Rates of dinitrogen fixation and the abundance of diazotrophs in North American coastal waters between Cape Hatteras and Georges Bank. *Limnology and Oceanography*, **57(4)**, 1067-1083.

- Polovina, J.J., J.P. Dunne, P.A. Woodworth and E.A. Howell, 2011:** Projected expansion of the subtropical biome and contraction of the temperate and equatorial upwelling biomes in the North Pacific under global warming. *ICES Journal of Marine Science*, **68(6)**, 986-995.
- Rykaczewski, R.R. and J.P. Dunne, 2011:** A measured look at ocean chlorophyll trends. *Nature*, **472(7342)**, E5-E6.
- Saba, V.S., M.A.M. Friedrichs, D. Antoine, R.A. Armstrong, I. Asanuma, M.J. Behrenfeld, A.M. Ciotti, M. Dowell, N. Hoepffner, K.J.W. Hyde, J. Ishizaka, T. Kameda, J. Marra, F. Mélin, A. Morel, J. O'Reilly, M. Scardi, W.O. Smith Jr., T.J. Smyth, S. Tang, J. Uitz, K. Waters and T.K. Westberry, 2011:** An evaluation of ocean color model estimates of marine primary productivity in coastal and pelagic regions across the globe. *Biogeosciences*, **8(2)**, 489-503.
- Saba, V.S., M.A.M. Friedrichs, M.-E. Carr, D. Antoine, R.A. Armstrong, I. Asanuma, O. Aumont, N.R. Bates, M.J. Behrenfeld, V. Bennington, L. Bopp, J. Bruggeman, E.T. Buitenhuis, M.J. Church, A.M. Ciotti, S.C. Doney, M. Dowell, J. Dunne, S. Dutkiewicz, W. Gregg, N. Hoepffner, K.J.W. Hyde, J. Ishizaka, T. Kameda, D.M. Karl, I. Lima, M.W. Lomas, J. Marra, G.A. McKinley, F. Mélin, J.K. Moore, A. Morel, J. O'Reilly, B. Salihoglu, M. Scardi, T.J. Smyth, S.L. Tang, J. Tjiputra, J. Uitz, M. Vichi, K. Waters, T.K. Westberry and A. Yool, 2010:** Challenges of modeling depth-integrated marine primary productivity over multiple decades: a case study at BATS and HOT. *Global Biogeochemical Cycles*, **24**, GB3020.
- Sarmiento, J.L., R. Slater, R. Barber, L. Bopp, S.C. Doney, A.C. Hirst, J. Kleypas, R. Matear, U. Mikolajewicz, P. Monfray, V. Soldatov, S.A. Spall and R. Stouffer, 2004:** Response of ocean ecosystems to climate warming. *Global Biogeochemical Cycles*, **18(3)**, GB3003.
- Schmittner, A., A. Oschlies, H.D. Matthews and E.D. Galbraith, 2008:** Future changes in climate, ocean circulation, ecosystems, and biogeochemical cycling simulated for a business-as-usual CO₂ emission scenario until year 4000 AD. *Global Biogeochemical Cycles*, **22(1)**, GB1013.
- Steinacher, M., F. Joos, T.L. Frölicher, L. Bopp, P. Cadule, V. Cocco, S.C. Doney, M. Gehlen, K. Lindsay, J.K. Moore, B. Schneider and J. Segsneider, 2010:** Projected 21st century decrease in marine productivity: a multi-model analysis. *Biogeosciences*, **7(3)**, 979-1005.
- Westberry, T., Behrenfeld, M.J., Siegel, D.A., and Boss, E. 2008.** Carbon-based primary productivity modeling with vertically resolved photoacclimation. *Global Biogeochemical Cycles*, **22(2)**: GB2024. DOI: 10.1029/2007GB003078
- Wilson, S.T., D. Bottjer, M.J. Church and D.M. Karl, 2012:** Comparative assessment of nitrogen fixation methodologies, conducted in the oligotrophic North Pacific Ocean. *Applied and Environmental Microbiology*, **78(18)**, 6516-6523.

Box CC-UP. Uncertain Trends in Major Upwelling Ecosystems

[Salvador E. Lluch-Cota (Mexico), Ove Hoegh-Guldberg (Australia), David Karl (USA), Hans O. Pörtner (Germany), Svein Sundby (Norway), Jean-Pierre Gattuso (France)]

Upwelling is the vertical transport of cold, dense, nutrient-rich, relatively low-pH and often oxygen-poor waters to the euphotic zone where light is abundant. These waters trigger high levels of primary production and a high biomass of benthic and pelagic organisms. The driving forces of upwelling include wind stress and the interaction of ocean currents with bottom topography. Upwelling intensity also depends on water column stratification. The major upwelling systems of the Planet, the Equatorial Upwelling System (EUS, 30.5.2, Figure 30.1A) and the Eastern Boundary Upwelling Ecosystems (EBUE, 30.5.5, Figure 30.1A), represent only 10% of the ocean surface but contribute nearly 25 % to global fish production (Figure 30.1B, Table S30.1).

Marine ecosystems associated with upwelling systems can be influenced by a range of 'bottom-up' trophic mechanisms, with upwelling, transport, and chlorophyll concentrations showing strong seasonal and interannual couplings and variability. These, in turn, influence trophic transfer up the food chain, affecting zooplankton, foraging fish, seabirds and marine mammals.

There is considerable speculation as to how upwelling systems might change in a warming and acidifying ocean. Globally, the heat gain of the surface ocean has increased stratification by 4% (WGI 3.2, 3.4.4, 3.8), which means that more wind energy is needed to bring deep waters to the surface. It is as yet unclear to what extent wind stress can offset the increased stratification, due to the uncertainty in wind speed trends (WGI, 3.4.4). In the tropics, observations of reductions in trade winds over several decades contrast more recent evidence indicating their strengthening since the early 1990s (WGI, 9.4.1.3.4). Observations and modelling efforts in fact show diverging trends in coastal upwelling at the eastern boundaries of the Pacific and the Atlantic. Bakun (1990) proposed that the the difference in heat gaining rates between land and ocean causes an increase in the pressure gradient, which results in increased alongshore winds and leads to intensified offshore transport of surface water through Ekman pumping, and the upwelling of nutrient rich, cold waters (Figure CC-UP). Some regional records support this hypothesis, others do not. There is considerable variability in warming and cooling trends over the past decades both within and

among systems making it difficult to predict changes in the intensity of all Eastern Boundary Upwelling Ecosystems (30.5.5).

Understanding whether upwelling and climate change will impact resident biota in an additive, synergistic or antagonistic manner is important for projections of how ecological goods and services provided for human society will change. Even though upwellings may prove more resilient to climate change than other ocean ecosystems because of their ability to function under extremely variable conditions (Capone and Hutchins, 2013), consequences of their shifts are highly relevant since these are the most biologically active systems in the ocean. Increased upwelling would enhance fisheries yields. However, the export of organic material from surface to deeper layers of the ocean may increase and stimulate its decomposition by microbial activity, thereby enhancing oxygen depletion and CO₂ enrichment in deeper water layers. Once this water returns to the surface through upwelling benthic and pelagic coastal communities will be exposed to acidified and deoxygenated water which may combine with anthropogenic impact to negatively affect marine biota and ecosystem structure of the upper ocean (high confidence, 6.3.2, 6.3.3; 30.3.2.2, 30.3.2.3). Extreme hypoxia may result in abnormal mortalities of fishes and invertebrates (Keller *et al.*, 2010), reduce the fisheries catch potential and impact aquaculture in coastal areas (5.4.3.3, 6.3.7, 30.5.1.1.2, 30.5.5.1.3, Barton *et al.*, 2012). Shifts in upwelling also coincide with an apparent increase in the frequency of submarine eruptions of methane and hydrogen sulphide gas, caused by enhanced formation and sinking of phytoplankton biomass to the hypoxic or anoxic sea floor. This combination of factors has been implicated in the extensive mortality of coastal fishes and invertebrates (Bakun and Weeks, 2004), resulting in significant reductions in fishing productivity, such as Cape hake (*Merluccius capensis*), Namibia's most valuable fishery (Hamukuaya *et al.*, 1998).

Reduced upwelling would also reduce the productivity of important pelagic fisheries, such as for sardines, anchovies and mackerel, with major consequences for the economies of several countries (6.4.1, Chp 7, Figure 30.1A, B, Table S30.1). However, under projected scenarios of reduced upward supply of nutrients due to stratification of the open ocean, upwelling of both nutrients and trace elements may become increasingly important to maintaining upper ocean nutrient and trace metal inventories. It has been suggested that upwelling areas may also increase nutrient content and productivity under enhanced stratification, and that upwelled and partially denitrified waters containing excess phosphate may select for N₂-fixing microorganisms (Deutsch *et al.*, 2007; Deutsch and Weber, 2012), but field observations of N₂ fixation in these regions have not supported these predictions (Fernandez *et al.*, 2011; Franz *et al.*, 2012). The role of this process in global primary production thus needs to be validated (*low confidence*).

The central question therefore is whether or not upwelling will intensify, and if so, whether the effects of intensified upwelling on O₂ and CO₂ inventories will outweigh its benefits for primary production and associated fisheries and aquaculture (*low confidence*). In any case increasing atmospheric CO₂ concentrations will equilibrate with upwelling waters that may cause them to become more corrosive, depending upon pCO₂ of the upwelled water, and potentially increasingly impact the biota of Eastern Boundary Upwelling Ecosystems.

[INSERT FIGURE UP-1 HERE]

Figure UP-1: Upper panel: Schematic hypothetical mechanism of increasing coastal wind-driven upwelling at eastern boundary systems, where differential warming rates between land and ocean results in increased land-ocean pressure gradients (1) that produce stronger alongshore winds (2) and offshore movement of surface water through Ekman transport (3), and increased upwelling of deep cold nutrient rich waters to replace it (4). Lower panel: potential consequences of climate change in upwelling systems. Increasing stratification and uncertainty in wind stress trends result in uncertain trends in upwelling. Increasing upwelling may result in higher input of nutrients to the euphotic zone, and increased primary production, which in turn may enhance pelagic fisheries, but also decreased coastal fisheries due to an augmented exposure of coastal fauna to hypoxic, low pH waters. Decreased upwelling may result in lower primary production in these systems with direct impacts on pelagic fisheries productivity.]

Box CC-UP References

- Bakun, A.**, 1990: Global climate change and intensification of coastal ocean upwelling, *Science*, **247(4939)**, 198-201.
- Bakun, A.** and S.J. Weeks, 2004: Greenhouse gas buildup, sardines, submarine eruptions and the possibility of abrupt degradation of intense marine upwelling ecosystems. *Ecology Letters*, **7(11)**, 1015-1023.
- Barton, A.**, B. Hales, G.G. Waldbusser, C. Langdon, R.A. Feely, 2012: The Pacific oyster, *Crassostrea gigas*, shows negative correlation to naturally elevated carbon dioxide levels: Implications for near-term ocean acidification effects, *Limnology and Oceanography*, **57(3)**: 698-710.
- Capone, D.G.** and D.A. Hutchins, 2013: Microbial biogeochemistry of coastal upwelling regimes in a changing ocean. *Nature geoscience*, 711-717.
- Deutsch, C.** and T. Weber, 2012: Nutrient ratios as a tracer and driver of ocean biogeochemistry. *Annual Review of Marine Science*, **4**, 113-114.
- Deutsch, C.**, J.L. Sarmiento, D.M. Sigman, N. Gruber and J.P. Dunne, 2007: Spatial coupling of nitrogen inputs and losses in the ocean. *Nature*, **445(7124)**, 163-167.
- Fernandez, C.**, L. Farías and O. Ulloa, 2011: Nitrogen fixation in denitrified marine waters. *PLoS ONE*, **6(6)**, e20539.
- Franz, J.**, G. Krahnemann, G. Lavik, P. Grasse, T. Dittmar and U. Riebesell, 2012: Dynamics and stoichiometry of nutrients and phytoplankton in waters influenced by the oxygen minimum zone in the eastern tropical Pacific. *Deep Sea Research Part I: Oceanographic Research Papers*, **62**, 20-31.
- Hamukuaya, H.**, M.J. O'Toole and P.M.J. Woodhead, 1998: Observations of severe hypoxia and offshore displacement of Cape hake over the Namibian shelf in 1994. *South African Journal of Marine Science*, **19(1)**, 57-59.
- Keller, AA.**, Simon V, Chan F, Wakefield WW, Clarke ME, et al., 2010: Demersal fish and invertebrate biomass in relation to an offshore hypoxic zone along the US West Coast. *Fisheries Oceanography* **19**:76–87.

Chapter References

- Abed, R.M.**, F. Garcia-Pichel and M. Hernandez-Marine, 2002: Polyphasic characterization of benthic, moderately halophilic, moderately thermophilic cyanobacteria with very thin trichomes and the proposal of *Halomicronema excentricum* gen. nov., sp. nov. *Archives of Microbiology*, **177(5)**, 361-370.
- Adger, W.N.**, T.P. Hughes, C. Folke, S.R. Carpenter and J. Rockström, 2005: Social-ecological resilience to coastal disasters. *Science*, **309(5737)**, 1036-1039.
- Ainsworth, C.H.**, J.F. Samhuri, D.S. Busch, W.W.L. Cheung, J. Dunne and T.A. Okey, 2011: Potential impacts of climate change on Northeast Pacific marine foodwebs and fisheries. *ICES Journal of Marine Science*, **68(6)**, 1217-1229.
- Alheit, J.**, T. Pohlmann, M. Casini, W. Greve, R. Hinrichs, M. Mathis, K. O'Driscoll, R. Vorberg and C. Wagner, 2012: Climate variability drives anchovies and sardines into North Sea and Baltic Sea. *Progress in Oceanography*, **96(1)**, 128-139.
- Alker, A.P.**, G.W. Smith and K. Kim, 2001: Characterization of *Aspergillus sydowii* (Thom et Church), a fungal pathogen of Caribbean sea fan corals. *Hydrobiologia*, **460**, 105-111.
- Allison, E.H.**, A.L. Perry, M.-C. Badjeck, W.N. Adger, K. Brown, D. Conway, A.S. Halls, G.M. Pilling, J.D. Reynolds, N.L. Andrew and N.K. Dulvy, 2009: Vulnerability of national economies to the impacts of climate change on fisheries. *Fish and Fisheries*, **10(2)**, 173-196.
- Altizer, S.**, R.S. Ostfeld, P.T.J. Johnson, S. Kutz and C.D. Harvell, 2013: Climate change and infectious diseases: from evidence to a predictive framework. *Science*, **341(6145)**, 514-519.
- Amelung, B.**, S. Nicholls and D. Viner, 2007: Implications of global climate change for tourism flows and seasonality. *Journal of Travel Research*, **45(3)**, 285-296.
- Andersson, A.J.**, F.T. Mackenzie and J.-P. Gattuso, 2011: 7- Effects of ocean acidification on benthic processes, organisms, and ecosystems. In: *Ocean Acidification* [Gattuso, J.-P. and L. Hansson (eds.)]. Oxford University Press, Oxford, pp. 122-153.
- Angilletta, M.J.J.**, 2009: *Thermal Adaptation. A Theoretical and Empirical Synthesis* Oxford University Press, New York, 320 pp.
- Anthony, K.R.**, D.I. Kline, G. Diaz-Pulido, S. Dove and O. Hoegh-Guldberg, 2008: Ocean acidification causes bleaching and productivity loss in coral reef builders. *Proceedings of the National Academy of Sciences of the United States of America*, **105(45)**, 17442-17446.

- Archer, D.**, M. Eby, V. Brovkin, A. Ridgwell, L. Cao, U. Mikolajewicz, K. Caldeira, K. Matsumoto, G. Munhoven, A. Montenegro and K. Tokos, 2009: Atmospheric lifetime of fossil fuel carbon dioxide. *Annual Review of Earth and Planetary Sciences*, **37(1)**, 117-134.
- Armbrust, E.V.**, 2009: The life of diatoms in the world's oceans. *Nature*, **459(7244)**, 185-192.
- Armstrong, J.L.**, J.L. Boldt, A.D. Cross, J.H. Moss, N.D. Davis, K.W. Myers, R.V. Walker, D.A. Beauchamp and L.J. Halderson, 2005: Distribution, size, and interannual, seasonal and diel food habits of northern Gulf of Alaska juvenile pink salmon, *Oncorhynchus gorbuscha*. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **52(1-2)**, 247-265.
- Arnason, R.**, 2007: Climate change and fisheries: assessing the economic impact in Iceland and Greenland. *Natural Resource Modeling*, **20(2)**, 163-197.
- Arnold, K.E.**, H.S. Findlay, J.I. Spicer, C.L. Daniels and D. Boothroyd, 2009: Effect of CO₂-related acidification on aspects of the larval development of the European lobster, *Homarus gammarus* (L.). *Biogeosciences*, **6(8)**, 1747-1754.
- Arnold, T.**, C. Mealey, H. Leahey, A.W. Miller, J.M. Hall-Spencer, M. Milazzo and K. Maers, 2012: Ocean acidification and the loss of phenolic substances in marine plants. *PLoS ONE*, **7(4)**, e35107.
- Arrieta, J.M.**, S. Arnaud-Haond and C.M. Duarte, 2010: What lies underneath: conserving the oceans' genetic resources. *Proceedings of the National Academy of Sciences of the United States of America*, **107(43)**, 18318-18324.
- Arrigo, K.R.** and G.L. van Dijken, 2011: Secular trends in Arctic Ocean net primary production. *Journal of Geophysical Research*, **116(C9)**, C09011.
- Arrigo, K.R.**, D.K. Perovich, R.S. Pickart, Z.W. Brown, G.L. van Dijken, K.E. Lowry, M.M. Mills, M.A. Palmer, W.M. Balch, F. Bahr, N.R. Bates, C. Benitez-Nelson, B. Bowler, E. Brownlee, J.K. Ehn, K.E. Frey, R. Garley, S.R. Laney, L. Lubelczyk, J. Mathis, A. Matsuoka, B.G. Mitchell, G.W.K. Moore, E. Ortega-Retuerta, S. Pal, C.M. Polashenski, R.A. Reynolds, B. Schieber, H.M. Sosik, M. Stephens and J.H. Swift, 2012: Massive phytoplankton blooms under Arctic sea ice. *Science*, **336(6087)**, 1408.
- Auel, H.**, W. Hagen, W. Ekau and H.M. Verheye, 2005: Metabolic adaptations and reduced respiration of the copepod *Calanoides carinatus* during diapause at depth in the Angola-Benguela Front and northern Benguela upwelling regions. *African Journal of Marine Science*, **27(3)**, 653-657.
- Baker, A.C.**, 2001: Ecosystems: reef corals bleach to survive change. *Nature*, **411(6839)**, 765-766.
- Baker-Austin, C.**, J.A. Trinanes, N.G.H. Taylor, R. Hartnell, A. Siitonen and J. Martinez-Urtaza, 2013: Emerging *Vibrio* risk at high latitudes in response to ocean warming. *Nature Climate Change*, **3(1)**, 73-77.
- Balazs, G.H.** and M. Chaloupka, 2004: Thirty-year recovery trend in the once depleted Hawaiian green sea turtle stock. *Biological Conservation*, **117(5)**, 491-498.
- Banse, K.**, 1991: Rates of phytoplankton cell division in the field and in iron enrichment experiments. *Limnology and Oceanography*, **36(8)**, 1886-1898.
- Barange, M.**, W.W.L. Cheung, G. Merino and R.I. Perry, 2010: Modelling the potential impacts of climate change and human activities on the sustainability of marine resources. *Current Opinion in Environmental Sustainability*, **2(5-6)**, 326-333.
- Barker, S.** and H. Elderfield, 2002: Foraminiferal calcification response to glacial-interglacial changes in atmospheric CO₂. *Science*, **297(5582)**, 833-836.
- Barker, S.**, P. Diz, M.J. Vautravers, J. Pike, G. Knorr, I.R. Hall and W.S. Broecker, 2009: Interhemispheric Atlantic seesaw response during the last deglaciation. *Nature*, **457(7233)**, 1097-1102.
- Barton, A.**, B. Hales, G.G. Waldbusser, C. Langdon and R.A. Feely, 2012: The Pacific oyster, *Crassostrea gigas*, shows negative correlation to naturally elevated carbon dioxide levels: Implications for near-term ocean acidification effects. *Limnology and Oceanography*, **57(3)**, 698-710.
- Baumann, H.**, S.C. Talmage and C.J. Gobler, 2012: Reduced early life growth and survival in a fish in direct response to increased carbon dioxide. *Nature Climate Change*, **2(1)**, 38-41.
- Baumgartner, M.**, K.O. Stetter and W. Foissner, 2002: Morphological, small subunit rRNA, and physiological characterization of *Trimyema minutum* (Kahl, 1931), an anaerobic ciliate from submarine hydrothermal vents growing from 28°C to 52°C. *Journal of Eukaryotic Microbiology*, **49(3)**, 227-238.
- Bazzino, G.**, W.F. Gilly, U. Markaida, C.A. Salinas-Zavala and J. Ramos-Castillejos, 2010: Horizontal movements, vertical-habitat utilization and diet of the jumbo squid (*Dosidicus gigas*) in the Pacific Ocean off Baja California Sur, Mexico. *Progress in Oceanography*, **86(1-2)**, 59-71.

- Beare, D.**, F. Burns, E. Jones, K. Peach, E. Portilla, T. Greig, E. McKenzie and D. Reid, 2004: An increase in the abundance of anchovies and sardines in the north-western North Sea since 1995. *Global Change Biology*, **10(7)**, 1209-1213.
- Beaufort, L.**, I. Probert, T. de Garidel-Thoron, E.M. Bendif, D. Ruiz-Pino, N. Metzl, C. Goyet, N. Buchet, P. Coupel, M. Grelaud, B. Rost, R.E.M. Rickaby and C. de Vargas, 2011: Sensitivity of coccolithophores to carbonate chemistry and ocean acidification. *Nature*, **476(7358)**, 80-83.
- Beaugrand, G.**, 2009: Decadal changes in climate and ecosystems in the North Atlantic Ocean and adjacent seas. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **56(8-10)**, 656-673.
- Beaugrand, G.**, C. Luczak and M. Edwards, 2009: Rapid biogeographical plankton shifts in the North Atlantic Ocean. *Global Change Biology*, **15(7)**, 1790-1803.
- Beaugrand, G.**, M. Edwards and L. Legendre, 2010: Marine biodiversity, ecosystem functioning, and carbon cycles. *Proceedings of the National Academy of Sciences of the United States of America*, **107(22)**, 10120-10124.
- Beaugrand, G.**, A. McQuatters-Gollop, M. Edwards and E. Goberville, 2013: Long-term responses of North Atlantic calcifying plankton to climate change. *Nature Climate Change*, **3(3)**, 263-267.
- Beaugrand, G.**, P.C. Reid, F. Ibañez, J.A. Lindley and M. Edwards, 2002: Reorganization of North Atlantic marine copepod biodiversity and climate. *Science*, **296(5573)**, 1692-1694.
- Beaugrand, G.**, M. Edwards, K. Brander, C. Luczak and F. Ibañez, 2008: Causes and projections of abrupt climate-driven ecosystem shifts in the North Atlantic. *Ecology Letters*, **11(11)**, 1157-1168.
- Beaulieu, C.**, S.A. Henson, J.L. Sarmiento, J.P. Dunne, S.C. Doney, R.R. Rykaczewski and L. Bopp, 2013: Factors challenging our ability to detect long-term trends in ocean chlorophyll. *Biogeosciences*, **10(4)**, 2711-2724.
- Beaumont, N.J.**, M.C. Austen, J.P. Atkins, D. Burdon, S. Degraer, T.P. Dentinho, S. Derous, P. Holm, T. Horton, E. van Ierland, A.H. Marboe, D.J. Starkey, M. Townsend, T. Zarzycki, 2007: Identification, definition and quantification of goods and services provided by marine biodiversity: implications for the ecosystem approach. *Marine Pollution Bulletin* **54(3)**, 253-265.
- Becker, B.H.**, M.Z. Peery and S.R. Beissinger, 2007: Ocean climate and prey availability affect the trophic level and reproductive success of the marbled murrelet, an endangered seabird. *Marine Ecology Progress Series*, **329**, 267-279.
- Bednaršek, N.**, G.A. Tarling, D.C.E. Bakker, S. Fielding, E.M. Jones, H.J. Venables, P. Ward, A. Kuzirian, B. Lézé, R.A. Feely and E.J. Murphy, 2012: Extensive dissolution of live pteropods in the Southern Ocean. *Nature Geoscience*, **5(12)**, 881-885.
- Behrenfeld, M.**, 2011: Uncertain future for ocean algae. *Nature Climate Change*, **1(1)**, 33-34.
- Beman, J.M.**, C.-E. Chow, A.L. King, Y. Feng, J.A. Fuhrman, A. Andersson, N.R. Bates, B.N. Popp and D.A. Hutchins, 2011: Global declines in oceanic nitrification rates as a consequence of ocean acidification. *Proceedings of the National Academy of Sciences of the United States of America*, **108(1)**, 208-213.
- Beniash, E.**, A. Ivanina, N.S. Lieb, I. Kurochkin and I.M. Sokolova, 2010: Elevated level of carbon dioxide affects metabolism and shell formation in oysters *Crassostrea virginica*. *Marine Ecology Progress Series*, **419**, 95-108.
- Benson, S.**, P. Cook, J. Anderson, S. Bachu, H.B. Nimir, B. Basu, J. Bradshaw, G. Deguchi, J. Gale, G. von Goerne, W. Heidug, S. Holloway, R. Kamal, D. Keith, P. Lloyd, P. Rocha, B. Senior, J. Thomson, T. Torp, T. Wildenborg, M. Wilson, F. Zarlenga and D. Zhou, 2005: Underground geological storage. In: *Carbon Dioxide Capture and Storage: A Special Report of IPCC Working Group III* [Metz, B., O. Davidson, H. de Coninck, M. Loos and L. Meyer (eds.)]. Cambridge University Press, Cambridge, pp. 195-276.
- Bertrand, A.**, M. Ballón and A. Chaigneau, 2010: Acoustic observation of living organisms reveals the upper limit of the oxygen minimum zone. *PLoS ONE*, **5(4)**, e10330.
- Bertrand, E.M.**, M.A. Saito, J.M. Rose, C.R. Riesselman, M.C. Lohan, A.E. Noble, P.A. Lee and G.R. DiTullio, 2007: Vitamin B-12 and iron colimitation of phytoplankton growth in the Ross Sea. *Limnology and Oceanography*, **52(3)**, 1079-1093.
- Bianchi, D.**, E.D. Galbraith, D.A. Carozza, K.A.S. Mislán and C.A. Stock, 2013: Intensification of open-ocean oxygen depletion by vertically migrating animals. *Nature Geoscience*, **6(7)**, 545-548.
- Billé, R.**, R. Kelly, E. Harrould-Kolieb, D. Herr, F. Joos, K.J. Kroeker, D. Laffoley, A. Oschlies and J.-P. Gattuso, 2013: Taking action against ocean acidification: a review of management and policy options. *Environmental Management*, **52**, 761-779.
- Bissinger, J.E.**, D.J.S. Montagnes, J. Sharples and D. Atkinson, 2008: Predicting marine phytoplankton maximum growth rates from temperature: improving on the Eppley curve using quantile regression *Limnology and Oceanography*, **53(2)**, 487-493.

- Blanchard, J.L.**, S. Jennings, R. Holmes, J. Harle, G. Merino, J.I. Allen, J. Holt, N.K. Dulvy and M. Barange, 2012: Potential consequences of climate change for primary production and fish production in large marine ecosystems. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **367(1605)**, 2979-2989.
- Bode, A.**, J.A. Hare, W.K.W. Li, X.A.G. Morán and L. Valdés, 2011: Chlorophyll and primary production in the North Atlantic. In: *ICES Cooperative Research Report No. 310* [Reid, P.C. and L. Valdés (eds.)]. International Council for the Exploration of the Sea, pp. 77-102.
- Boetius, A.**, S. Albrecht, K. Bakker, C. Bienhold, J. Felden, M. Fernández-Méndez, S. Hendricks, C. Katlein, C. Lalande, T. Krumpfen, M. Nicolaus, I. Peeken, B. Rabe, A. Rogacheva, E. Rybakova, R. Somavilla, F. Wenzhöfer and RV Polarstern ARK27-3-Shipboard Science Party, 2013: Export of algal biomass from the melting arctic sea ice. *Science*, **339(6126)**, 1430-1432.
- Bolton, J.J.** and K. Lüning, 1982: Optimal growth and maximal survival temperatures of Atlantic *Laminaria* species (Phaeophyta) in culture. *Marine Biology*, **66(1)**, 89-94.
- Bopp, L.**, C. Le Quéré, M. Heimann, A.C. Manning and P. Monfray, 2002: Climate-induced oceanic oxygen fluxes: Implications for the contemporary carbon budget. *Global Biogeochemical Cycles*, **16(2)**, 1022.
- Bopp, L.**, L. Resplandy, J.D. Orr, D. S.C., J.P. Dunne, M. Gehlen, P. Halloran, C. Heinze, T. Ilyina, R. Sférian, J. Tijiputra and M. Vichi, 2013: Multiple stressors of ocean ecosystems in the 21st century: projections with CMIP5 models. *Biogeosciences*, **10(10)**, 6225-6245.
- Bossdorf, O.**, C.L. Richards and M. Pigliucci, 2008: Epigenetics for ecologists. *Ecology Letters*, **11(2)**, 106-115.
- Botsford, L.W.**, M.D. Holland, J.F. Samhouri, J.W. White and A. Hastings, 2011: Importance of age structure in models of the response of upper trophic levels to fishing and climate change. *ICES Journal of Marine Science*, **68(6)**, 1270-1283.
- Bowler, C.**, A. Vardi and A.E. Allen, 2010: Oceanographic and biogeochemical insights from diatom genomes. *Annual Review of Marine Science*, **2(1)**, 333-365.
- Bown, P.R.**, J.A. Lees and J.R. Young, 2004: Calcareous nannoplankton evolution and diversity through time. In: *Coccolithophores - From Molecular Processes to Global Impact* [Thierstein, H.R. and J.R. Young (eds.)]. Springer, Heidelberg, pp. 481-508.
- Boyce, D.G.**, M.R. Lewis and B. Worm, 2010: Global phytoplankton decline over the past century. *Nature*, **466(7306)**, 591-596.
- Boyd, P.W.**, 2002: Environmental factors controlling phytoplankton processes in the Southern Ocean. *Journal of Phycology*, **38(5)**, 844-861.
- Boyd, P.W.**, 2008: Ranking geo-engineering schemes. *Nature Geoscience*, **1(11)**, 722-724.
- Boyd, P.W.**, 2011: Beyond ocean acidification. *Nature Geoscience*, **4(5)**, 273-274.
- Boyd, P.W.** and S.C. Doney, 2002: Modelling regional responses by marine pelagic ecosystems to global climate change. *Geophysical Research Letters*, **29(16)**, 1806.
- Boyd, P.W.** and D.A. Hutchins, 2012: Understanding the responses of ocean biota to a complex matrix of cumulative anthropogenic change. *Marine Ecology Progress Series*, **470**, 125-135.
- Boyd, P.W.**, C.S. Law and S.C. Doney, 2011: A climate change atlas for the ocean. *Oceanography*, **24(2)**, 13-16.
- Boyd, P.W.**, R. Strzepek, F.X. Fu and D.A. Hutchins, 2010: Environmental control of open-ocean phytoplankton groups: now and in the future. *Limnology and Oceanography*, **55(3)**, 1353-1376.
- Boyd, P.W.**, T. Jickells, C.S. Law, S. Blain, E.A. Boyle, K.O. Buesseler, K.H. Coale, J.J. Cullen, H.J. de Baar, M. Follows, M. Harvey, C. Lancelot, M. Levasseur, N.P. Owens, R. Pollard, R.B. Rivkin, J. Sarmiento, V. Schoemann, V. Smetacek, S. Takeda, A. Tsuda, S. Turner and A.J. Watson, 2007: Mesoscale iron enrichment experiments 1993-2005: synthesis and future directions. *Science*, **315(5812)**, 612-617.
- Bradshaw, W.E.** and C.M. Holzapfel, 2010: Light, time, and the physiology of biotic response to rapid climate change in animals. *Annual Review of Physiology*, **72(1)**, 147-166.
- Brainard, R.E.**, C. Birkeland, C.M. Eakin, P. McElhany, M.W. Miller, M. Patterson and G.A. Piniak, 2011: *Status review report of 82 candidate coral species petitioned under the U.S. Endangered Species Act*. U.S. Department of Commerce, NOAA Technical Memorandum, 530 pp.
- Bralower, T.J.**, 2002: Evidence of surface water oligotrophy during the Paleocene-Eocene thermal maximum: nanofossil assemblage data from Ocean Drilling Program Site 690, Maud Rise, Weddell Sea. *Paleoceanography*, **17(2)**, 1-15.
- Branch, T.A.**, B.M. DeJoseph, L.J. Ray and C.A. Wagner, 2013: Impacts of ocean acidification on marine seafood. *Trends in Ecology and Evolution*, **28(3)**, 178-186.

- Brander, K.**, 2008: Tackling the old familiar problems of pollution, habitat alteration and overfishing will help with adapting to climate change. *Marine Pollution Bulletin*, **56(12)**, 1957-1958.
- Brander, K., G. Blom, M.F. Borges, K. Erzini, G. Henderson, B.R. MacKenzie, H. Mendes, J. Ribeiro, A.M.P. Santos and R. Toresen**, 2003: Changes in fish distribution in the eastern North Atlantic: are we seeing a coherent response to changing temperature? *ICES Marine Science Symposia*, **219**, 261-270.
- Brander, K.M.**, 2007: Global fish production and climate change. *Proceedings of the National Academy of Sciences of the United States of America*, **104(50)**, 19709-19714.
- Brander, L.M., K. Rehdanz, R.S.J. Tol and P.J.H. Van Beukering**, 2012: The economic impact of ocean acidification on coral reefs. *Climate Change Economics*, **3(1)**, 1250002.
- Brandes, J.A., A.H. Devol and C. Deutsch**, 2007: New developments in the marine nitrogen cycle. *Chemical Reviews*, **107(2)**, 577-589.
- Braun-McNeill, J., C.R. Sasso, S.P. Epperly and C. Rivero**, 2008: Feasibility of using sea surface temperature imagery to mitigate cheloniid sea turtle-fishery interactions off the coast of northeastern USA. *Endangered Species Research*, **5(2-3)**, 257-266.
- Breau, C., R.A. Cunjak and S.J. Peake**, 2011: Behaviour during elevated water temperatures: can physiology explain movement of juvenile Atlantic salmon to cool water? *Journal of Animal Ecology*, **80(4)**, 844-853.
- Brewer, P.G. and E.T. Peltzer**, 2009: Limits to marine life. *Science*, **324(5925)**, 347-348.
- Brinton, E. and A. Townsend**, 2003: Decadal variability in abundances of the dominant euphausiid species in southern sectors of the California Current. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **50(14-16)**, 2449-2472.
- Broderick, A.C., B.J. Godley, S. Reece and J.R. Downie**, 2000: Incubation periods and sex ratios of green turtles: highly female biased hatchling production in the eastern Mediterranean. *Marine Ecology Progress Series*, **202**, 273-281.
- Brotz, L., W.W.L. Cheung, K. Kleisner, E. Pakhomov and D. Pauly**, 2012: Increasing jellyfish populations: trends in large marine ecosystems. *Hydrobiologia*, **690(1)**, 3-20.
- Brown, C.J., E.A. Fulton, A.J. Hobday, R.J. Matear, H.P. Possingham, C. Bulman, V. Christensen, R.E. Forrest, P.C. Gehrke, N.A. Gribble, S.P. Griffiths, H. Lozano-Montes, J.M. Martin, S. Metcalf, T.A. Okey, R. Watson and A.J. Richardson**, 2010: Effects of climate-driven primary production change on marine food webs: implications for fisheries and conservation. *Global Change Biology*, **16(4)**, 1194-1212.
- Brunel, T. and M. Dickey-Collas**, 2010: Effects of temperature and population density on von Bertalanffy growth parameters in Atlantic herring: a macro-ecological analysis. *Marine Ecology Progress Series*, **405**, 15-28.
- Bruno, J.F. and E.R. Selig**, 2007: Regional decline of coral cover in the Indo-Pacific: timing, extent, and subregional comparisons. *PLoS ONE*, **2(8)**, e711.
- Buesseler, K.O., S.C. Doney, D.M. Karl, P.W. Boyd, K. Caldeira, F. Chai, K.H. Coale, H.J. de Baar, P.G. Falkowski, K.S. Johnson, R.S. Lampitt, A.F. Michaels, S.W.A. Naqvi, V. Smetacek, S. Takeda and A.J. Watson**, 2008: Ocean iron fertilization - moving forward in a sea of uncertainty. *Science*, **319**, 162.
- Bunce, A., F. Norman, N. Brothers and R. Gales**, 2002: Long-term trends in the Australasian gannet (*Morus serrator*) population in Australia: the effect of climate change and commercial fisheries. *Marine Biology*, **141(2)**, 263-269.
- Burge, C.A., C.M. Eakin, C.S. Friedman, B. Froelich, P.K. Hershberger, E.E. Hofmann, L.E. Petes, K.C. Prager, E. Weil, B.L. Willis, S.E. Ford and C.D. Harvell**, 2013 in press: Climate change influences on marine infectious diseases: implications for management and society. *Annual Review of Marine Science*.
- Burleson, M.L. and P.E. Silva**, 2011: Cross tolerance to environmental stressors: effects of hypoxic acclimation on cardiovascular responses of channel catfish (*Ictalurus punctatus*) to a thermal challenge. *Journal of Thermal Biology*, **36(4)**, 250-254.
- Burrows, M.T., D.S. Schoeman, L.B. Buckley, P. Moore, E.S. Poloczanska, K.M. Brander, C. Brown, J.F. Bruno, C.M. Duarte, B.S. Halpern, J. Holding, C.V. Kappel, W. Kiessling, M.I. O'Connor, J.M. Pandolfi, C. Parmesan, F.B. Schwing, W.J. Sydeman and A.J. Richardson**, 2011: The pace of shifting climate in marine and terrestrial ecosystems. *Science*, **334(6056)**, 652-655.
- Cai, W.-J., X. Hu, W.-J. Huang, M.C. Murrell, J.C. Lehrter, S.E. Lohrenz, W.-C. Chou, W. Zhai, J.T. Hollibaugh, Y. Wang, P. Zhao, X. Guo, K. Gundersen, M. Dai and G.-C. Gong**, 2011: Acidification of subsurface coastal waters enhanced by eutrophication. *Nature Geoscience*, **4(11)**, 766-770.
- Cairns, D.K., A.J. Gaston and F. Huettmann**, 2008: Endothermy, ectothermy and the global structure of marine vertebrate communities. *Marine Ecology Progress Series*, **356**, 239-250.

- Calambokidis, J.**, J. Barlow, J.K.B. Ford, T.E. Chandler and A.B. Douglas, 2009: Insights into the population structure of blue whales in the Eastern North Pacific from recent sightings and photographic identification. *Marine Mammal Science*, **25(4)**, 816-832.
- Caldeira, K.** and L. Wood, 2008: Global and Arctic climate engineering: numerical model studies. *Philosophical Transactions of the Royal Society A: Mathematical, Physical, and Engineering Sciences*, **366(1882)**, 4039-4056.
- Caldeira, K.**, M. Akai, P. Brewer, B. Chen, P. Haugan, T. Iwama, P. Johnston, H. Kheshgi, Q. Li, T. Ohsumi, H.-O. Pörtner, C. Sabine, Y. Shirayama and J. Thomson, 2005: Ocean Storage. In: *Carbon Dioxide Capture and Storage: A Special Report of IPCC Working Group III* [Metz, B. and O. Davidson (eds.)]. Cambridge University Press, Cambridge UK, pp. 277-318.
- Calosi, P.**, S.P.S. Rastrick, M. Graziano, S.C. Thomas, C. Baggini, H.A. Carter, J.M. Hall-Spencer, M. Milazzo and J.I. Spicer, 2013: Distribution of sea urchins living near shallow water CO₂ vents is dependent upon species acid-base and ion-regulatory abilities. *Marine Pollution Bulletin*, **73(2)**, 470-484.
- Campbell, J.**, D. Antoine, R. Armstrong, K. Arrigo, W. Balch, R. Barber, M. Behrenfeld, R. Bidigare, J. Bishop, M.-E. Carr, W. Esaias, P. Falkowski, N. Hoepffner, R. Iverson, D. Kiefer, S. Lohrenz, J. Marra, A. Morel, J. Ryan, V. Vedernikov, K. Waters, C. Yentsch and J. Yoder, 2002: Comparison of algorithms for estimating ocean primary production from surface chlorophyll, temperature, and irradiance. *Global Biogeochemical Cycles*, **16(3)**, 1035.
- Campbell, S.J.**, L.J. McKenzie and S.P. Kerville, 2006: Photosynthetic responses of seven tropical seagrasses to elevated seawater temperature. *Journal of Experimental Marine Biology and Ecology*, **330(2)**, 455-468.
- Cao, L.** and K. Caldeira, 2010: Can ocean iron fertilization mitigate ocean acidification? *Climatic Change*, **99(1-2)**, 303-311.
- Capotondi, A.**, M.A. Alexander, N.A. Bond, E.N. Curchitser and J.D. Scott, 2012: Enhanced upper ocean stratification with climate change in the CMIP3 models. *Journal of Geophysical Research*, **117(C4)**, C04031.
- Carlton, J.T.**, 2000: Global change and biological invasions in the oceans. In: *Invasive Species in a Changing World* [Mooney, H.A. and R.J. Hobbs (eds.)]. Island Press, Covelo, CA, pp. 31-53.
- Carpenter, S.R.** and W.A. Brock, 2006: Rising variance: a leading indicator of ecological transition. *Ecology Letters*, **9(3)**, 311-318.
- Carrillo, C.J.**, R.C. Smith and D.M. Karl, 2004: Processes regulating oxygen and carbon dioxide in surface waters west of the Antarctic Peninsula. *Marine Chemistry*, **84(3-4)**, 161-179.
- Casini, M.**, J. Hjelm, J.C. Molinero, J. Lövgren, M. Cardinale, V. Bartolino, A. Belgrano and G. Kornilovs, 2009: Trophic cascades promote threshold-like shifts in pelagic marine ecosystems. *Proceedings of the National Academy of Sciences of the United States of America*, **106(1)**, 197-202.
- CBD**, 2009: *Scientific Synthesis of the Impacts of Ocean Acidification on Marine Biodiversity by the Secretariat of the Convention on Biological Diversity*. Technical Series No. 46, Montreal, Canada, 61 pp.
- Cesar, H.**, L. Burke and L. Pet-Soede, 2003: *The Economics of Worldwide Coral Reef Degradation*. Cesar Environmental Economics Consulting (CEEC), Arnhem, 23 pp.
- Chaloupka, M.**, N. Kamezaki and C. Limpus, 2008: Is climate change affecting the population dynamics of the endangered Pacific loggerhead sea turtle? *Journal of Experimental Marine Biology and Ecology*, **356(1-2)**, 136-143.
- Chambers, L.E.**, L. Hughes and M.A. Weston, 2005: Climate change and its impact on Australia's avifauna. *Emu*, **105(1)**, 1-20.
- Chambers, L.E.**, C.A. Devney, B.C. Congdon, N. Dunlop, E.J. Woehler and P. Dann, 2011: Observed and predicted effects of climate on Australian seabirds. *Emu*, **111(3)**, 235-251.
- Chan, F.**, J.A. Barth, J. Lubchenco, A. Kirincich, H. Weeks, W.T. Peterson and B.A. Menge, 2008: Emergence of anoxia in the California Current large marine ecosystem. *Science*, **319(5865)**, 920.
- Charpy-Roubaud, C.** and A. Sournia, 1990: The comparative estimation of phytoplanktonic, microphytobenthic and macrophytobenthic primary production in the oceans. *Marine Microbial Food Webs*, **4(1)**, 31-57.
- Chavez, F.P.**, M. Messie and J.T. Pennington, 2011: Marine primary production in relation to climate variability and change. *Annual Review of Marine Science*, **3(1)**, 227-260.
- Chavez, F.P.**, J. Ryan, S.E. Lluch-Cota and M. Niquen C, 2003: From anchovies to sardines and back: multidecadal change in the Pacific Ocean. *Science*, **299(5604)**, 217-221.
- Checkley Jr, D.M.**, A.G. Dickson, M. Takahashi, J.A. Radich, N. Eisenkolb and R. Asch, 2009: Elevated CO₂ enhances otolith growth in young fish. *Science*, **324(5935)**, 1683.

- Cheung, W.W.L., R. Watson and D. Pauly, 2013a:** Signature of ocean warming in global fisheries catches. *Nature*, **497**, 365-368.
- Cheung, W.W.L., J. Dunne, J.L. Sarmiento and D. Pauly, 2011:** Integrating ecophysiology and plankton dynamics into projected maximum fisheries catch potential under climate change in the Northeast Atlantic. *ICES Journal of Marine Science*, **68(6)**, 1008-1018.
- Cheung, W.W.L., C. Close, V. Lam, R. Watson and D. Pauly, 2008:** Application of macroecological theory to predict effects of climate change on global fisheries potential. *Marine Ecology Progress Series*, **365**, 187-197.
- Cheung, W.W.L., V.W.Y. Lam, J.L. Sarmiento, K. Kearney, R. Watson and D. Pauly, 2009:** Projecting global marine biodiversity impacts under climate change scenarios. *Fish and Fisheries*, **10(3)**, 235-251.
- Cheung, W.W.L., V.W.Y. Lam, J.L. Sarmiento, K. Kearney, R. Watson, D. Zeller and D. Pauly, 2010:** Large-scale redistribution of maximum fisheries catch in the global ocean under climate change. *Global Change Biology*, **16(1)**, 24-35.
- Cheung, W.W.L., J.L. Sarmiento, J. Dunne, T.L. Frölicher, V.W.Y. Lam, M.L.D. Palomares, R. Watson and D. Pauly, 2013b:** Shrinking of fishes exacerbates impacts of global ocean changes on marine ecosystems. *Nature Climate Change*, **3(3)**, 254-258.
- Chevaldonné, P., C. Fisher, J. Childress, D. Desbruyères, D. Jollivet, F. Zal and A. Toulmond, 2000:** Thermotolerance and the 'Pompeii worms'. *Marine Ecology Progress Series*, **208**, 2093-2295.
- Chevin, L.-M., R. Lande and G.M. Mace, 2010:** Adaptation, plasticity, and extinction in a changing environment: towards a predictive theory. *PLoS Biology*, **8(4)**, e1000357.
- Christen, N., P. Calosi, C.L. McNeill and S. Widdicombe, 2013:** Structural and functional vulnerability to elevated $p\text{CO}_2$ in marine benthic communities. *Marine Biology*, **160**, 2113-2128.
- Christian, J.R. and D.M. Karl, 1995:** Bacterial ectoenzymes in marine waters - activity ratios and temperature responses in 3 oceanographic provinces. *Limnology and Oceanography*, **40(6)**, 1042-1049.
- Claiborne, J.B., S.L. Edwards and A.I. Morrison-Shetlar, 2002:** Acid-base regulation in fishes: cellular and molecular mechanisms. *Journal of Experimental Zoology*, **293(3)**, 302-319.
- Clark, D., M. Lamare and M. Barker, 2009:** Response of sea urchin pluteus larvae (Echinodermata: Echinoidea) to reduced seawater pH: a comparison among a tropical, temperate, and a polar species. *Marine Biology*, **156(6)**, 1125-1137.
- CLIMAP Project Members, 1976:** The surface of the ice-age earth. *Science*, **191(4232)**, 1131-1137.
- Colbourne, E., J. Craig, C. Fitzpatrick, D. Senciall, P. Stead and W. Bailey, 2011:** *An assessment of the physical oceanographic environment on the Newfoundland and Labrador Shelf during 2010*. DFO Canadian Science Advisory Secretariat Science Advisory Report 2011/089, iv + 31p. pp.
- Coll, M., L.J. Shannon, D. Yemane, J.S. Link, H. Ojaveer, S. Neira, D. Jouffre, P. Labrosse, J.J. Heymans, E.A. Fulton and Y.-J. Shin, 2010:** Ranking the ecological relative status of exploited marine ecosystems. *ICES Journal of Marine Science*, **67(4)**, 769-786.
- Comeau, A.M., W.K.W. Li, J.E. Tremblay, E.C. Carmack and C. Lovejoy, 2011:** Arctic ocean microbial community structure before and after the 2007 record sea ice minimum. *PLoS ONE*, **6(11)**, e27492.
- Comeau, S., G. Gorsky, R. Jeffree, J.L. Teyssié and J.-P. Gattuso, 2009:** Impact of ocean acidification on a key Arctic pelagic mollusc (*Limacina helicina*). *Biogeosciences*, **6(9)**, 1877-1882.
- Condon, R.H., W.M. Graham, C.M. Duarte, K.A. Pitt, C.H. Lucas, S.H.D. Haddock, K.R. Sutherland, K.L. Robinson, M.N. Dawson, M.B. Decker, C.E. Mills, J.E. Purcell, A. Malej, H. Mianzan, S.-I. Uye, S. Gelcich and L.P. Madin, 2012:** Questioning the rise of gelatinous zooplankton in the world's oceans. *Bio Science*, **62(2)**, 160-169.
- Condon, R.H., C.M. Duarte, K.A. Pitt, K.L. Robinson, C.H. Lucas, K.R. Sutherland, H.W. Mianzan, M. Bogeberg, J.E. Purcell, M.B. Decker, S. Uye, L.P. Madin, R.D. Brodeur, S.H.D. Haddock, A. Malej, G.D. Parry, E. Eriksen, J. Quiñones, M. Acha, M. Harvey, J.M. Arthur and W.M. Graham, 2013:** Recurrent jellyfish blooms are a consequence of global oscillations. *Proceedings of the National Academy of Sciences of the United States of America*, **110(3)**, 1000-1005.
- Cooley, S.R., 2012:** How humans could "feel" changing biogeochemistry. *Current Opinion in Environmental Sustainability*, **4(3)**, 258-263.
- Cooley, S.R. and S.C. Doney, 2009:** Anticipating ocean acidification's economic consequences for commercial fisheries. *Environmental Research Letters*, **4(2)**, 024007.
- Cooley, S.R., H.L. Kite-Powell and S.C. Doney, 2009:** Ocean acidification's potential to alter global marine ecosystem services. *Oceanography*, **22(4)**, 172-181.

- Cooley, S.R., N. Lucey, H. Kite-Powell and S.C. Doney, 2012: Nutrition and income from molluscs today imply vulnerability to ocean acidification tomorrow. *Fish and Fisheries*, **13**(2), 182-215.
- Corbett, J.J., D.A. Lack, J.J. Winebrake, S. Harder, J.A. Silberman and M. Gold, 2010: Arctic shipping emissions inventories and future scenarios. *Atmospheric Chemistry and Physics*, **10**(19), 9689-9704.
- Costello, J.H., B.K. Sullivan and D.J. Gifford, 2006: A physical-biological interaction underlying variable phenological responses to climate change by coastal zooplankton. *Journal of Plankton Research*, **28**(11), 1099-1105.
- Crain, C.M., K. Kroeker and B.S. Halpern, 2008: Interactive and cumulative effects of multiple human stressors in marine systems. *Ecology Letters*, **11**(12), 1304-1315.
- Crook, E.D., D. Potts, M. Rebolledo-Vieyra, L. Hernandez and A. Paytan, 2012: Calcifying coral abundance near low-pH springs: implications for future ocean acidification. *Coral Reefs*, **31**(1), 239-245.
- Crutzen, P., 2006: Albedo enhancement by stratospheric sulfur injections: a contribution to resolve a policy dilemma? *Climatic Change*, **77**(3-4), 211-220.
- Cubillos, J.C., S.W. Wright, G. Nash, M.F. de Salas, B. Griffiths, B. Tilbrook, A. Poisson and G.M. Hallegraeff, 2007: Calcification morphotypes of the coccolithophorid *Emiliania huxleyi* in the Southern Ocean: Changes in 2001 to 2006 compared to historical data. *Marine Ecology Progress Series*, **348**, 47-54.
- Cuevas, E., F.A. Abreu-Grobois, V. Guzmán-Hernández, M.A. Liceaga-Correa and R.P. van Dam, 2008: Post-nesting migratory movements of hawksbill turtles *Eretmochelys imbricata* in waters adjacent to the Yucatan Peninsula, Mexico. *Endangered Species Research*, **10**, 123-133.
- Cui, Y., L.R. Kump, A.J. Ridgwell, A.J. Charles, C.K. Junium, A.F. Diefendorf, K.H. Freeman, N.M. Urban and I.C. Harding, 2011: Slow release of fossil carbon during the Palaeocene-Eocene Thermal Maximum. *Nature Geoscience*, **4**(7), 481-485.
- Cury, P., L. Shannon and Y.-J. Shin, 2003: The functioning of marine ecosystems: a fisheries perspective. In: *Responsible Fisheries in the Marine Ecosystem* [Sinclair, M. and G. Valdimarsson (eds.)]. FAO and CABI Publishing, Wallingford, U. K., pp. 103-124.
- Czerny, J., J. Barcelos e Ramos and U. Riebesell, 2009: Influence of elevated CO₂ concentrations on cell division and nitrogen fixation rates in the bloom-forming cyanobacterium *Nodularia spumigena*. *Biogeosciences*, **6**(9), 1865-1875.
- Dale, B., M. Edwards and P.C. Reid, 2006: Climate change and harmful algal blooms. In: *Ecology of Harmful Algae* [Granéli, E. and J.T. Turner (eds.)]. Springer, Berlin, pp. 367-378.
- Danovaro, R., C. Corinaldesi, A. Dell'Anno, J.A. Fuhrman, J.J. Middelburg, R.T. Noble and C.A. Suttle, 2011: Marine viruses and global climate change. *FEMS Microbiology Reviews*, **35**(6), 993-1034.
- Daskalov, G.M., 2003: Long-term changes in fish abundance and environmental indices in the Black Sea. *Marine Ecology Progress Series*, **255**, 259-270.
- Daufresne, M., K. Lengfellner and U. Sommer, 2009: Global warming benefits the small in aquatic ecosystems. *Proceedings of the National Academy of Sciences of the United States of America*, **106**(31), 12788-12793.
- de Baar, H.J.W., J.T.M. de Jong, D.C.E. Bakker, B.M. Löscher, C. Veth, U. Bathmann and V. Smetacek, 1995: Importance of iron for plankton blooms and carbon dioxide drawdown in the Southern Ocean. *Nature*, **373**, 412-415.
- de Baar, H.J.W., P.W. Boyd, K.H. Coale, M.R. Landry, A. Tsuda, P. Assmy, D.C.E. Bakker, Y. Bozec, R.T. Barber, M.A. Brzezinski, K.O. Buesseler, M. Boyé, P.L. Croot, F. Gervais, M.Y. Gorbunov, P.J. Harrison, W.T. Hiscock, P. Laan, C. Lancelot, C.S. Law, M. Levasseur, A. Marchetti, F.J. Millero, J. Nishioka, Y. Nojiri, T. van Oijen, U. Riebesell, M.J.A. Rijkenberg, H. Saito, S. Takeda, K.R. Timmermans, M.J.W. Veldhuis, A.M. Waite and C.-S. Wong, 2005: Synthesis of iron fertilization experiments: from the Iron Age in the age of enlightenment. *Journal of Geophysical Research*, **110**(C9), C09S16.
- De Jonckheere, J.F., M. Baumgartner, F.R. Opperdoes and K.O. Stetter, 2009: *Marinamoeba thermophila*, a new marine heterolobosean amoeba growing at 50°C. *European Journal of Protistology*, **45**(3), 231-236.
- De Jonckheere, J.F., M. Baumgartner, S. Eberhardt, F.R. Opperdoes and K.O. Stetter, 2011: *Oramoeba fumarolia* gen. nov., sp. nov., a new marine heterolobosean amoebflagellate growing at 54°C. *European Journal of Protistology*, **47**(1), 16-23.
- de Moel, H., G.M. Ganssen, F.J.C. Peeters, S.J.A. Jung, D. Kroon, G.J.A. Brummer and R.E. Zeebe, 2009: Planktic foraminiferal shell thinning in the Arabian Sea due to anthropogenic ocean acidification? *Biogeosciences*, **6**(9), 1917-1925.

- De'ath, G., J.M. Lough and K.E. Fabricius, 2009: Declining coral calcification on the Great Barrier Reef. *Science*, **323(5910)**, 116-119.
- De'ath, G., J.M. Lough and K.E. Fabricius, 2009 (corrigendum): Declining coral calcification on the Great Barrier Reef. *Science*, **323(5910)**, 116-119.
- De'ath, G., K.E. Fabricius, H. Sweatman and M. Puotinen, 2012: The 27-year decline of coral cover on the Great Barrier Reef and its causes. *Proceedings of the National Academy of Sciences of the United States of America*, **109(44)**, 17995-17999.
- Deigweiher, K., N. Koschnick, H.-O. Pörtner and M. Lucassen, 2008: Acclimation of ion regulatory capacities in gills of marine fish under environmental hypercapnia. *American Journal of Physiology: Regulatory, Integrative and Comparative Physiology*, **295(5)**, R1660-1670.
- Denman, K., J.R. Christian, N. Steiner, H.-O. Pörtner and Y. Nojiri, 2011: Potential impacts of future ocean acidification on marine ecosystems and fisheries: present knowledge and recommendations for future research. *ICES Journal of Marine Science*, **68(6)**, 1019-1029.
- Deutsch, C., H. Brix, T. Ito, H. Frenzel and L. Thompson, 2011: Climate-forced variability of ocean hypoxia. *Science*, **333(6040)**, 336-339.
- deYoung, B., R. Harris, J. Alheit, G. Beaugrand, N. Mantua and L. Shannon, 2004: Detecting regime shifts in the ocean: data considerations. *Progress in Oceanography*, **60(2-4)**, 143-164.
- deYoung, B., M. Barange, G. Beaugrand, R. Harris, R.I. Perry, M. Scheffer and F. Werner, 2008: Regime shifts in marine ecosystems: detection, prediction and management. *Trends in Ecology and Evolution*, **23(7)**, 402-409.
- DFO, 2011a: *Assessment of capelin in SA 2 + Div. 3KL in 2010*. DFO Canadian Science Advisory Secretariat Science Advisory Report 2010/090, 16 pp.
- DFO, 2011b: *Recovery potential assessment for the Newfoundland and Labrador designatable unit (NAFO Divs. 2GHJ, 3KLNO) of Atlantic Cod (Gadus morhua)*. DFO Canadian Science Advisory Secretariat Science Advisory Report 2011/037, 30 pp.
- Díaz, R.J. and R. Rosenberg, 2008: Spreading dead zones and consequences for marine ecosystems. *Science*, **321(5891)**, 926-929.
- Dierssen, H.M., 2010: Perspectives on empirical approaches for ocean color remote sensing of chlorophyll in a changing climate. *Proceedings of the National Academy of Sciences of the United States of America*, **107(40)**, 17073-17078.
- Doak, D.F., J.A. Estes, B.S. Halpern, U. Jacob, D.R. Lindberg, J. Lovvorn, D.H. Monson, M.T. Tinker, T.M. Williams, J.T. Wootton, I. Carroll, M. Emmerson, F. Micheli and M. Novak, 2008: Understanding and predicting ecological dynamics: are major surprises inevitable? *Ecology*, **89(4)**, 952-961.
- Dobson, A., 2009: Climate variability, global change, immunity, and the dynamics of infectious diseases. *Ecology*, **90(4)**, 920-927.
- Dodds, L.A., J.M. Roberts, A.C. Taylor and F. Marubini, 2007: Metabolic tolerance of the cold-water coral *Lophelia pertusa* (Scleractinia) to temperature and dissolved oxygen change. *Journal of Experimental Marine Biology and Ecology*, **349(2)**, 205-214.
- Dodson, J.J., S. Tremblay, F. Colombani, J.E. Carscadden and F. Lecomte, 2007: Trans-Arctic dispersals and the evolution of a circumpolar marine fish species complex, the capelin (*Mallotus villosus*). *Molecular Ecology*, **16(23)**, 5030-5043.
- Domenici, P., B. Allan, M.I. McCormick and P.L. Munday, 2012: Elevated carbon dioxide affects behavioural lateralization in a coral reef fish. *Biology Letters*, **8(1)**, 78-81.
- Donelson, J.M., P.L. Munday, M.I. McCormick and C.R. Pitcher, 2012: Rapid transgenerational acclimation of a tropical reef fish to climate change. *Nature Climate Change*, **2(1)**, 30-32.
- Doney, S.C., 2006: Oceanography - Plankton in a warmer world. *Nature*, **444(7120)**, 695-696.
- Doney, S.C., 2010: The growing human footprint on coastal and open-ocean biogeochemistry. *Science*, **328(5985)**, 1512-1516.
- Donner, S.D., W.J. Skirving, C.M. Little, M. Oppenheimer and O. Hoegh-Guldberg, 2005: Global assessment of coral bleaching and required rates of adaptation under climate change. *Global Change Biology*, **11(12)**, 2251-2265.
- Dore, J.E., R. Lukas, D.W. Sadler, M.J. Church and D.M. Karl, 2009: Physical and biogeochemical modulation of ocean acidification in the central North Pacific. *Proceedings of the National Academy of Sciences of the United States of America*, **106(30)**, 12235-12240.

- Douvere, F.**, 2008: The importance of marine spatial planning in advancing ecosystem-based sea use management. *Marine Policy*, **32(5)**, 762-771.
- Dove, S.G., D.I. Kline, O. Pantos, F.E. Angly, G.W. Tyson and O. Hoegh-Guldberg**, 2013 in press: Future reef decalcification under a business-as-usual CO₂ emission scenario. *Proceedings of the National Academy of Sciences of the United States of America*.
- Dowsett, H.J.**, 2007: The PRISM palaeoclimate reconstruction and Pliocene sea-surface temperature. In: *Deep-time Perspectives on Climate Change: Marrying the Signal from Computer Models and Biological Proxies* [Williams, M., A.M. Haywood, F.J. Gregory and D.N. Schmidt (eds.)]. The Micropalaeontological Society Special Publication, The Geological Society, London, UK, pp. 459-480.
- Drinkwater, K.F.**, 2006: The regime shift of the 1920s and 1930s in the North Atlantic. *Progress in Oceanography*, **68(2-4)**, 134-151.
- Drinkwater, K.F., G. Beaugrand, M. Kaeriyama, S. Kim, G. Ottersen, R.I. Perry, H.-O. Pörtner, J.J. Polovina and A. Takasuka**, 2010: On the processes linking climate to ecosystem changes. *Journal of Marine Systems*, **79(3-4)**, 374-388.
- Duce, R.A., J. LaRoche, K. Altieri, K.R. Arrigo, A.R. Baker, D.G. Capone, S. Cornell, F. Dentener, J. Galloway, R.S. Ganeshram, R.J. Geider, T. Jickells, M.M. Kuypers, R. Langlois, P.S. Liss, S.M. Liu, J.J. Middelburg, C.M. Moore, S. Nickovic, A. Oschlies, T. Pedersen, J. Prospero, R. Schlitzer, S. Seitzinger, L.L. Sorensen, M. Uematsu, O. Ulloa, M. Voss, B. Ward and L. Zamora**, 2008: Impacts of atmospheric anthropogenic nitrogen on the open ocean. *Science*, **320(5878)**, 893-897.
- Dulvy, N.K., S.I. Rogers, S. Jennings, V. Stelzenmüller, S.R. Dye and H.R. Skjoldal**, 2008: Climate change and deepening of the North Sea fish assemblage: a biotic indicator of warming seas. *Journal of Applied Ecology*, **45(4)**, 1029-1039.
- Dunkley Jones, T., D.J. Lunt, D.N. Schmidt, A. Ridgwell, A. Sluijs, P.J. Valdes and M. Maslin**, 2013: Climate model and proxy data constraints on ocean warming across the Paleocene–Eocene Thermal Maximum. *Earth-Science Reviews*, **125**, 123-145.
- Dunne, J.A. and R.J. Williams**, 2009: Cascading extinctions and community collapse in model food webs. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **364(1524)**, 1711-1723.
- Dupont, S., B. Lundve and M. Thorndyke**, 2010: Near future ocean acidification increases growth rate of the lecithotrophic larvae and juveniles of the sea star *Crossaster papposus*. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution*, **314(5)**, 382-389.
- Dupont, S., N. Dorey, M. Stumpp, F. Melzner and M. Thorndyke**, 2012: Long-term and trans-life-cycle effects of exposure to ocean acidification in the green sea urchin *Strongylocentrotus droebachiensis*. *Marine Biology*, **160(8)**, 1835-1843.
- Durack, P.J., S.E. Wijffels and R.J. Matear**, 2012: Ocean salinities reveal strong global water cycle intensification during 1950 to 2000. *Science*, **336(6080)**, 455-458.
- Dyhrman, S.T., S.T. Haley, S.R. Birkeland, L.L. Wurch, M.J. Cipriano and A.G. McArthur**, 2006: Long serial analysis of gene expression for gene discovery and transcriptome profiling in the widespread marine coccolithophore *Emiliania huxleyi*. *Applied and Environmental Microbiology*, **72(1)**, 252-260.
- Easterling, D.R., G.A. Meehl, C. Parmesan, S.A. Changnon, T.R. Karl and L.O. Mearns**, 2000: Climate extremes: observations, modeling, and impacts. *Science*, **289(5487)**, 2068-2074.
- Edmunds, P.J.**, 2011: Zooplanktivory ameliorates the effects of ocean acidification on the reef coral *Porites* spp. *Limnology and Oceanography*, **56(6)**, 2402-2410.
- Edwards, M. and A.J. Richardson**, 2004: Impact of climate change on marine pelagic phenology and trophic mismatch. *Nature*, **430(7002)**, 881-884.
- Edwards, M., P.C. Reid and B. Planque**, 2001: Long-term and regional variability of phytoplankton biomass in the Northeast Atlantic (1960–1995). *ICES Journal of Marine Science*, **58(1)**, 39-49.
- Edwards, M., D.G. Johns, S.C. Leterme, E. Svendsen and A.J. Richardson**, 2006: Regional climate change and harmful algal blooms in the Northeast Atlantic. *Limnology and Oceanography*, **51(2)**, 820-829.
- Edwards, M., G. Beaugrand, P. Helaouët, J. Alheit and S. Coombs**, 2013: Marine ecosystem response to the Atlantic Multidecadal Oscillation. *PLoS ONE*, **8(2)**, e57212.
- Eero, M., B.R. MacKenzie, F.W. Koster and H. Gislason**, 2011: Multi-decadal responses of a cod (*Gadus morhua*) population to human-induced trophic changes, fishing, and climate. *Ecological Applications*, **21(1)**, 214-226.

- Eggert, A., R.J.W. Visser, P.R. Van Hasselt and A.M. Breeman, 2006:** Differences in acclimation potential of photosynthesis in seven isolates of the tropical to warm temperate macrophyte *Valonia utricularis* (Chlorophyta). *Phycologia*, **45(5)**, 546-556.
- Eide, A., 2007:** Economic impacts of global warming: the case of the Barents Sea fisheries. *Natural Resource Modeling*, **20(2)**, 199-221.
- Eide, A. and K. Heen, 2002:** Economic impacts of global warming - a study of the fishing industry in North Norway. *Fisheries Research*, **56(3)**, 261-274.
- Ekau, W., H. Auel, H.-O. Pörtner and D. Gilbert, 2010:** Impacts of hypoxia on the structure and processes in pelagic communities (zooplankton, macro-invertebrates and fish). *Biogeosciences*, **7(5)**, 1669-1699.
- Eliason, E.J., T.D. Clark, M.J. Hague, L.M. Hanson, Z.S. Gallagher, K.M. Jeffries, M.K. Gale, D.A. Patterson, S.G. Hinch and A.P. Farrell, 2011:** Differences in thermal tolerance among sockeye salmon populations. *Science*, **332(6025)**, 109-112.
- Elmqvist, T., C. Folke, M. Nyström, G. Peterson, J. Bengtsson, B. Walker and J. Norberg, 2003:** Response diversity, ecosystem change, and resilience. *Frontiers in Ecology and the Environment*, **1(9)**, 488-494.
- Enfield, D.B., A.M. Mestas-Núñez and P.J. Trimble, 2001:** The Atlantic Multidecadal Oscillation and its relation to rainfall and river flows in the continental U.S. *Geophysical Research Letters*, **28(10)**, 2077-2080.
- Engel, A., S. Thoms, U. Riebesell, E. Rochelle-Newall and I. Zondervan, 2004:** Polysaccharide aggregation as a potential sink of marine dissolved organic carbon. *Nature*, **428(6986)**, 929-932.
- Eppley, R.W., 1972:** Temperature and phytoplankton growth in the sea. *Fishery Bulletin*, **70(4)**, 1063-1085.
- Etheridge, D.M., L.P. Steele, R.L. Langenfelds, R.J. Francey, J.M. Barnola and V.I. Morgan, 1996:** Natural and anthropogenic changes in atmospheric CO₂ over the last 1000 years from air in Antarctic ice and firn. *Journal of Geophysical Research*, **101(D2)**, 4115-4128.
- Fabricius, K.E., C. Langdon, S. Uthicke, C. Humphrey, S. Noonan, G. De'ath, R. Okazaki, N. Muehllehner, M.S. Glas and J.M. Lough, 2011:** Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations. *Nature Climate Change*, **1(3)**, 165-169.
- Fairweather, T.P., C.D. van der Lingen, A.J. Booth, L. Drapeau and J.J. van der Westhuizen, 2006:** Indicators of sustainable fishing for South African sardine *Sardinops sagax* and anchovy *Engraulis encrasicolus*. *African Journal of Marine Science*, **28(3-4)**, 661-680.
- Falkenberg, L.J., S.D. Connell and B.D. Russell, 2013:** Disrupting the effects of synergies between stressors: improved water quality dampens the effects of future CO₂ on a marine habitat. *Journal of Applied Ecology*, **50(1)**, 51-58.
- Falkowski, P.G., 1997:** Evolution of the nitrogen cycle and its influence on the biological sequestration of CO₂ in the ocean. *Nature*, **387(6630)**, 272-275.
- Falkowski, P.G. and J.A. Raven, 1997:** *Aquatic Photosynthesis*. Blackwell Science, Oxford, U.K., 375 pp.
- FAO, 2003:** *The Ecosystem Approach to Fisheries*. FAO Technical Guidelines for Responsible Fisheries, No. 4, Suppl. 2, FAO, Rome, Italy, 112 pp.
- FAO, 2012a:** PART 3 – Feeding the world. Trends in the livestock sector. In: *FAO Statistical Yearbook 2012. World Food and Agriculture* Food and Agriculture Organization of the United Nations, pp. 198-213.
- FAO, 2012b:** *The state of world fisheries and aquaculture 2012*. Food and Agriculture Organization of the United Nations, Rome, Italy, 230 pp.
- Fashchuk, D.Y., 2011:** *Marine Ecological Geography. Theory and Experience*. Springer, Berlin, 433 pp.
- Feely, R.A., C.L. Sabine, J.M. Hernandez-Ayon, D. Ianson and B. Hales, 2008:** Evidence for upwelling of corrosive "acidified" water onto the continental shelf. *Science*, **320(5882)**, 1490-1492.
- Feely, R.A., S.R. Alin, J. Newton, C.L. Sabine, M. Warner, A. Devol, C. Krembs and C. Maloy, 2010:** The combined effects of ocean acidification, mixing, and respiration on pH and carbonate saturation in an urbanized estuary. *Estuarine, Coastal and Shelf Science*, **88(4)**, 442-449.
- Feng, Y., C.E. Hare, K. Leblanc, J.M. Rose, Y. Zhang, G.R. DiTullio, P.A. Lee, S.W. Wilhelm, J.M. Rowe, J. Sun, N. Nemcek, C. Gueguen, U. Passow, I. Benner, C. Brown and D.A. Hutchins, 2009:** Effects of increased pCO₂ and temperature on the North Atlantic spring bloom. I. The phytoplankton community and biogeochemical response. *Marine Ecology Progress Series*, **388**, 13-25.
- Fernandez, C., L. Farías and O. Ulloa, 2011:** Nitrogen fixation in denitrified marine waters. *PLoS ONE*, **6(6)**, e20539.

- Fernández-Reiriz**, M.J., P. Range, X.A. Álvarez-Salgado and U. Labarta, 2011: Physiological energetics of juvenile clams (*Ruditapes decussatus*) in a high CO₂ coastal ocean. *Marine Ecology Progress Series*, **433**, 97-105.
- Fernando**, H.J.S., J.L. McCulley, S.G. Mendis and K. Perera, 2005: Coral poaching worsens Tsunami destruction in Sri Lanka. *Eos Transactions of the American Geophysical Union*, **86(33)**, 301-304.
- Ferrari**, M.C.O., D.L. Dixon, P.L. Munday, M.I. McCormick, M.G. Meekan, A. Sih and D.P. Chivers, 2011: Intrageneric variation in antipredator responses of coral reef fishes affected by ocean acidification: implications for climate change projections on marine communities. *Global Change Biology*, **17(9)**, 2980-2986.
- Field**, C.B., M.J. Behrenfeld, J.T. Randerson and P. Falkowski, 1998: Primary production of the biosphere: integrating terrestrial and oceanic components. *Science*, **281(5374)**, 237-240.
- Field**, D.B., T.R. Baumgartner, C.D. Charles, V. Ferreira-Bartrina and M.D. Ohman, 2006: Planktonic foraminifera of the California Current reflect 20th-century warming. *Science*, **311(5757)**, 63-66.
- Findlay**, H.S., M.A. Kendall, J.I. Spicer and S. Widdicombe, 2010: Post-larval development of two intertidal barnacles at elevated CO₂ and temperature. *Marine Biology*, **157(4)**, 725-735.
- Fish**, M.R., A. Lombana and C. Drews, 2009: *Climate Change and Marine Turtles in the Wider Caribbean: Regional Climate Projections*. WWF, San José, CA, USA, 20 pp.
- Flombaum**, P., J.L. Gallegos, R.A. Gordillo, J. Rincón, L.L. Zabala, N. Jiao, D.M. Karl, W.K.W. Li, M.W. Lomas, D. Veneziano, C.S. Vera, J.A. Vrugt and A.C. Martiny, 2013: Present and future global distributions of the marine Cyanobacteria *Prochlorococcus* and *Synechococcus*. *Proceedings of the National Academy of Sciences of the United States of America*, **110(24)**, 9824-9829.
- Folke**, C., S. Carpenter, B. Walker, M. Scheffer, T. Elmqvist, L. Gunderson and C.S. Holling, 2004: Regime shifts, resilience, and biodiversity in ecosystem management. *Annual Review of Ecology, Evolution, and Systematics*, **35(1)**, 557-581.
- Folt**, C.L., C.Y. Chen, M.V. Moore and J. Burnaford, 1999: Synergism and antagonism among multiple stressors. *Limnology and Oceanography*, **44(3)**, 864-877.
- Form**, A. and U. Riebesell, 2012: Acclimation to ocean acidification during long-term CO₂ exposure in the cold-water coral *Lophelia pertusa*. *Global Change Biology*, **18(3)**, 843-853.
- Foster**, L.C., D.N. Schmidt, E. Thomas, S. Arndt and A. Ridgwell, 2013: Surviving rapid climate change in the deep sea during the Paleogene hyperthermals. *Proceedings of the National Academy of Sciences of the United States of America*, **110(23)**, 9273-9276.
- Frank**, K.T., B. Petrie, J.S. Choi and W.C. Leggett, 2005: Trophic cascades in a formerly cod-dominated ecosystem. *Science*, **308(5728)**, 1621-1623.
- Franz**, J., G. Krahnemann, G. Lavik, P. Grasse, T. Dittmar and U. Riebesell, 2012: Dynamics and stoichiometry of nutrients and phytoplankton in waters influenced by the oxygen minimum zone in the eastern tropical Pacific. *Deep-Sea Research Part I: Oceanographic Research Papers*, **62**, 20-31.
- Friedland**, K.D. and C.D. Todd, 2012: Changes in Northwest Atlantic Arctic and Subarctic conditions and the growth response of Atlantic salmon. *Polar Biology*, **35(4)**, 593-609.
- Friedland**, K.D., C. Stock, K.F. Drinkwater, J.S. Link, R.T. Leaf, B.V. Shank, J.M. Rose, C.H. Pilskaln and M.J. Fogarty, 2012: Pathways between primary production and fisheries yields of large marine ecosystems. *PLoS ONE*, **7(1)**, e28945.
- Friedrich**, T., A. Timmermann, A. Abe-Ouchi, N.R. Bates, M.O. Chikamoto, M.J. Church, J.E. Dore, D.K. Gledhill, M. González-Dávila, M. Heinemann, T. Ilyina, J.H. Jungclaus, E. McLeod, A. Mouchet and J.M. Santana-Casiano, 2012: Detecting regional anthropogenic trends in ocean acidification against natural variability. *Nature Climate Change*, **2(3)**, 167-171.
- Frommel**, A.Y., R. Maneja, D. Lowe, A.M. Malzahn, A.J. Geffen, A. Folkvord, U. Piatkowski, T.B.H. Reusch and C. Clemmesen, 2012: Severe tissue damage in Atlantic cod larvae under increasing ocean acidification. *Nature Climate Change*, **2(1)**, 42-46.
- Fu**, F.-X., A.O. Tatters and D.A. Hutchins, 2012: Global change and the future of harmful algal blooms in the ocean. *Marine Ecology Progress Series*, **470**, 207-233.
- Fu**, F.-X., A.R. Place, N.S. Garcia and D.A. Hutchins, 2010: CO₂ and phosphate availability control the toxicity of the harmful bloom dinoflagellate *Karlodinium veneficum*. *Aquatic Microbial Ecology*, **59(1)**, 55-65.
- Fu**, F.-X., M.E. Warner, Y. Zhang, Y. Feng and D.A. Hutchins, 2007: Effects of increased temperature and CO₂ on photosynthesis, growth, and elemental ratios in marine *Synechococcus* and *Prochlorococcus* (Cyanobacteria). *Journal of Phycology*, **43(3)**, 485-496.

- Fu, F.-X., Y. Zhang, M.E. Warner, Y. Feng, J. Sun and D.A. Hutchins, 2008: A comparison of future increased CO₂ and temperature effects on sympatric *Heterosigma akashiwo* and *Prorocentrum minimum*. *Harmful Algae*, **7(1)**, 76-90.
- Fuentes, M.M.P.B., J.A. Maynard, M. Guinea, I.P. Bell, P.J. Werdell and M. Hamann, 2009: Proxy indicators of sand temperature help project impacts of global warming on sea turtles in northern Australia. *Endangered Species Research*, **9(1)**, 33-40.
- Fulton, E.A., 2011: Interesting times: winners, losers, and system shifts under climate change around Australia. *ICES Journal of Marine Science*, **68(6)**, 1329-1342.
- Fulton, E.A., J.S. Link, I.C. Kaplan, M. Savina-Rolland, P. Johnson, C. Ainsworth, P. Horne, R. Gorton, R.J. Gamble, A.D.M. Smith and D.C. Smith, 2011: Lessons in modelling and management of marine ecosystems: the Atlantis experience. *Fish and Fisheries*, **12(2)**, 171-188.
- Galbraith, H., R. Jones, R. Park, J. Clough, S. Herrod-Julius, B. Harrington and G. Page, 2005: Global climate and sea level rise: potential losses of intertidal habitat for shorebirds. In: *Bird Conservation Implementation and Integration in the Americas: Proceedings of the Third International Partners in Flight Conference, 2002, March 20-24, Asilomar, CA, Volume 2, General Technical Report, PSW-GTR-191* [Ralph, C.J. and T.D. Rich (eds.)]. US Dept. of Agriculture, Forest Service, Pacific Southwest Research Station, Albany, CA, USA, pp. 1119-1122.
- Garcia, S.M. and A.A. Rosenberg, 2010: Food security and marine capture fisheries: characteristics, trends, drivers and future perspectives. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **365(1554)**, 2869-2880.
- Gardner, B., P.J. Sullivan, S. Epperly and S.J. Morreale, 2008: Hierarchical modeling of bycatch rates of sea turtles in the western North Atlantic. *Endangered Species Research*, **5**, 279-289.
- Gazeau, F., L.M. Parker, S. Comeau, J.-P. Gattuso, W.A. O'Connor, S. Martin, H.-O. Pörtner and P.M. Ross, 2013: Impacts of ocean acidification on marine shelled molluscs. *Marine Biology*, **160(8)**, 2207-2245.
- Genner, M.J., D.W. Sims, V.J. Wearmouth, E.J. Southall, A.J. Southward, P.A. Henderson and S.J. Hawkins, 2004: Regional climatic warming drives long-term community changes of British marine fish. *Proceedings of the Royal Society B: Biological Sciences*, **271(1539)**, 655-661.
- Genner, M.J., D.W. Sims, A.J. Southward, G.C. Budd, P. Masterson, M. McHugh, P. Rendle, E.J. Southall, V.J. Wearmouth and S.J. Hawkins, 2010: Body size-dependent responses of a marine fish assemblage to climate change and fishing over a century-long scale. *Global Change Biology*, **16(2)**, 517-527.
- Gibbs, S.J., P.R. Bown, J.A. Sessa, T.J. Bralower and P.A. Wilson, 2006: Nannoplankton extinction and origination across the Paleocene-Eocene Thermal Maximum. *Science*, **314(5806)**, 1770-1773.
- Gilly, W.F., J.M. Beman, S.Y. Litvin and B.H. Robison, 2013: Oceanographic and biological effects of shoaling of the oxygen minimum zone. *Annual Review of Marine Science*, **5**, 393-420.
- Gilly, W.F., U. Markaida, C.H. Baxter, B.A. Block, A. Boustany, L. Zeidberg, K. Reisenbichler, B. Robison, G. Bazzino and C. Salinas, 2006: Vertical and horizontal migrations by the jumbo squid *Dosidicus gigas* revealed by electronic tagging. *Marine Ecology Progress Series*, **324**, 1-17.
- Giordano, M., J. Beardall and J.A. Raven, 2005: CO₂ concentrating mechanisms in algae: mechanisms, environmental modulation, and evolution. *Annual Review of Plant Biology*, **56**, 99-131.
- Giovannoni, S.J. and K.L. Vergin, 2012: Seasonality in ocean microbial communities. *Science*, **335(6069)**, 671-676.
- Glynn, P.W. and L. D'Croz, 1990: Experimental evidence for high temperature stress as the cause of El Niño-coincident coral mortality. *Coral Reefs*, **8(4)**, 181-191.
- Godfrey, M.H., A.F. D'Amato, M.Â. Marcovaldi and N. Mrosovsky, 1999: Pivotal temperature and predicted sex ratios for hatchling hawksbill turtles from Brazil. *Canadian Journal of Zoology*, **77(9)**, 1465-1473.
- Gómez, I., A. Wulff, M. Roleda, P. Huovinen, U. Karsten, M.L. Quartino, K. Dunton and C. Wiencke, 2011: Light and temperature demands of benthic algae in the polar regions. In: *Biology of Polar Benthic Algae* [Wiencke, C. (ed.)]. de Gruyter, Berlin, pp. 195-220.
- González-Taboada, F. and R. Anadón, 2012: Patterns of change in sea surface temperature in the North Atlantic during the last three decades: beyond mean trends. *Climatic Change*, **115(2)**, 419-431.
- Gooday, A.J., B.J. Bett, E. Escobar, B. Ingole, L.A. Levin, C. Neira, A.V. Raman and J. Sellanes, 2010: Habitat heterogeneity and its relationship to biodiversity in oxygen minimum zones. *Marine Ecology*, **31**, 125-147.
- Gooding, R.A., C.D.G. Harley and E. Tang, 2009: Elevated water temperature and carbon dioxide concentration increase the growth of a keystone echinoderm. *Proceedings of the National Academy of Sciences of the United States of America*, **106(23)**, 9316-9321.

- Goreau, T.J.** and R.L. Hayes, 1994: Coral bleaching and ocean "Hot Spots". *Ambio*, **23(3)**, 176-180.
- Granier, C.**, U. Niemeier, J.H. Jungclaus, L. Emmons, P. Hess, J.F. Lamarque, S. Walters and G.P. Brasseur, 2006: Ozone pollution from future ship traffic in the Arctic northern passages. *Geophysical Research Letters*, **33(13)**, L13807.
- Gravelle, G.** and N. Mimura, 2008: Vulnerability assessment of sea-level rise in Viti Levu, Fiji Islands. *Sustainability Science*, **3(2)**, 171-180.
- Gray, J.S.**, R.S.S. Wu and Y.Y. Or, 2002: Effects of hypoxia and organic enrichment on the coastal marine environment. *Marine Ecology Progress Series*, **238**, 249-279.
- Green, J.L.**, B.J.M. Bohannan and R.J. Whitaker, 2008: Microbial biogeography: from taxonomy to traits. *Science*, **320(5879)**, 1039-1043.
- Greene, C.H.** and A.J. Pershing, 2003: The flip-side of the North Atlantic Oscillation and modal shifts in slope-water circulation patterns. *Limnology and Oceanography*, **48(1)**, 319-322.
- Greene, C.H.** and A.J. Pershing, 2007: Climate drives sea change. *Science*, **315(5815)**, 1084-1085.
- Greene, C.H.**, A.J. Pershing, R.D. Kenney and J.W. Jossi, 2003: Impact of climate variability on the recovery of endangered North Atlantic right whales. *Oceanography*, **16(4)**, 98-103.
- Grémillet, D.** and T. Boulinier, 2009: Spatial ecology and conservation of seabirds facing global climate change: a review. *Marine Ecology Progress Series*, **391**, 121-137.
- Grieshaber, M.**, I. Hardewig, U. Kreutzer and H.-O. Pörtner, 1994: Physiological and metabolic responses to hypoxia in invertebrates. In: *Reviews of Physiology, Biochemistry and Pharmacology* [Blaustein, M.P., H. Grunicke, E. Habermann, D. Pette, H. Reuter, B. Sakmann, M. Schweiger, E. Weibel and E.M. Wright (eds.)]. Springer, Berlin/Heidelberg, pp. 43-147.
- Griffith, G.P.**, E.A. Fulton and A.J. Richardson, 2011: Effects of fishing and acidification-related benthic mortality on the southeast Australian marine ecosystem. *Global Change Biology*, **17(10)**, 3058-3074.
- Griffith, G.P.**, E.A. Fulton, R. Gorton and A.J. Richardson, 2012: Predicting interactions among fishing, ocean warming, and ocean acidification in a marine system with whole-ecosystem models. *Conservation Biology*, **26(6)**, 1145-1152.
- Grossart, H.P.**, M. Allgaier, U. Passow and U. Riebesell, 2006: Testing the effect of CO₂ concentration on the dynamics of marine heterotrophic bacterioplankton. *Limnology and Oceanography*, **51(1)**, 1-11.
- Gruber, N.**, 2011: Warming up, turning sour, losing breath: ocean biogeochemistry under global change. *Philosophical Transactions of the Royal Society A: Mathematical, Physical, and Engineering Sciences*, **369(1943)**, 1980-1996.
- Guinotte, J.M.**, J. Orr, S. Cairns, A. Freiwald, L. Morgan and R. George, 2006: Will human-induced changes in seawater chemistry alter the distribution of deep-sea scleractinian corals? *Frontiers in Ecology and the Environment*, **4(3)**, 141-146.
- Gutowska, M.A.**, H.-O. Pörtner and F. Melzner, 2008: Growth and calcification in the cephalopod *Sepia officinalis* under elevated seawater pCO₂. *Marine Ecology Progress Series*, **373**, 303-309.
- Haines, A.**, R.S. Kovats, D. Campbell-Lendrum and C. Corvalan, 2006: Climate change and human health: impacts, vulnerability, and mitigation. *The Lancet*, **367(9528)**, 2101-2109.
- Hales, S.**, P. Weinstein and A. Woodward, 1999: Ciguatera (fish poisoning), El Niño, and Pacific sea surface temperatures. *Ecosystem Health*, **5(1)**, 20-25.
- Halfar, J.**, S. Hetzinger, W. Adey, T. Zack, G. Gamboa, B. Kunz, B. Williams and D.E. Jacob, 2011: Coralline algal growth-increment widths archive North Atlantic climate variability. *Palaeogeography, Palaeoclimatology, Palaeoecology*, **302(1-2)**, 71-80.
- Hall-Spencer, J.M.**, R. Rodolfo-Metalpa, S. Martin, E. Ransome, M. Fine, S.M. Turner, S.J. Rowley, D. Tedesco and M.-C. Buia, 2008: Volcanic carbon dioxide vents show ecosystem effects of ocean acidification. *Nature*, **454(7200)**, 96-99.
- Hallegraeff, G.M.**, 2010: Ocean climate change, phytoplankton community responses, and harmful algal blooms: a formidable predictive challenge. *Journal of Phycology*, **46(2)**, 220-235.
- Hannah, C.**, A. Vezina and M. St John, 2010: The case for marine ecosystem models of intermediate complexity. *Progress in Oceanography*, **84(1-2)**, 121-128.
- Hannesson, R.**, 2007: Global warming and fish migrations. *Natural Resource Modeling*, **20(2)**, 301-319.
- Hansen, P.J.**, N. Lundholm and B. Rost, 2007: Growth limitation in marine red-tide dinoflagellates: effects of pH versus inorganic carbon availability. *Marine Ecology Progress Series*, **334**, 63-71.

- Hare, J.A., M.J. Wuenschel and M.E. Kimball, 2012:** Projecting range limits with coupled thermal tolerance - climate change models: An example based on gray snapper (*Lutjanus griseus*) along the US East coast. *PLoS ONE*, **7(12)**, e52294.
- Harley, C.D.G., 2011:** Climate change, keystone predation, and biodiversity loss. *Science*, **334(6059)**, 1124-1127.
- Harley, C.D.G., K.M. Anderson, K.W. Demes, J.P. Jorve, R.L. Kordas, T.A. Coyle and M.H. Graham, 2012:** Effects of climate change on global seaweed communities. *Journal of Phycology*, **48(5)**, 1064-1078.
- Harvell, D., S. Altizer, I.M. Cattadori, L. Harrington and E. Weil, 2009:** Climate change and wildlife diseases: When does the host matter the most? *Ecology*, **90(4)**, 912-920.
- Harvey, B.P., D. Gwynn-Jones and P.J. Moore, 2013:** Meta-analysis reveals complex marine biological responses to the interactive effects of ocean acidification and warming. *Ecology and Evolution*, **3(4)**, 1016-1030.
- Hashioka, T. and Y. Yamanaka, 2007:** Ecosystem change in the western North Pacific associated with global warming using 3D-NEMURO. *Ecological Modelling*, **202(1-2)**, 95-104.
- Hawkes, L.A., A.C. Broderick, M.H. Godfrey and B.J. Godley, 2009:** Climate change and marine turtles. *Endangered Species Research*, **7(2)**, 137-154.
- Hawkins, S.J., 2012:** Marine conservation in a rapidly changing world. *Aquatic Conservation-Marine and Freshwater Ecosystems*, **22(3)**, 281-287.
- Hays, G.C., A.C. Broderick, F. Glen and B.J. Godley, 2003:** Climate change and sea turtles: a 150-year reconstruction of incubation temperatures at a major marine turtle rookery. *Global Change Biology*, **9(4)**, 642-646.
- Haywood, A.M., M.A. Chandler, P.J. Valdes, U. Salzmann, D.J. Lunt and H.J. Dowsett, 2009:** Comparison of mid-Pliocene climate predictions produced by the HadAM3 and GCMAM3 General Circulation Models. *Global and Planetary Change*, **66(3-4)**, 208-224.
- Hazen, E.L., J.K. Craig, C.P. Good and L.B. Crowder, 2009:** Vertical distribution of fish biomass in hypoxic waters on the Gulf of Mexico shelf. *Marine Ecology Progress Series*, **375**, 195-207.
- Hazen, E.L., S. Jorgensen, R.R. Rykaczewski, S.J. Bograd, D.G. Foley, I.D. Jonsen, S.A. Shaffer, J.P. Dunne, D.P. Costa, L.B. Crowder and B.A. Block, 2013:** Predicted habitat shifts of Pacific top predators in a changing climate. *Nature Climate Change*, **3**, 234-238.
- Head, E.J.H. and P. Pepin, 2010:** Spatial and inter-decadal variability in plankton abundance and composition in the Northwest Atlantic (1958–2006). *Journal of Plankton Research*, **32(12)**, 1633-1648.
- Heath, M., M. Edwards, R. Furness, J. Pinnegar and S. Wanless, 2009:** A view from above: changing seas, seabirds and food sources. In: *Marine Climate Change Ecosystem Linkages Report Card* [Baxter, J.M., P.J. Buckley and M.T. Frost (eds.)]. MCCIP, Lowestoft, UK, pp. 24.
- Heisler, N. (ed.), 1986:** *Acid-base Regulation in Animals*. Elsevier, Amsterdam, Netherlands, 492 pp.
- Helm, K.P., N.L. Bindoff and J.A. Church, 2010:** Changes in the global hydrological-cycle inferred from ocean salinity. *Geophysical Research Letters*, **37(18)**, L18701.
- Henson, S., H. Cole, C. Beaulieu and A. Yool, 2013:** The impact of global warming on seasonality of ocean primary production. *Biogeosciences*, **10**, 4357-4369.
- Henson, S.A., J.L. Sarmiento, J.P. Dunne, L. Bopp, I. Lima, S.C. Doney, J. John and C. Beaulieu, 2010:** Detection of anthropogenic climate change in satellite records of ocean chlorophyll and productivity. *Biogeosciences*, **7(2)**, 621-640.
- Higginson, M.J., M.A. Altabet, D.W. Murray, R.W. Murray and T.D. Herbert, 2004:** Geochemical evidence for abrupt changes in relative strength of the Arabian monsoons during a stadial/interstadial climate transition. *Geochimica et Cosmochimica Acta*, **68(19)**, 3807-3826.
- Hill, V.J., P.A. Matrai, E. Olson, S. Suttles, M. Steele, L.A. Codispoti and R.C. Zimmerman, 2013:** Synthesis of integrated primary production in the Arctic Ocean: II. *In situ* and remotely sensed estimates. *Progress in Oceanography*, **110**, 107-125.
- Hinder, S.L., G.C. Hays, M. Edwards, E.C. Roberts, A.W. Walne and M.B. Gravenor, 2012:** Changes in marine dinoflagellate and diatom abundance under climate change. *Nature Climate Change*, **2(4)**, 271-275.
- Hoegh-Guldberg, O., 1999:** Climate change, coral bleaching and the future of the world's coral reefs. *Marine and Freshwater Research*, **50(8)**, 839-866.
- Hoegh-Guldberg, O., 2011:** Coral reef ecosystems and anthropogenic climate change. *Regional Environmental Change*, **11**, 215-227.

- Hoegh-Guldberg, O.** and G.J. Smith, 1989: The effect of sudden changes in temperature, light and salinity on the population density and export of zooxanthellae from the reef corals *Stylophora pistillata* Esper and *Seriatopora hystrix* Dana. *Journal of Experimental Marine Biology and Ecology*, **129(3)**, 279-303.
- Hoegh-Guldberg, O.** and B. Salvat, 1995: Periodic mass-bleaching and elevated sea temperatures: bleaching of outer reef slope communities in Moorea, French Polynesia. *Marine Ecology Progress Series*, **121**, 181-190.
- Hoegh-Guldberg, O.** and J.F. Bruno, 2010: The impact of climate change on the world's marine ecosystems. *Science*, **328(5985)**, 1523-1528.
- Hoel, A.H.**, 2009: *Best Practices in Ecosystem Based Ocean Management in the Arctic*. Norsk Polarinstitutt, Tromsø, 116 pp.
- Hoffmann, L.J.**, E. Breitbart, P.W. Boyd and K.A. Hunter, 2012: Influence of ocean warming and acidification on trace metal biogeochemistry. *Marine Ecology Progress Series*, **470**, 191-205.
- Holcomb, M.**, A.L. Cohen and D.C. McCorkle, 2012: An investigation of the calcification response of the scleractinian coral *Astrangia poculata* to elevated $p\text{CO}_2$ and the effects of nutrients, zooxanthellae and gender. *Biogeosciences*, **9(1)**, 29-39.
- Holt, J.**, S. Wakelin, J. Lowe and J. Tinker, 2010: The potential impacts of climate change on the hydrography of the northwest European continental shelf. *Progress in Oceanography*, **86(3-4)**, 361-379.
- Hönisch, B.**, A. Ridgwell, D.N. Schmidt, E. Thomas, S.J. Gibbs, A. Sluijs, R. Zeebe, L. Kump, R.C. Martindale, S.E. Greene, W. Kiessling, J. Ries, J.C. Zachos, D.L. Royer, S. Barker, T.M. Marchitto, R. Moyer, C. Pelejero, P. Ziveri, G.L. Foster and B. Williams, 2012: The geological record of ocean acidification. *Science*, **335(6072)**, 1058-1063.
- Hoppe, C.J.M.**, G. Langer and B. Rost, 2011: *Emiliania huxleyi* shows identical responses to elevated $p\text{CO}_2$ in TA and DIC manipulations. *Journal of Experimental Marine Biology and Ecology*, **406(1-2)**, 54-62.
- Hoppe, H.-G.**, K. Gocke, R. Koppe and C. Begler, 2002: Bacterial growth and primary production along a north-south transect of the Atlantic Ocean. *Nature*, **416(6877)**, 168-171.
- House, K.Z.**, C.H. House, D.P. Schrag and M.J. Aziz, 2007: Electrochemical acceleration of chemical weathering as an energetically feasible approach to mitigating anthropogenic climate change. *Environmental Science & Technology*, **41(24)**, 8464-8470.
- Howarth, R.**, F. Chan, D.J. Conley, J. Garnier, S.C. Doney, R. Marino and G. Billen, 2011: Coupled biogeochemical cycles: eutrophication and hypoxia in temperate estuaries and coastal marine ecosystems. *Frontiers in Ecology and the Environment*, **9(1)**, 18-26.
- Howells, E.J.**, V.H. Beltran, N.W. Larsen, L.K. Bay, B.L. Willis and M.J.H. van Oppen, 2012: Coral thermal tolerance shaped by local adaptation of photosymbionts. *Nature Climate Change*, **2(2)**, 116-120.
- Hsieh, C.-H.**, C.S. Reiss, R.P. Hewitt and G. Sugihara, 2008: Spatial analysis shows that fishing enhances the climatic sensitivity of marine fishes. *Canadian Journal of Fisheries and Aquatic Sciences*, **65(5)**, 947-961.
- Hsieh, C.-H.**, C.S. Reiss, J.R. Hunter, J.R. Beddington, R.M. May and G. Sugihara, 2006: Fishing elevates variability in the abundance of exploited species. *Nature*, **443(7113)**, 859-862.
- Huber, R.**, T.A. Langworthy, H. König, M. Thomm, C.R. Woese, U.B. Sleytr and K.O. Stetter, 1986: *Thermotoga maritima* sp. nov. represents a new genus of unique extremely thermophilic eubacteria growing up to 90°C. *Archives of Microbiology*, **144(4)**, 324-333.
- Huey, R.B.** and J.G. Kingsolver, 1989: Evolution of thermal sensitivity of ectotherm performance. *Trends in Ecology and Evolution*, **4(5)**, 131-135.
- Hughes, L.**, 2000: Biological consequences of global warming: is the signal already apparent? *Trends in Ecology and Evolution*, **15(2)**, 56-61.
- Hughes, R.G.**, 2004: Climate change and loss of saltmarshes: consequences for birds. *Ibis*, **146**, 21-28.
- Hughes, T.P.**, A.H. Baird, E.A. Dinsdale, N.A. Moltschanowskyj, M.S. Pratchett, J.E. Tanner and B.L. Willis, 2012: Assembly rules of reef corals are flexible along a steep climatic gradient. *Current Biology*, **22(8)**, 736-741.
- Hunt, B.P.V.**, E.A. Pakhomov, G.W. Hosie, V. Siegel, P. Ward and K. Bernard, 2008: Pteropods in Southern Ocean ecosystems. *Progress in Oceanography*, **78(3)**, 193-221.
- Huntley, B.**, R.E. Green, Y.C. Collingham and S.G. Willis, 2007: *A climatic atlas of European breeding birds*. Lynx Editions, Barcelona, 1-521 pp.
- Hutchins, D.A.**, M.R. Mulholland and F.-X. Fu, 2009: Nutrient cycles and marine microbes in a CO_2 -enriched ocean. *Oceanography*, **22(4)**, 128-145.
- Hutchins, D.A.**, F.-X. Fu, E.A. Webb, N. Walworth and A. Tagliabue, 2013: Taxon-specific response of marine nitrogen fixers to elevated carbon dioxide concentrations. *Nature Geoscience*, **6**, 790-795.

- Hutchins, D.A., F.-X. Fu, Y. Zhang, M.E. Warner, Y. Feng, K. Portune, P.W. Bernhardt and M.R. Mulholland, 2007:** CO₂ control of *Trichodesmium* N₂ fixation, photosynthesis, growth rates, and elemental ratios: implications for past, present, and future ocean biogeochemistry. *Limnology and Oceanography*, **52(4)**, 1293-1304.
- Iglesias-Rodriguez, M.D., P.R. Halloran, R.E. Rickaby, I.R. Hall, E. Colmenero-Hidalgo, J.R. Gittins, D.R. Green, T. Tyrrell, S.J. Gibbs, P. von Dassow, E. Rehm, E.V. Armbrust and K.P. Boessenkool, 2008:** Phytoplankton calcification in a high-CO₂ world. *Science*, **320(5874)**, 336-340.
- Ilyina, T., R.E. Zeebe and P.G. Brewer, 2010:** Future ocean increasingly transparent to low-frequency sound owing to carbon dioxide emissions. *Nature Geoscience*, **3(1)**, 18-22.
- IPCC, 2000 - Nakicenovic, N., and R. Swart (Eds.)** *Special report: Emissions scenarios*. Cambridge University Press, Cambridge, UK, 570 pp.
- IPCC, 2012a - Field, C.B., V. Barros, T.F. Stocker, D. Qin, D.J. Dokken, K.L. Ebi, M.D. Mastrandrea, K.J. Mach, G.-K. Plattner, S.K. Allen, M. Tignor, and P.M. Midgley (Eds.)** *Special report: Managing the Risks of Extreme Events and Disasters to Advance Climate Change Adaptation*. Cambridge University Press, Cambridge, UK, 582 pp.
- IPCC, 2012b:** *Meeting Report of the Intergovernmental Panel on Climate Change Expert Meeting on Geoengineering. Prepared by IPCC Working Group III Technical Support Unit*. Potsdam Institute for Climate Impact Research Potsdam, Germany, 99 pp.
- Ishimatsu, A. and A. Dissanayake, 2010:** Life threatened in acidic coastal waters. In: *Coastal Environmental and Ecosystem Issues of the East China Sea* [Ishimatsu, A. and H.-J. Lie (eds.)]. TERRAPUB and Nagasaki University, Nagasaki, pp. 283–303.
- Ishimatsu, A., M. Hayashi and T. Kikkawa, 2008:** Fishes in high-CO₂, acidified oceans. *Marine Ecology Progress Series*, **373**, 295-302.
- Ito, S., K.A. Rose, A.J. Miller, K. Drinkwater, K.M. Brander, J.E. Overland, S. Sundby, E. Curchitser, J.W. Hurrell and Y. Yamanaka, 2010:** Ocean ecosystem responses to future global change scenarios: a way forward. In: *Global Change and Marine Ecosystems* [Barange, M., J.G. Field, R.H. Harris, E. Hofmann, R.I. Perry and F. Werner (eds.)]. Oxford University Press, pp. 287-322.
- Jaccard, S.L. and E.D. Galbraith, 2012:** Large climate-driven changes of oceanic oxygen concentrations during the last deglaciation. *Nature Geoscience*, **5(2)**, 151-156.
- Jackson, G.A. and A.B. Burd, 2001:** A model for the distribution of particle flux in the mid-water column controlled by subsurface biotic interactions. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **49(1-3)**, 193-217.
- Jackson, J.B.C. and K.G. Johnson, 2000:** Life in the last few million years. *Paleobiology*, **26(4)**, 221-235.
- Jacobs, S.S. and C.F. Giulivi, 2010:** Large multidecadal salinity trends near the Pacific-Antarctic continental margin. *Journal of Climate*, **23(17)**, 4508-4524.
- Jantzen, C., V. Häussermann, G. Försterra, J. Laudien, M. Ardelan, S. Maier and C. Richter, 2013:** Occurrence of a cold-water coral along natural pH gradients (Patagonia, Chile). *Marine Biology*, **160(10)**, 2597-2607.
- Jarre, A. and L.J. Shannon, 2010:** Regime shifts: physical-biological interactions under climatic and anthropogenic pressures. In: *Marine Ecosystems and Global Change* [Barange, M., J.G. Field, R.P. Harris, E.E. Hofmann, R.I. Perry and F. Werner (eds.)]. Oxford University Press, Oxford, UK, pp. 215-216.
- Jenkyns, H.C., 2010:** Geochemistry of oceanic anoxic events. *Geochemistry Geophysics Geosystems*, **11**, Q03004.
- Jenouvrier, S., M. Holland, J. Stroeve, C. Barbraud, H. Weimerskirch, M. Serreze and H. Caswell, 2012:** Effects of climate change on an emperor penguin population: analysis of coupled demographic and climate models. *Global Change Biology*, **18(9)**, 2756-2770.
- Jessen, G.L., R.A. Quiñones and R.R. González, 2009:** Aerobic and anaerobic enzymatic activity and allometric scaling of the deep benthic polychaete *Hyalinoecia artifex* (Polychaeta: Onuphidae). *Journal of the Marine Biological Association of the United Kingdom*, **89(6)**, 1171-1175.
- Jin, D., E. Thunberg and P. Hoagland, 2008:** Economic impact of the 2005 red tide event on commercial shellfish fisheries in New England. *Ocean & Coastal Management*, **51(5)**, 420-429.
- Jin, X. and N. Gruber, 2003:** Offsetting the radiative benefit of ocean iron fertilization by enhancing N₂O emissions. *Geophysical Research Letters*, **30(24)**, 2249.
- Johns, D.G., M. Edwards and S.D. Batten, 2001:** Arctic boreal plankton species in the Northwest Atlantic. *Canadian Journal of Fisheries and Aquatic Sciences*, **58(11)**, 2121-2124.

- Johns**, D.G., M. Edwards, A. Richardson and J.I. Spicer, 2003: Increased blooms of a dinoflagellate in the NW Atlantic. *Marine Ecology Progress Series*, **265**, 283-287.
- Johnson**, K.S., S.C. Riser and D.M. Karl, 2010: Nitrate supply from deep to near-surface waters of the North Pacific subtropical gyre. *Nature*, **465(7301)**, 1062-1065.
- Joint**, I., S.C. Doney and D.M. Karl, 2011: Will ocean acidification affect marine microbes? *ISME Journal*, **5(1)**, 1-7.
- Jones**, A.M., R. Berkelmans, M.J.H. van Oppen, J.C. Mieog and W. Sinclair, 2008: A community change in the algal endosymbionts of a scleractinian coral following a natural bleaching event: field evidence of acclimatization. *Proceedings of the Royal Society B: Biological Sciences*, **275(1641)**, 1359-1365.
- Jones**, M.C., S.R. Dye, J.A. Fernandes, T.L. Frölicher, J.K. Pinnegar, R. Warren and W.W.L. Cheung, 2013: Predicting the impact of climate change on threatened species in UK waters. *PLoS ONE*, **8(1)**, e54216.
- Jones**, P.D., M. New, D.E. Parker, S. Martin and I.G. Rigor, 1999: Surface air temperature and its changes over the past 150 years. *Reviews of Geophysics*, **37(2)**, 173-199.
- Jones**, R.J., O. Hoegh-Guldberg, A.W.D. Larkum and U. Schreiber, 1998: Temperature-induced bleaching of corals begins with impairment of the CO₂ fixation mechanism in zooxanthellae. *Plant, Cell and Environment*, **21(12)**, 1219-1230.
- Jutfelt**, F., K. Bresolin de Souza, A. Vuylsteke and J. Sturve, 2013: Behavioural disturbances in a temperate fish exposed to sustained high-CO₂ levels. *PLoS ONE*, **8(6)**, e65825.
- Kaniewska**, P., P.R. Campbell, D.I. Kline, M. Rodriguez-Lanetty, D.J. Miller, S. Dove and O. Hoegh-Guldberg, 2012: Major cellular and physiological impacts of ocean acidification on a reef building coral. *PLoS ONE*, **7(4)**, e34659.
- Karl**, D.M., N. Bates, S. Emerson, P.J. Harrison, C. Jeandel, O. Llinás, K.K. Liu, J.-C. Matry, A.F. Michaels, J.C. Miquel, S. Neuer, Y. Nojiri and C.S. Wong, 2003: Temporal studies of biogeochemical processes determined from ocean time-series observations during the JGOFS era. In: *Ocean Biogeochemistry: The Role of the Ocean Carbon Cycle in Global Change* [Fasham, M.J.R. (ed.)]. Springer, Berlin, Germany, pp. 239-267.
- Karl**, D.M., R.R. Bidigare and R.M. Letelier, 2001: Long-term changes in plankton community structure and productivity in the North Pacific Subtropical Gyre: The domain shift hypothesis. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **48(8-9)**, 1449-1470.
- Karstensen**, J., L. Stramma and M. Visbeck, 2008: Oxygen minimum zones in the eastern tropical Atlantic and Pacific oceans. *Progress in Oceanography*, **77(4)**, 331-350.
- Kaschner**, K., D.P. Tittensor, J. Ready, T. Gerrodette and B. Worm, 2011: Current and future patterns of global marine mammal biodiversity. *PLoS ONE*, **6(5)**, e19653.
- Kashefi**, K. and D.R. Lovley, 2003: Extending the upper temperature limit for life. *Science*, **301(5635)**, 934.
- Katsikatsou**, M., A. Anestis, H.-O. Pörtner, A. Vratsistas, K. Aligizaki and B. Michaelidis, 2012: Field studies and projections of climate change effects on the bearded horse mussel *Modiolus barbatus* in the Gulf of Thermaikos, Greece. *Marine Ecology Progress Series*, **449**, 183-196.
- Keeling**, C.D., S.C. Piper, R.B. Bacastow, M. Wahlen, T.P. Whorf, M. Heimann and H.A. Meijer, 2005: Atmospheric CO₂ and ¹³CO₂ exchange with the terrestrial biosphere and oceans from 1978 to 2000: observations and carbon cycle implications. In: *A History of Atmospheric CO₂ and its Effects on Plants, Animals, and Ecosystems* [Baldwin, I.T., M.M. Caldwell, G. Heldmaier, R.B. Jackson, O.L. Lange, H.A. Mooney, E.-D. Schulze and U. Sommer (eds.)]. Springer, New York, NY, USA, pp. 83-113.
- Keeling**, R.F., A. Körtzinger and N. Gruber, 2010: Ocean deoxygenation in a warming world. *Annual Review of Marine Science*, **2(1)**, 199-229.
- Kelly**, M.W., E. Sanford and R.K. Grosberg, 2012: Limited potential for adaptation to climate change in a broadly distributed marine crustacean. *Proceedings of the Royal Society B: Biological Sciences*, **279(1727)**, 349-356.
- Kennett**, J.P. and L.D. Stott, 1991: Abrupt deep-sea warming, palaeoceanographic changes and benthic extinctions at the end of the Paleocene. *Nature*, **353(6341)**, 225-229.
- Kiessling**, W. and C. Simpson, 2011: On the potential for ocean acidification to be a general cause of ancient reef crises. *Global Change Biology*, **17(1)**, 56-67.
- Kiessling**, W., C. Simpson, B. Beck, H. Mewis and J.M. Pandolfi, 2012: Equatorial decline of reef corals during the last Pleistocene interglacial. *Proceedings of the National Academy of Sciences of the United States of America*, **109(52)**, 21378-21383.

- Kim, J.-M., K. Lee, K. Shin, E.J. Yang, A. Engel, D.M. Karl and H.-C. Kim, 2011:** Shifts in biogenic carbon flow from particulate to dissolved forms under high carbon dioxide and warm ocean conditions. *Geophysical Research Letters*, **38(8)**, L08612.
- Kirby, R.R. and G. Beaugrand, 2009:** Trophic amplification of climate warming. *Proceedings of the Royal Society B: Biological Sciences*, **276(1676)**, 4095-4103.
- Kirby, R.R., G. Beaugrand and J.A. Lindley, 2009:** Synergistic effects of climate and fishing in a marine ecosystem. *Ecosystems*, **12(4)**, 548-561.
- Kirchman, D.L., X.A. Morán and H. Ducklow, 2009:** Microbial growth in the polar oceans - role of temperature and potential impact of climate change. *Nature Reviews Microbiology*, **7(6)**, 451-459.
- Kirk, J.T.O., 1994:** *Light and Photosynthesis in Aquatic Ecosystems*. Cambridge University Press, 662 pp.
- Klaas, C. and D.E. Archer, 2002:** Association of sinking organic matter with various types of mineral ballast in the deep sea: Implications for the rain ratio. *Global Biogeochemical Cycles*, **16(4)**, 1116.
- Kleypas, J.A. and C. Langdon, 2006:** Coral reefs and changing seawater chemistry. In: *Coral Reefs and Climate Change: Science and Management* [Phinney, J., O. Hoegh-Guldberg, J. Kleypas, W. Skirving and A.E. Strong (eds.)]. American Geophysical Union, Washington, D.C., USA, pp. 73-110.
- Knies, J.L., R. Izem, K.L. Supler, J.G. Kingsolver and C.L. Burch, 2006:** The genetic basis of thermal reaction norm evolution in lab and natural phage populations. *PLoS Biology*, **4(7)**, e201.
- Knoll, A., R.K. Bambach, J.L. Payne, S. Pruss and W.W. Fischer, 2007:** Paleophysiology and end-Permian mass extinction. *Earth and Planetary Science Letters*, **256(3-4)**, 295-313.
- Knoll, A.H. and W.W. Fischer, 2011:** 4- Skeletons and ocean chemistry: the long view. In: *Ocean Acidification* [Gattuso, J.-P. and L. Hansson (eds.)]. Oxford University Press, Oxford, pp. 67-82.
- Köhler, P., J. Hartmann and D.A. Wolf-Gladrow, 2010:** Geoengineering potential of artificially enhanced silicate weathering of olivine. *Proceedings of the National Academy of Sciences of the United States of America*, **107(47)**, 20228-20233.
- Koslow, J.A., R. Goericke, A. Lara-Lopez and W. Watson, 2011:** Impact of declining intermediate-water oxygen on deepwater fishes in the California Current. *Marine Ecology Progress Series*, **436**, 207-218.
- Kovach, R.P., A.J. Gharrett and D.A. Tallmon, 2012:** Genetic change for earlier migration timing in a pink salmon population. *Proceedings of the Royal Society B: Biological Sciences*, **279(1743)**, 3870-3878.
- Kovats, R.S., M.J. Bouma, S. Hajat, E. Worrall and A. Haines, 2003:** El Niño and health. *The Lancet*, **362(9394)**, 1481-1489.
- Kranz, S., M. Eichner and B. Rost, 2011:** Interactions between CCM and N₂ fixation in *Trichodesmium*. *Photosynthesis Research*, **109(1-3)**, 73-84.
- Kranz, S.A., O. Levitan, K.U. Richter, O. Prášil, I. Berman-Frank and B. Rost, 2010:** Combined effects of CO₂ and light on the N₂-fixing cyanobacterium *Trichodesmium* IMS101: physiological responses. *Plant Physiology*, **154(1)**, 334-345.
- Krause, E., A. Wichels, L. Giménez, M. Lunau, M.B. Schilhabel and G. Gerdt, 2012:** Small changes in pH have direct effects on marine bacterial community composition: a microcosm approach. *PLoS ONE*, **7(10)**, e47035.
- Kroeker, K.J., F. Micheli, M.C. Gambi and T.R. Martz, 2011:** Divergent ecosystem responses within a benthic marine community to ocean acidification. *Proceedings of the National Academy of Sciences of the United States of America*, **108(35)**, 14515-14520.
- Kroeker, K.J., R.L. Kordas, R. Crim, I.E. Hendriks, L. Ramajo, G.S. Singh, C.M. Duarte and J.-P. Gattuso, 2013:** Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. *Global Change Biology*, **19(6)**, 1884-1896.
- Kübler, J.E. and I.R. Davison, 1995:** Thermal acclimation of light use characteristics of *Chondrus crispus* (Rhodophyta). *European Journal of Phycology*, **30(3)**, 189-195.
- Kurihara, H. and Y. Shirayama, 2004:** Effects of increased atmospheric CO₂ on sea urchin early development. *Marine Ecology Progress Series*, **274**, 161-169.
- La Sorte, F.A. and W. Jetz, 2010:** Avian distributions under climate change: towards improved projections. *Journal of Experimental Biology*, **213(6)**, 862-869.
- Lafferty, K.D., 2009:** Calling for an ecological approach to studying climate change and infectious diseases. *Ecology*, **90(4)**, 932-933.
- Laidre, K.L., I. Stirling, L.F. Lowry, O. Wiig, M.P. Heide-Jørgensen and S.H. Ferguson, 2008:** Quantifying the sensitivity of Arctic marine mammals to climate-induced habitat change. *Ecological Applications*, **18(2 Suppl)**, S97-S125.

- Lam, P.J. and J.K.B. Bishop, 2008: The continental margin is a key source of iron to the HNLC North Pacific Ocean. *Geophysical Research Letters*, **35(7)**, L07608.
- Lambert, E., C. Hunter, G.J. Pierce and C.D. MacLeod, 2010: Sustainable whale-watching tourism and climate change: towards a framework of resilience. *Journal of Sustainable Tourism*, **18(3)**, 409-427.
- Langdon, C. and M.J. Atkinson, 2005: Effect of elevated $p\text{CO}_2$ on photosynthesis and calcification of corals and interactions with seasonal change in temperature/irradiance and nutrient enrichment. *Journal of Geophysical Research*, **110(C9)**, C09S07.
- Langenbuch, M. and H.-O. Pörtner, 2002: Changes in metabolic rate and N excretion in the marine invertebrate *Sipunculus nudus* under conditions of environmental hypercapnia: identifying effective acid-base variables. *Journal of Experimental Biology*, **205(8)**, 1153-1160.
- Langenbuch, M. and H.-O. Pörtner, 2003: Energy budget of hepatocytes from Antarctic fish (*Pachycara brachycephalum* and *Lepidonotothen kempfi*) as a function of ambient CO_2 : pH-dependent limitations of cellular protein biosynthesis? *Journal of Experimental Biology*, **206(22)**, 3895-3903.
- Langenbuch, M., C. Bock, D. Leibfritz and H.-O. Pörtner, 2006: Effects of environmental hypercapnia on animal physiology: A ^{13}C NMR study of protein synthesis rates in the marine invertebrate *Sipunculus nudus*. *Comparative Biochemistry and Physiology A: Molecular and Integrative Physiology*, **144(4)**, 479-484.
- Langer, G., I. Probert, G. Nehrke and P. Ziveri, 2011: The morphological response of *Emiliania huxleyi* to seawater carbonate chemistry changes: an inter-strain comparison. *Journal of Nannoplankton Research*, **32(1)**, 27-32.
- Langer, G., G. Nehrke, I. Probert, J. Ly and P. Ziveri, 2009: Strain-specific responses of *Emiliania huxleyi* to changing seawater carbonate chemistry. *Biogeosciences*, **6(11)**, 2637-2646.
- Langer, G., M. Geisen, K.-H. Baumann, J. Kläs, U. Riebesell, S. Thoms and J.R. Young, 2006: Species-specific responses of calcifying algae to changing seawater carbonate chemistry. *Geochemistry Geophysics Geosystems*, **7(9)**, Q09006.
- Lauer, A., V. Eyring, J.J. Corbett, C.F. Wang and J.J. Winebrake, 2009: Assessment of near-future policy instruments for oceangoing shipping: impact on atmospheric aerosol burdens and the Earth's radiation budget. *Environmental Science & Technology*, **43(15)**, 5592-5598.
- Lavaniegos, B.E. and M.D. Ohman, 2003: Long-term changes in pelagic tunicates of the California Current. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **50(14-16)**, 2473-2498.
- Law, C.S., 2008: Predicting and monitoring the effects of large-scale ocean iron fertilization on marine trace gas emissions. *Marine Ecology Progress Series*, **364**, 283-288.
- Law, C.S., E. Breitbarth, L.J. Hoffmann, C.M. McGraw, R.J. Langlois, J. LaRoche, A. Marriner and K.A. Safi, 2012: No stimulation of nitrogen fixation by non-filamentous diazotrophs under elevated CO_2 in the South Pacific. *Global Change Biology*, **18(10)**, 3004-3014.
- Le Borgne, R., V. Allain, S.P. Griffiths, R.J. Matear, A.D. McKinnon, A.J. Richardson and J.W. Young, 2011: Vulnerability of oceanic food webs in the tropical Pacific to climate change. In: *Vulnerability of Tropical Pacific Fisheries and Aquaculture to Climate Change* [Bell, J.D., J.E. Johnson and A.J. Hobday (eds.)]. Secretariat of the Pacific Community, Noumea, New Caledonia, pp. 189-250.
- Le Quesne, W.J.F. and J.K. Pinnegar, 2012: The potential impacts of ocean acidification: scaling from physiology to fisheries. *Fish and Fisheries*, **13(3)**, 333-344.
- Leckie, R.M., T.J. Bralower and R. Cashman, 2002: Oceanic anoxic events and planktonic evolution: biotic response to tectonic forcing during the mid-Cretaceous. *Paleoceanography*, **17(3)**, doi:10.1029/2001PA000623.
- Leclercq, N., J.-P. Gattuso and J. Jaubert, 2002: Primary production, respiration, and calcification of a coral reef mesocosm under increased CO_2 partial pressure. *Limnology and Oceanography*, **47(2)**, 558-564.
- Lehodey, P., 2000: *Impacts of the El Niño Southern Oscillation on tuna populations and fisheries in the tropical Pacific Ocean, SCTB13 Working Paper RG-1*. 13th Meeting of the Standing Committee on Tuna and Billfish, Noumea, New Caledonia, 5-12 July 2000, Secretariat of the Pacific Community, pp. 1-32.
- Lehodey, P., J. Hampton, R.W. Brill, S. Nicol, I. Senina, B. Calmetters, H.-O. Pörtner, L. Bopp, T. Llyina, J.D. Bell and J. Sibert, 2011: Vulnerability of oceanic fisheries in the tropical Pacific to climate change. In: *Vulnerability of Tropical Pacific Fisheries and Aquaculture to Climate Change* [Bell, J.D., J.E. Johnson and A.J. Hobday (eds.)]. Secretariat of the Pacific Community, Noumea, New Caledonia, pp. 433-492.
- Lenoir, S., G. Beaugrand and É. Lecuyer, 2011: Modelled spatial distribution of marine fish and projected modifications in the North Atlantic Ocean. *Global Change Biology*, **17(1)**, 115-129.
- Levin, L.A., 2003: Oxygen minimum zone benthos: adaptation and community response to hypoxia. *Oceanography and Marine Biology: an Annual Review*, **41**, 1-45.

- Levin, L.A.** and M. Sibuet, 2012: Understanding continental margin biodiversity: a new imperative. *Annual Review of Marine Science*, **4(1)**, 79-112.
- Levin, L.A.**, W. Ekau, A.J. Gooday, F. Jorissen, J.J. Middelburg, S.W.A. Naqvi, C. Neira, N.N. Rabalais and J. Zhang, 2009: Effects of natural and human-induced hypoxia on coastal benthos. *Biogeosciences*, **6(10)**, 2063-2098.
- Levitan, O.**, S.A. Kranz, D. Spungin, O. Prášil, B. Rost and I. Berman-Frank, 2010: Combined effects of CO₂ and light on the N₂-fixing cyanobacterium *Trichodesmium* IMS101: a mechanistic view. *Plant Physiology*, **154(1)**, 346-356.
- Levitan, O.**, G. Rosenberg, I. Setlik, E. Setlikova, J. Grigel, J. Klepetar, O. Prášil and I. Berman-Frank, 2007: Elevated CO₂ enhances nitrogen fixation and growth in the marine cyanobacterium *Trichodesmium*. *Global Change Biology*, **13(2)**, 531-538.
- Lewandowska, A.** and U. Sommer, 2010: Climate change and the spring bloom: a mesocosm study on the influence of light and temperature on phytoplankton and mesozooplankton. *Marine Ecology Progress Series*, **405**, 101-111.
- Lewis, P.N.**, M.J. Riddle and C.L. Hewitt, 2004: Management of exogenous threats to Antarctica and the sub-Antarctic islands: balancing risks from TBT and non-indigenous marine organisms. *Marine Pollution Bulletin*, **49(11-12)**, 999-1005.
- Liggett, D.**, A. McIntosh, A. Thompson, N. Gilbert and B. Storey, 2011: From frozen continent to tourism hotspot? Five decades of Antarctic tourism development and management, and a glimpse into the future. *Tourism Management*, **32(2)**, 357-366.
- Lima, F.P.**, P.A. Ribeiro, N. Queiroz, S.J. Hawkins and A.M. Santos, 2007: Do distributional shifts of northern and southern species of algae match the warming pattern? *Global Change Biology*, **13(12)**, 2592-2604.
- Lindgren, M.**, C. Möllmann, A. Nielsen, K. Brander, B.R. MacKenzie and N.C. Stenseth, 2010: Ecological forecasting under climate change: the case of Baltic cod. *Proceedings of the Royal Society B: Biological Sciences*, **277(1691)**, 2121-2130.
- Lindenmayer, D.B.**, G.E. Likens, C.J. Krebs and R.J. Hobbs, 2010: Improved probability of detection of ecological "surprises". *Proceedings of the National Academy of Sciences of the United States of America*, **107(51)**, 21957-21962.
- Lindley, J.A.**, G. Beaugrand, C. Luczak, J.M. Dewarumez and R.R. Kirby, 2010: Warm-water decapods and the trophic amplification of climate in the North Sea. *Biology Letters*, **6(6)**, 773-776.
- Lipp, E.K.**, A. Huq and R.R. Colwell, 2002: Effects of global climate on infectious disease: the cholera model. *Clinical Microbiology Reviews*, **15(4)**, 757-770.
- Lischka, S.**, J. Büdenbender, T. Boxhammer and U. Riebesell, 2011: Impact of ocean acidification and elevated temperatures on early juveniles of the polar shelled pteropod *Limacina helicina*: mortality, shell degradation, and shell growth. *Biogeosciences*, **8(4)**, 919-932.
- Liu, J.**, M.G. Weinbauer, C. Maier, M.H. Dai and J.-P. Gattuso, 2010: Effect of ocean acidification on microbial diversity and on microbe-driven biogeochemistry and ecosystem functioning. *Aquatic Microbial Ecology*, **61(3)**, 291-305.
- Liu, W.** and M. He, 2012: Effects of ocean acidification on the metabolic rates of three species of bivalve from southern coast of China. *Chinese Journal of Oceanology and Limnology*, **30(2)**, 206-211.
- Llewellyn, L.E.**, 2010: Revisiting the association between sea surface temperature and the epidemiology of fish poisoning in the South Pacific: reassessing the link between ciguatera and climate change. *Toxicon*, **56(5)**, 691-697.
- Lloret, J.** and H.-J. Rätz, 2000: Condition of cod (*Gadus morhua*) off Greenland during 1982–1998. *Fisheries Research*, **48(1)**, 79-86.
- Lobitz, B.**, L. Beck, A. Huq, B. Wood, G. Fuchs, A.S.G. Faruque and R. Colwell, 2000: Climate and infectious disease: Use of remote sensing for detection of *Vibrio cholerae* by indirect measurement. *Proceedings of the National Academy of Sciences of the United States of America*, **97(4)**, 1438-1443.
- Lohbeck, K.T.**, U. Riebesell and T.B.H. Reusch, 2012: Adaptive evolution of a key phytoplankton species to ocean acidification. *Nature Geoscience*, **5(5)**, 346-351.
- Lomas, M.W.**, B.M. Hopkinson, J.L. Losh, D.E. Ryan, D.L. Shi, Y. Xu and F.M.M. Morel, 2012: Effect of ocean acidification on cyanobacteria in the subtropical North Atlantic. *Aquatic Microbial Ecology*, **66(3)**, 211-222.
- Lombard, F.**, R.E. da Rocha, J. Bijma and J.-P. Gattuso, 2010: Effect of carbonate ion concentration and irradiance on calcification in planktonic foraminifera. *Biogeosciences*, **7(1)**, 247–255.

- Lovelock, J.E.** and C.G. Rapley, 2007: Ocean pipes could help the Earth to cure itself. *Nature*, **449(7161)**, 403.
- Loya, Y., K. Sakai, K. Yamazato, Y. Nakano, H. Sambali and R. van Woesik**, 2001: Coral bleaching: the winners and the losers. *Ecology Letters*, **4(2)**, 122-131.
- Luczak, C., G. Beaugrand, M. Jaffré and S. Lenoir**, 2011: Climate change impact on Balearic shearwater through a trophic cascade. *Biology Letters*, **7(5)**, 702-705.
- Maas, A.E., K.F. Wishner and B.A. Seibel**, 2012: The metabolic response of pteropods to ocean acidification reflects natural CO₂-exposure in oxygen minimum zones. *Biogeosciences*, **9(2)**, 747-757.
- Mackas, D.L.**, 2011: Does blending of chlorophyll data bias temporal trend? *Nature*, **472(7342)**, E4-E5.
- Mackas, D.L., R.H. Goldblatt and A.G. Lewis**, 1998: Interdecadal variation in developmental timing of *Neocalanus plumchrus* populations at Ocean Station P in the subarctic North Pacific. *Canadian Journal of Fisheries and Aquatic Sciences*, **55(8)**, 1878-1893.
- MacLeod, C.D.**, 2009: Global climate change, range changes and potential implications for the conservation of marine cetaceans: a review and synthesis. *Endangered Species Research*, **7**, 125-136.
- MacLeod, C.D., S.M. Bannon, G.J. Pierce, C. Schweder, J.A. Learmonth, J.S. Herman and R.J. Reid**, 2005: Climate change and the cetacean community of north-west Scotland. *Biological Conservation*, **124(4)**, 477-483.
- Maier, C., J. Hegeman, M.G. Weinbauer and J.-P. Gattuso**, 2009: Calcification of the cold-water coral *Lophelia pertusa* under ambient and reduced pH. *Biogeosciences*, **6(8)**, 1671-1680.
- Maier, C., A. Schubert, M.M. Berzunza Sánchez, M.G. Weinbauer, P. Watremez and J.-P. Gattuso**, 2013: End of the century pCO₂ levels do not impact calcification in mediterranean cold-water corals. *PLoS ONE*, **8(4)**, e62655.
- Manzello, D.P., J.A. Kleypas, D.A. Budd, C.M. Eakin, P.W. Glynn and C. Langdon**, 2008: Poorly cemented coral reefs of the eastern tropical Pacific: possible insights into reef development in a high-CO₂ world. *Proceedings of the National Academy of Sciences of the United States of America*, **105(30)**, 10450-10455.
- Marañón, E., P. Cermeño, M. Latasa and R.D. Tadonlécé**, 2012: Temperature, resources, and phytoplankton size structure in the ocean. *Limnology and Oceanography*, **57(5)**, 1266-1278.
- Marbà, N. and C.M. Duarte**, 2010: Mediterranean warming triggers seagrass (*Posidonia oceanica*) shoot mortality. *Global Change Biology*, **16(8)**, 2366-2375.
- Margalef, R.**, 1978: Life-forms of phytoplankton as survival alternatives in an unstable environment. *Oceanologica Acta*, **1(4)**, 493-509.
- MARGO Project Members**, 2009: Constraints on the magnitude and patterns of ocean cooling at the Last Glacial Maximum. *Nature Geoscience*, **2(2)**, 127-132.
- Mark, F.C., C. Bock and H.-O. Pörtner**, 2002: Oxygen-limited thermal tolerance in Antarctic fish investigated by MRI and ³¹P-MRS. *American Journal of Physiology: Regulatory, Integrative and Comparative Physiology*, **283(5)**, R1254-1262.
- Martin, S. and J.-P. Gattuso**, 2009: Response of Mediterranean coralline algae to ocean acidification and elevated temperature. *Global Change Biology*, **15(8)**, 2089-2100.
- Martin, S., R. Rodolfo-Metalpa, E. Ransome, S. Rowley, M.C. Buia, J.-P. Gattuso and J. Hall-Spencer**, 2008: Effects of naturally acidified seawater on seagrass calcareous epibionts. *Biology Letters*, **4(6)**, 689-692.
- Martin, S., S. Richier, M.-L. Pedrotti, S. Dupont, C. Castejon, Y. Gerakis, M.-E. Kerros, F. Oberhansli, J.-L. Teysse, R. Jeffree and J.-P. Gattuso**, 2011: Early development and molecular plasticity in the Mediterranean sea urchin *Paracentrotus lividus* exposed to CO₂-driven acidification. *Journal of Experimental Biology*, **214(8)**, 1357-1368.
- Martrat, B., J.O. Grimalt, C. Lopez-Martinez, I. Cacho, F.J. Sierro, J.A. Flores, R. Zahn, M. Canals, J.H. Curtis and D.A. Hodell**, 2004: Abrupt temperature changes in the Western Mediterranean over the past 250,000 years. *Science*, **306(5702)**, 1762-1765.
- Matear, R.J. and A.C. Hirst**, 1999: Climate change feedback on the future oceanic CO₂ uptake. *Tellus Series B-Chemical and Physical Meteorology*, **51(3)**, 722-733.
- Mazaris, A.D., A.S. Kallimanis, S.P. Sgardelis and J.D. Pantis**, 2008: Do long-term changes in sea surface temperature at the breeding areas affect the breeding dates and reproduction performance of Mediterranean loggerhead turtles? Implications for climate change. *Journal of Experimental Marine Biology and Ecology*, **367(2)**, 219-226.
- Mazaris, A.D., A.S. Kallimanis, J. Tzanopoulos, S.P. Sgardelis and J.D. Pantis**, 2009: Sea surface temperature variations in core foraging grounds drive nesting trends and phenology of loggerhead turtles in the Mediterranean Sea. *Journal of Experimental Marine Biology and Ecology*, **379(1-2)**, 23-27.

- McBryan**, T.L., K. Anttila, T.M. Healy and P.M. Schulte, 2013: Responses to temperature and hypoxia as interacting stressors in fish: implications for adaptation to environmental change. *Integrative and Comparative Biology*, **53**(4), 648-659.
- McClain**, C.R., 2009: A decade of satellite ocean color observations. *Annual Review of Marine Science*, **1**(1), 19-42.
- McClatchie**, S., R. Goericke, R. Cosgrove, G. Auad and R. Vetter, 2010: Oxygen in the Southern California Bight: multidecadal trends and implications for demersal fisheries. *Geophysical Research Letters*, **37**, L19602.
- McCulloch**, M., J. Falter, J. Trotter and P. Montagna, 2012: Coral resilience to ocean acidification and global warming through pH up-regulation. *Nature Climate Change*, **2**(8), 623-627.
- McDaniel**, L.D., E. Young, J. Delaney, F. Ruhnau, K.B. Ritchie and J.H. Paul, 2010: High frequency of horizontal gene transfer in the oceans. *Science*, **330**(6000), 50.
- McIntyre**, T., I.J. Ansorge, H. Bornemann, J. Plötz, C.A. Tosh and M.N. Bester, 2011: Elephant seal dive behaviour is influenced by ocean temperature: implications for climate change impacts on an ocean predator. *Marine Ecology Progress Series*, **441**, 257-272.
- McLeod**, E., K.R.N. Anthony, A. Andersson, R. Beeden, Y. Golbuu, J. Kleypas, K. Kroeker, D. Manzello, R.V. Salm, H. Schuttenberg and J.E. Smith, 2013: Preparing to manage coral reefs for ocean acidification: lessons from coral bleaching. *Frontiers in Ecology and the Environment*, **11**(1), 20-27.
- McMahon**, C.R. and G.C. Hays, 2006: Thermal niche, large-scale movements and implications of climate change for a critically endangered marine vertebrate. *Global Change Biology*, **12**(7), 1330-1338.
- McQuatters-Gollop**, A., P.C. Reid, M. Edwards, P.H. Burkill, C. Castellani, S. Batten, W. Gieskes, D. Beare, R.R. Bidigare, E. Head, R. Johnson, M. Kahru, J.A. Koslow and A. Pena, 2011: Is there a decline in marine phytoplankton? *Nature*, **472**(7342), E6-E7.
- Meinshausen**, M., S.J. Smith, K. Calvin, J.S. Daniel, M.L.T. Kainuma, J.F. Lamarque, K. Matsumoto, S.A. Montzka, S.C.B. Raper, K. Riahi, A. Thomson, G.J.M. Velders and D.P.P. van Vuuren, 2011: The RCP greenhouse gas concentrations and their extensions from 1765 to 2300. *Climatic Change*, **109**(1-2), 213-241.
- Meissner**, K.J., T. Lippmann and A. Sen Gupta, 2012: Large-scale stress factors affecting coral reefs: open ocean sea surface temperature and surface seawater aragonite saturation over the next 400 years. *Coral Reefs*, **31**(2), 309-319.
- Melzner**, F., S. Göbel, M. Langenbuch, M.A. Gutowska, H.-O. Pörtner and M. Lucassen, 2009: Swimming performance in Atlantic Cod (*Gadus morhua*) following long-term (4-12 months) acclimation to elevated seawater P(CO₂). *Aquatic Toxicology*, **92**(1), 30-37.
- Melzner**, F., P. Stange, K. Trubenbach, J. Thomsen, I. Casties, U. Panknin, S.N. Gorb and M.A. Gutowska, 2011: Food supply and seawater pCO₂ impact calcification and internal shell dissolution in the blue mussel *Mytilus edulis*. *PLoS ONE*, **6**(9), e24223.
- Merico**, A., T. Tyrrell, E.J. Lessard, T. Oguz, P.J. Stabeno, S.I. Zeeman and T.E. Whitledge, 2004: Modelling phytoplankton succession on the Bering Sea shelf: role of climate influences and trophic interactions in generating *Emiliania huxleyi* blooms 1997-2000. *Deep-Sea Research Part I: Oceanographic Research Papers*, **51**(12), 1803-1826.
- Metzger**, R.A. and G. Benford, 2001: Sequestering of atmospheric carbon through permanent disposal of crop residue. *Climatic Change*, **49**(1-2), 11-19.
- Michaelidis**, B., C. Ouzounis, A. Palaras and H.-O. Pörtner, 2005: Effects of long-term moderate hypercapnia on acid-base balance and growth rate in marine mussels *Mytilus galloprovincialis*. *Marine Ecology Progress Series*, **293**, 109-118.
- Milazzo**, M., S. Mirto, P. Domenici and M. Gristina, 2013: Climate change exacerbates interspecific interactions in sympatric coastal fishes. *Journal of Animal Ecology*, **82**(2), 468-477.
- Miller**, A.W., A.C. Reynolds, C. Sobrino and G.F. Riedel, 2009: Shellfish face uncertain future in high CO₂ world: influence of acidification on oyster larvae calcification and growth in estuaries. *PLoS ONE*, **4**(5), e5661.
- Millero**, F.J., 1995: Thermodynamics of the carbon dioxide system in the oceans. *Geochimica et Cosmochimica Acta*, **59**(4), 661-677.
- Mills**, C.E., 2001: Jellyfish blooms: are populations increasing globally in response to changing ocean conditions? *Hydrobiologia*, **451**, 55-68.
- Milly**, P.C.D., J. Betancourt, M. Falkenmark, R.M. Hirsch, Z.W. Kundzewicz, D.P. Lettenmaier and R.J. Stouffer, 2008: Stationarity is dead: whither water management? *Science*, **319**(5863), 573-574.
- Molinero**, J.B., F. Ibanez, P. Nival, E. Buecher and S. Souissi, 2005: North Atlantic climate and northwestern Mediterranean plankton variability. *Limnology and Oceanography*, **50**(4), 1213-1220.

- Moloney, C.L., M.A. St John, K.L. Denman, D.M. Karl, F.W. Köster, S. Sundby and R.P. Wilson, 2011:** Weaving marine food webs from end to end under global change. *Journal of Marine Systems*, **84(3-4)**, 106-116.
- Moore, J.K., S.C. Doney, D.M. Glover and I.Y. Fung, 2002:** Iron cycling and nutrient-limitation patterns in surface waters of the World Ocean. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **49(1-3)**, 463-507.
- Moore, S.E. and H.P. Huntington, 2008:** Arctic marine mammals and climate change: impacts and resilience. *Ecological Applications*, **18(2)**, S157-S165.
- Moore, W.R., 2010:** The impact of climate change on Caribbean tourism demand. *Current Issues in Tourism*, **13(5)**, 495-505.
- Mora, C., C.-L. Wei, A. Rollo, T. Amaro, A.R. Baco, D. Billett, L. Bopp, Q. Chen, M. Collier, R. Danovaro, A.J. Gooday, B.M. Grube, P.R. Halloran, J. Ingels, D.O.B. Jones, L.A. Levin, H. Nakano, K. Norling, E. Ramirez-Llodra, M. Rex, H.A. Ruhl, C.R. Smith, A.K. Sweetman, A.R. Thurber, J.F. Tjiputra, P. Usseglio, L. Watling, T. Wu and M. Yasuhara, 2013 in press:** Biotic and human vulnerability to projected changes in ocean biogeochemistry over the 21st century. *PLoS Biology*.
- Morán, X.A.G., Á. López-Urrutia, A. Calvo-Díaz and W.K.W. Li, 2010:** Increasing importance of small phytoplankton in a warmer ocean. *Global Change Biology*, **16(3)**, 1137-1144.
- Moss, R.H., J.A. Edmonds, K.A. Hibbard, M.R. Manning, S.K. Rose, D.P. van Vuuren, T.R. Carter, S. Emori, M. Kainuma, T. Kram, G.A. Meehl, J.F.B. Mitchell, N. Nakicenovic, K. Riahi, S.J. Smith, R.J. Stouffer, A.M. Thomson, J.P. Weyant and T.J. Wilbanks, 2010:** The next generation of scenarios for climate change research and assessment. *Nature*, **463(7282)**, 747-756.
- Mountain, D.G. and J. Kane, 2010:** Major changes in the Georges Bank ecosystem, 1980s to the 1990s. *Marine Ecology Progress Series*, **398**, 81-91.
- Mouriño-Carballido, B., R. Graña, A. Fernández, A. Bode, M. Varela, J.F. Domínguez, J. Escánez, D. de Armas and E. Marañón, 2011:** Importance of N₂ fixation vs. nitrate eddy diffusion along a latitudinal transect in the Atlantic Ocean. *Limnology and Oceanography*, **56(3)**, 999-1007.
- Moy, A.D., W.R. Howard, S.G. Bray and T.W. Trull, 2009:** Reduced calcification in modern Southern Ocean planktonic foraminifera. *Nature Geoscience*, **2(4)**, 276-280.
- Moya, A., L. Huisman, E.E. Ball, D.C. Hayward, L.C. Grasso, C.M. Chua, H.N. Woo, J.-P. Gattuso, S. Forêt and D.J. Miller, 2012:** Whole transcriptome analysis of the coral *Acropora millepora* reveals complex responses to CO₂-driven acidification during the initiation of calcification. *Molecular Ecology*, **21(10)**, 2440-2454.
- Müller, R., T. Laepple, I. Bartsch and C. Wiencke, 2009:** Impact of oceanic warming on the distribution of seaweeds in polar and cold-temperate waters. *Botanica Marina*, **52(6)**, 617-638.
- Munday, P.L., N.E. Crawley and G.E. Nilsson, 2009b:** Interacting effects of elevated temperature and ocean acidification on the aerobic performance of coral reef fishes. *Marine Ecology Progress Series*, **388**, 235-242.
- Munday, P.L., J.M. Donelson, D.L. Dixon and G.G. Endo, 2009a:** Effects of ocean acidification on the early life history of a tropical marine fish. *Proceedings of the Royal Society B: Biological Sciences*, **276(1671)**, 3275-3283.
- Munday, P.L., V. Hernaman, D.L. Dixon and S.R. Thorrold, 2011b:** Effect of ocean acidification on otolith development in larvae of a tropical marine fish. *Biogeosciences*, **8(2)**, 1631-1641.
- Munday, P.L., M. Gagliano, J.M. Donelson, D.L. Dixon and S.R. Thorrold, 2011a:** Ocean acidification does not affect the early life history development of a tropical marine fish. *Marine Ecology Progress Series*, **423**, 211-221.
- Munday, P.L., D.L. Dixon, M.I. McCormick, M. Meekan, M.C.O. Ferrari and D.P. Chivers, 2010:** Replenishment of fish populations is threatened by ocean acidification. *Proceedings of the National Academy of Sciences of the United States of America*, **107(29)**, 12930-12934.
- Naqvi, S.W.A., D.A. Jayakumar, P.V. Narvekar, H. Naik, V.V.S.S. Sarma, W. D'Souza, S. Joseph and M.D. George, 2000:** Increased marine production of N₂O due to intensifying anoxia on the Indian continental shelf. *Nature*, **408(6810)**, 346-349.
- Narita, D., K. Rehdanz and R.S.J. Tol, 2012:** Economic costs of ocean acidification: a look into the impacts on global shellfish production. *Climatic Change*, **113(3-4)**, 1049-1063.
- Navarro, J.M., R. Torres, K. Acuña, C. Duarte, P.H. Manriquez, M. Lardies, N.A. Lagos, C. Vargas and V. Aguilera, 2013:** Impact of medium-term exposure to elevated pCO₂ levels on the physiological energetics of the mussel *Mytilus chilensis*. *Chemosphere*, **90(3)**, 1242-1248.
- Neuheimer, A.B. and P. Grønkvær, 2012:** Climate effects on size-at-age: growth in warming waters compensates for earlier maturity in an exploited marine fish. *Global Change Biology*, **18(6)**, 1812-1822.

- Neuheimer, A.B., R.E. Thresher, J.M. Lyle and J.M. Semmens, 2011:** Tolerance limit for fish growth exceeded by warming waters. *Nature Climate Change*, **1(2)**, 110-113.
- Neutel, A.M., J.A.P. Heesterbeek, J. van de Koppel, G. Hoenderboom, A. Vos, C. Kaldewey, F. Berendse and P.C. de Ruiter, 2007:** Reconciling complexity with stability in naturally assembling food webs. *Nature*, **449**, 599-602.
- Nilsson, G.E., S. Östlund-Nilsson and P.L. Munday, 2010:** Effects of elevated temperature on coral reef fishes: loss of hypoxia tolerance and inability to acclimate. *Comparative Biochemistry and Physiology A: Molecular and Integrative Physiology*, **156(4)**, 389-393.
- Nilsson, G.E., N. Crawley, I.G. Lunde and P.L. Munday, 2009:** Elevated temperature reduces the respiratory scope of coral reef fishes. *Global Change Biology*, **15(6)**, 1405-1412.
- Nilsson, G.E., D.L. Dixon, P. Domenici, M.I. McCormick, C. Sørensen, S.-A. Watson and P.L. Munday, 2012:** Near-future carbon dioxide levels alter fish behaviour by interfering with neurotransmitter function. *Nature Climate Change*, **2(3)**, 201-204.
- NOAA, 2012:** NOAA Extended Reconstructed Sea Surface Temperature (SST) Version 3b. From: PSD Climate and Weather Data, NOAA/OAR/ESRL Physical Sciences Division, Boulder, CO, USA. From: url: <http://www.esrl.noaa.gov/psd/data/gridded/data.noaa.ersst.html>.
- Nuttall, M., 1998:** *Protecting the Arctic: Indigenous Peoples and Cultural Survival*. Routledge, London, 195 pp.
- O'Connor, M.I., M.F. Piehler, D.M. Leech, A. Anton and J.F. Bruno, 2009:** Warming and resource availability shift food web structure and metabolism. *PLoS Biology*, **7(8)**, e1000178.
- O'Donnell, M.J., A.E. Todgham, M.A. Sewell, L.M. Hammond, K. Ruggiero, N.A. Fanguie, M.L. Zippay and G.E. Hofmann, 2010:** Ocean acidification alters skeletogenesis and gene expression in larval sea urchins. *Marine Ecology Progress Series*, **398**, 157-171.
- Occhipinti-Ambrogi, A., 2007:** Global change and marine communities: alien species and climate change. *Marine Pollution Bulletin*, **55(7-9)**, 342-352.
- Ohman, M.D., B.E. Lavaniegos and A.W. Townsend, 2009:** Multi-decadal variations in calcareous holozooplankton in the California Current System: Thecosome pteropods, heteropods, and foraminifera. *Geophysical Research Letters*, **36**, L18608.
- Olafsson, J., S.R. Olafsdottir, A. Benoit-Cattin, M. Danielsen, T.S. Arnarson and T. Takahashi, 2009:** Rate of Iceland Sea acidification from time series measurements. *Biogeosciences*, **6(11)**, 2661-2668.
- Orr, J.C., V.J. Fabry, O. Aumont, L. Bopp, S.C. Doney, R.A. Feely, A. Gnanadesikan, N. Gruber, A. Ishida, F. Joos, R.M. Key, K. Lindsay, E. Maier-Reimer, R. Matear, P. Monfray, A. Mouchet, R.G. Najjar, G.K. Plattner, K.B. Rodgers, C.L. Sabine, J.L. Sarmiento, R. Schlitzer, R.D. Slater, I.J. Totterdell, M.F. Weirig, Y. Yamanaka and A. Yool, 2005:** Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature*, **437(7059)**, 681-686.
- Oschlies, A., M. Pahlow, A. Yool and R.J. Matear, 2010:** Climate engineering by artificial ocean upwelling: channelling the sorcerer's apprentice. *Geophysical Research Letters*, **37**, L04701.
- Österblom, H., S. Hansson, U. Larsson, O. Hjerne, F. Wulff, R. Elmgren and C. Folke, 2007:** Human-induced trophic cascades and ecological regime shifts in the Baltic Sea. *Ecosystems*, **10(6)**, 877-889.
- Ottersen, G., D.O. Hjermann and N.C. Stenseth, 2006:** Changes in spawning stock structure strengthen the link between climate and recruitment in a heavily fished cod (*Gadus morhua*) stock. *Fisheries Oceanography*, **15(3)**, 230-243.
- Ottersen, G., S. Kim, G. Huse, J.J. Polovina and N.C. Stenseth, 2010:** Major pathways by which climate may force marine fish populations. *Journal of Marine Systems*, **79(3-4)**, 343-360.
- Pagani, M., Z. Liu, J. LaRiviere and A.C. Ravelo, 2010:** High Earth-system climate sensitivity determined from Pliocene carbon dioxide concentrations. *Nature Geoscience*, **3(1)**, 27-30.
- Pakker, H., A.M. Breeman, W.F.P. van Reine and C. van den Hoek, 1995:** A comparative study of temperature responses of Caribbean seaweeds from different biogeographic groups. *Journal of Phycology*, **31(4)**, 499-507.
- Palacios, S.L. and R.C. Zimmerman, 2007:** Response of eelgrass *Zostera marina* to CO₂ enrichment: possible impacts of climate change and potential for remediation of coastal habitats. *Marine Ecology Progress Series*, **344**, 1-13.
- Pancost, R.D., N. Crawford, S. Magness, A. Turner, H.C. Jenkyns and J.R. Maxwell, 2004:** Further evidence for the development of photic-zone euxinic conditions during Mesozoic oceanic anoxic events. *Journal of the Geological Society*, **161**, 353-364.
- Pane, E.F. and J.P. Barry, 2007:** Extracellular acid-base regulation during short-term hypercapnia is effective in a shallow-water crab, but ineffective in a deep-sea crab. *Marine Ecology Progress Series*, **334**, 1-9.

- Parker, L.M., P.M. Ross and W.A. O'Connor**, 2011: Populations of the Sydney rock oyster, *Saccostrea glomerata*, vary in response to ocean acidification. *Marine Biology*, **158(3)**, 689-697.
- Parker, L.M., P.M. Ross, W.A. O'Connor, L. Borysko, D.A. Raftos and H.-O. Pörtner**, 2012: Adult exposure influences offspring response to ocean acidification in oysters. *Global Change Biology*, **18(1)**, 82-92.
- Parmesan, C. and J. Matthews**, 2005: Biological impacts of climate change. In: *Principles of Conservation Biology* [Groom, M.J., G.K. Meffe and C.R. Carroll (eds.)]. Sinauer, Sunderland, MA, pp. 333-374.
- Parmesan, C., C. Duarte, E. Poloczanska, A.J. Richardson and M.C. Singer**, 2011: Overstretching attribution. *Nature Climate Change*, **1(1)**, 2-4.
- Parsons, L.S. and W.H. Lear**, 2001: Climate variability and marine ecosystem impacts: a North Atlantic perspective. *Progress in Oceanography*, **49(1-4)**, 167-188.
- Pascual, M., X. Rodo, S.P. Ellner, R. Colwell and M.J. Bouma**, 2000: Cholera dynamics and El Niño-Southern Oscillation. *Science*, **289(5485)**, 1766-1769.
- Paulmier, A. and D. Ruiz-Pino**, 2009: Oxygen minimum zones (OMZs) in the modern ocean. *Progress in Oceanography*, **80(3-4)**, 113-128.
- Paulmier, A., D. Ruiz-Pino and V. Garçon**, 2011: CO₂ maximum in the oxygen minimum zone (OMZ). *Biogeosciences*, **8(2)**, 239-252.
- Pauly, D., V. Christensen, J. Dalsgaard, R. Froese and F. Torres Jr.**, 1998: Fishing down marine food webs. *Science*, **279(5352)**, 860-863.
- Peck, L.S., S.A. Morley and M.S. Clark**, 2010: Poor acclimation capacities in Antarctic marine ectotherms. *Marine Biology*, **157(9)**, 2051-2059.
- Péron, C., H. Weimerskirch and C.-A. Bost**, 2012: Projected poleward shift of king penguins' (*Aptenodytes patagonicus*) foraging range at the Crozet Islands, southern Indian Ocean. *Proceedings of the Royal Society B: Biological Sciences*, **279(1738)**, 2515-2523.
- Perry, A.L., P.J. Low, J.R. Ellis and J.D. Reynolds**, 2005: Climate change and distribution shifts in marine fishes. *Science*, **308(5730)**, 1912-1915.
- Perry, R.I., P. Cury, K. Brander, S. Jennings, C. Möllmann and B. Planque**, 2010: Sensitivity of marine systems to climate and fishing: concepts, issues and management responses. *Journal of Marine Systems*, **79(3-4)**, 427-435.
- Pershing, A.J., E.H.J. Head, C.H. Greene and J.W. Jossi**, 2010: Pattern and scale of variability among Northwest Atlantic Shelf plankton communities. *Journal of Plankton Research*, **32(12)**, 1661-1674.
- Pershing, A.J., C.H. Greene, J.W. Jossi, L. O'Brien, J.K.T. Brodziak and B.A. Bailey**, 2005: Interdecadal variability in the Gulf of Maine zooplankton community, with potential impacts on fish recruitment. *ICES Journal of Marine Science*, **62(7)**, 1511-1523.
- Pespeni, M.H., E. Sanford, B. Gaylord, T.M. Hill, J.D. Hofelt, H.K. Jaris, M. LaVigne, E.A. Lenz, A.D. Russell, M.K. Young and S.R. Palumbi**, 2013: Evolutionary change during experimental ocean acidification. *Proceedings of the National Academy of Sciences of the United States of America*, **110(17)**, 6937-6942.
- Petitgas, P., D. Reid, B. Planque, E. Nogueira, B. O'Hea and U. Cotano**, 2006: The entrainment hypothesis: an explanation for the persistence and innovation in spawning migrations and life cycle spatial patterns. In: *ICES Conference and Meeting Documents 2006/B:07*, [ICES (ed.)]. Maastricht, Netherlands, ICES, pp. 9.
- Philippart, C.J.M., R. Anadón, R. Danovaro, J.W. Dippner, K.F. Drinkwater, S.J. Hawkins, T. Oguz, G. O'Sullivan and P.C. Reid**, 2011: Impacts of climate change on European marine ecosystems: observations, expectations and indicators. *Journal of Experimental Marine Biology and Ecology*, **400(1-2)**, 52-69.
- Pierce, D.W., P.J. Gleckler, T.P. Barnett, B.D. Santer and P.J. Durack**, 2012: The fingerprint of human-induced changes in the ocean's salinity and temperature fields. *Geophysical Research Letters*, **39(21)**, L21704.
- Pike, D.A., R.L. Antworth and J.C. Stiner**, 2006: Earlier nesting contributes to shorter nesting seasons for the loggerhead sea turtle, *Caretta caretta*. *Journal of Herpetology*, **40(1)**, 91-94.
- Pinsky, M.L. and M. Fogarty**, 2012: Lagged social-ecological responses to climate and range shifts in fisheries. *Climatic Change*, **115(3-4)**, 883-891.
- Pinsky, M.L., B. Worm, M.J. Fogarty, J.L. Sarmiento and S.A. Levin**, 2013: Marine taxa track local climate velocities. *Science*, **341(6151)**, 1239-1242.
- Piontek, J., M. Lunau, N. Händel, C. Borchard, M. Wurst and A. Engel**, 2010: Acidification increases microbial polysaccharide degradation in the ocean. *Biogeosciences*, **7(5)**, 1615-1624.
- Pitchford, J.W. and J. Brindley**, 1999: Iron limitation, grazing pressure and oceanic high nutrient-low chlorophyll (HNLC) regions. *Journal of Plankton Research*, **21(3)**, 525-547.

- Planque, B., E. Bellier and C. Loots, 2011a:** Uncertainties in projecting spatial distributions of marine populations. *ICES Journal of Marine Science*, **68(6)**, 1045-1050.
- Planque, B., C. Loots, P. Petitgas, U. Lindstrøm and S. Vaz, 2011b:** Understanding what controls the spatial distribution of fish populations using a multi-model approach. *Fisheries Oceanography*, **20(1)**, 1-17.
- Planque, B., J.-M. Fromentin, P. Cury, K.F. Drinkwater, S. Jennings, R.I. Perry and S. Kifani, 2010:** How does fishing alter marine populations and ecosystems sensitivity to climate? *Journal of Marine Systems*, **79(3-4)**, 403-417.
- Poloczanska, E.S., S.J. Hawkins, A.J. Southward and M.T. Burrows, 2008:** Modeling the response of populations of competing species to climate change. *Ecology*, **89(11)**, 3138-3149.
- Poloczanska, E.S., C.J. Brown, W.J. Sydeman, W. Kiessling, D.S. Schoeman, P.J. Moore, K. Brander, J.F. Bruno, L.B. Buckley, M.T. Burrows, C.M. Duarte, B.S. Halpern, J. Holding, C.V. Kappel, M.I. O'Connor, J.M. Pandolfi, C. Parmesan, F. Schwing, S.A. Thompson and A.J. Richardson, 2013:** Global imprint of climate change on marine life. *Nature Climate Change*, **3(10)**, 919-925.
- Polovina, J.J., E.A. Howell and M. Abecassis, 2008:** Ocean's least productive waters are expanding. *Geophysical Research Letters*, **35(3)**, L03618.
- Polovina, J.J., J.P. Dunne, P.A. Woodworth and E.A. Howell, 2011:** Projected expansion of the subtropical biome and contraction of the temperate and equatorial upwelling biomes in the North Pacific under global warming. *ICES Journal of Marine Science*, **68(6)**, 986-995.
- Pörtner, H.-O., 2002a:** Climate variations and the physiological basis of temperature dependent biogeography: systemic to molecular hierarchy of thermal tolerance in animals. *Comparative Biochemistry and Physiology A: Molecular and Integrative Physiology*, **132(4)**, 739-761.
- Pörtner, H.-O., 2002b:** Environmental and functional limits to muscular exercise and body size in marine invertebrate athletes. *Comparative Biochemistry and Physiology A: Molecular and Integrative Physiology*, **133(2)**, 303-321.
- Pörtner, H.-O., 2006:** Climate-dependent evolution of Antarctic ectotherms: an integrative analysis. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **53(8-10)**, 1071-1104.
- Pörtner, H.-O., 2008:** Ecosystem effects of ocean acidification in times of ocean warming: a physiologist's view. *Marine Ecology Progress Series*, **373**, 203-217.
- Pörtner, H.-O., 2010:** Oxygen- and capacity-limitation of thermal tolerance: a matrix for integrating climate-related stressor effects in marine ecosystems. *Journal of Experimental Biology*, **213(6)**, 881-893.
- Pörtner, H.-O., 2012:** Integrating climate-related stressor effects on marine organisms: unifying principles linking molecule to ecosystem-level changes. *Marine Ecology Progress Series*, **470**, 273-290.
- Pörtner, H.-O. and M.K. Grieshaber, 1993:** Critical $P_{O_2}(s)$ in oxyconforming and oxyregulating animals: gas exchange, metabolic rate and the mode of energy production. In: *The Vertebrate Gas Transport Cascade: Adaptations to Environment and Mode of Life* [Bicudo, J.E.P.W. (ed.)]. CRC Press Inc, Boca Raton, FL, U.S.A., pp. 330-357.
- Pörtner, H.-O. and R. Knust, 2007:** Climate change affects marine fishes through the oxygen limitation of thermal tolerance. *Science*, **315(5808)**, 95-97.
- Pörtner, H.-O. and A.P. Farrell, 2008:** Ecology: Physiology and climate change. *Science*, **322(5902)**, 690-692.
- Pörtner, H.-O. and M.A. Peck, 2010:** Climate change effects on fishes and fisheries: towards a cause-and-effect understanding. *Journal of Fish Biology*, **77(8)**, 1745-1779.
- Pörtner, H.-O., A. Reipschläger and N. Heisler, 1998:** Acid-base regulation, metabolism and energetics in *Sipunculus nudus* as a function of ambient carbon dioxide level. *Journal of Experimental Biology*, **201(1)**, 43-55.
- Pörtner, H.-O., C. Bock and A. Reipschläger, 2000:** Modulation of the cost of pH_i regulation during metabolic depression: a ³¹P-NMR study in invertebrate (*Sipunculus nudus*) isolated muscle. *Journal of Experimental Biology*, **203(16)**, 2417-2428.
- Pörtner, H.-O., M. Langenbuch and B. Michaelidis, 2005:** Synergistic effects of temperature extremes, hypoxia, and increases in CO₂ on marine animals: From Earth history to global change. *Journal of Geophysical Research*, **110(C9)**, C09S10.
- Pörtner, H.-O., L.S. Peck and T. Hirse, 2006:** Hyperoxia alleviates thermal stress in the Antarctic bivalve, *Laternula elliptica*: evidence for oxygen limited thermal tolerance. *Polar Biology*, **29(8)**, 688-693.
- Pörtner, H.-O., L.S. Peck and G.N. Somero, 2012:** Mechanisms defining thermal limits and adaptation in marine ectotherms: an integrative view. In: *Antarctic Ecosystems: An Extreme Environment in a Changing World* [Rogers, A., N.M. Johnston, E.J. Murphy and A. Clarke (eds.)]. Wiley-Blackwell, Chichester, UK, pp. 360-396.

- Pörtner, H.-O., P.M. Schulte, C.M. Wood and F. Schiemer, 2010:** Niche dimensions in fishes: an integrative view. *Physiological and Biochemical Zoology*, **83(5)**, 808-826.
- Pörtner, H.-O., M. Gutowska, A. Ishimatsu, M. Lucassen, F. Melzner and B. Seibel, 2011:** 8- Effects of ocean acidification on nektonic organisms. In: *Ocean Acidification* [Gattuso, J.-P. and L. Hansson (eds.)]. Oxford University Press, Oxford, UK, pp. 154-175.
- Pörtner, H.-O., C. Bock, R. Knust, G. Lannig, M. Lucassen, F.C. Mark and F.J. Sartoris, 2008:** Cod and climate in a latitudinal cline: physiological analyses of climate effects in marine fishes. *Climate Research*, **37(2-3)**, 253-270.
- Porzio, L., M.C. Buia and J.M. Hall-Spencer, 2011:** Effects of ocean acidification on macroalgal communities. *Journal of Experimental Marine Biology and Ecology*, **400(1-2)**, 278-287.
- Prince, E.D., J. Luo, C.P. Goodyear, J.P. Hoolihan, D. Snodgrass, E.S. Orbesen, J.E. Serafy, M. Ortiz and M.J. Schirripa, 2010:** Ocean scale hypoxia-based habitat compression of Atlantic istiophorid billfishes. *Fisheries Oceanography*, **19(6)**, 448-462.
- Purcell, J.E. and M.B. Decker, 2005:** Effects of climate on relative predation by scyphomedusae and ctenophores on copepods in Chesapeake Bay during 1987-2000. *Limnology and Oceanography*, **50(1)**, 376-387.
- Quartino M.L., D. Deregibus, G.L. Campana, G.E.J. Latorre, F.R. Momo, 2013:** Evidence of macroalgal colonization on newly ice-free areas following glacial retreat in Potter Cove (South Shetland Islands), Antarctica. *PLoS ONE* **8(3)**, e58223. doi:10.1371/journal.pone.0058223.
- Queste, B.Y., L. Fernand, T.D. Jickells and K.J. Heywood, 2013:** Spatial extent and historical context of North Sea oxygen depletion in August 2010. *Biogeochemistry*, **113(1-3)**, 53-68.
- Raab, K., M. Llope, L.A.J. Nagelkerke, A.D. Rijnsdorp, L.R. Teal, P. Licandro, P. Ruardij and M. Dickey-Collas, 2013:** Influence of temperature and food availability on juvenile European anchovy *Engraulis encrasicolus* at its northern boundary. *Marine Ecology Progress Series*, **488**, 233-245.
- Ragazzola, F., L.C. Foster, A. Form, P.S.L. Anderson, T.H. Hansteen and J. Fietzke, 2012:** Ocean acidification weakens the structural integrity of coralline algae. *Global Change Biology*, **18(9)**, 2804-2812.
- Rando, O.J. and K.J. Verstrepen, 2007:** Timescales of genetic and epigenetic inheritance. *Cell*, **128(4)**, 655-668.
- Ratkowsky, D.A., R.K. Lowry, T.A. McMeekin, A.N. Stokes and R.E. Chandler, 1983:** Model for bacterial culture growth rate throughout the entire biokinetic temperature range. *Journal of Bacteriology*, **154(3)**, 1222-1226.
- Rau, G.H., 2011:** CO₂ mitigation via capture and chemical conversion in seawater. *Environmental Science & Technology*, **45(3)**, 1088-1092.
- Rau, G.H., E.L. McLeod and O. Hoegh-Guldberg, 2012:** The need for new ocean conservation strategies in a high-carbon dioxide world. *Nature Climate Change*, **2(10)**, 720-724.
- Raun, A.L. and J. Borum, 2013:** Combined impact of water column oxygen and temperature on internal oxygen status and growth of *Zostera marina* seedlings and adult shoots. *Journal of Experimental Marine Biology and Ecology*, **441**, 16-22.
- Raven, J.A. and C.M. Scrimgeour, 1997:** The influence of anoxia on plants of saline habitats with special reference to the sulphur cycle. *Annals of Botany*, **79**, 79-86.
- Raven, J.A., M. Giordano, J. Beardall and S.C. Maberly, 2012:** Algal evolution in relation to atmospheric CO₂: carboxylases, carbon-concentrating mechanisms and carbon oxidation cycles. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **367(1588)**, 493-507.
- Reid, P.C., M.F. Borges and E. Svendsen, 2001:** A regime shift in the North Sea circa 1988 linked to changes in the North Sea horse mackerel fishery. *Fisheries Research*, **50(1-2)**, 163-171.
- Reipschläger, A. and H.-O. Pörtner, 1996:** Metabolic depression during environmental stress: the role of extracellular versus intracellular pH in *Sipunculus nudus*. *Journal of Experimental Biology*, **199(8)**, 1801-1807.
- Reipschläger, A., G.E. Nilsson and H.-O. Pörtner, 1997:** A role for adenosine in metabolic depression in the marine invertebrate *Sipunculus nudus*. *American Journal of Physiology: Regulatory, Integrative and Comparative Physiology*, **272(1)**, R350-356.
- Reusch, T.B.H. and T.E. Wood, 2007:** Molecular ecology of global change. *Molecular Ecology*, **16(19)**, 3973-3992.
- Reusch, T.B.H. and P.W. Boyd, 2013:** Experimental evolution meets marine phytoplankton. *Evolution*, **67(7)**, 1849-1859.
- Reynaud, S., N. Leclercq, S. Romaine-Lioud, C. Ferrier-Pagès, J. Jaubert and J.-P. Gattuso, 2003:** Interacting effects of CO₂ partial pressure and temperature on photosynthesis and calcification in a scleractinian coral. *Global Change Biology*, **9(11)**, 1660-1668.

- Richards, E.J.**, 2006: Inherited epigenetic variation - revisiting soft inheritance. *Nature Reviews Genetics*, **7(5)**, 395-401.
- Richards, J.G.**, A.P. Farrell and C.J. Brauner (eds.), 2009: *Hypoxia*. Elsevier Academic Press, Amsterdam, 525 pp.
- Richardson, A.J.** and D.S. Schoeman, 2004: Climate impact on plankton ecosystems in the Northeast Atlantic. *Science*, **305(5690)**, 1609-1612.
- Richardson, A.J.** and M.J. Gibbons, 2008: Are jellyfish increasing in response to ocean acidification? *Limnology and Oceanography*, **53(5)**, 2040-2045.
- Richardson, A.J.**, A. Bakun, G.C. Hays and M.J. Gibbons, 2009: The jellyfish joyride: causes, consequences and management responses to a more gelatinous future. *Trends in Ecology and Evolution*, **24(6)**, 312-322.
- Richardson, A.J.**, A.W. Walne, A.W.G. John, T.D. Jonas, J.A. Lindley, D.W. Sims, D. Stevens and M. Witt, 2006: Using continuous plankton recorder data. *Progress in Oceanography*, **68(1)**, 27-74.
- Richier, S.**, M.E. Kerros, C. de Vargas, L. Haramaty, P.G. Falkowski and J.-P. Gattuso, 2009: Light-dependent transcriptional regulation of genes of biogeochemical interest in the diploid and haploid life cycle stages of *Emiliana huxleyi*. *Applied and Environmental Microbiology*, **75(10)**, 3366-3369.
- Ridgwell, A.** and D.N. Schmidt, 2010: Past constraints on the vulnerability of marine calcifiers to massive carbon dioxide release. *Nature Geoscience*, **3(3)**, 196-200.
- Riebesell, U.**, 2004: Effects of CO₂ enrichment on marine phytoplankton. *Journal of Oceanography*, **60(4)**, 719-729.
- Riebesell, U.**, R.G.J. Bellerby, H.P. Grossart and F. Thingstad, 2008: Mesocosm CO₂ perturbation studies: from organism to community level. *Biogeosciences*, **5(4)**, 1157-1164.
- Riebesell, U.**, J.-P. Gattuso, T.F. Thingstad and J. Middelburg (eds.), 2013: *Arctic ocean acidification: pelagic ecosystem and biogeochemical responses during a mesocosm study*. Biogeosciences Special issue.
- Riebesell, U.**, I. Zondervan, B. Rost, P.D. Tortell, R.E. Zeebe and F.M.M. Morel, 2000: Reduced calcification of marine plankton in response to increased atmospheric CO₂. *Nature*, **407(6802)**, 364-367.
- Riebesell, U.**, K.G. Schulz, R.G.J. Bellerby, M. Botros, P. Fritsche, M. Meyerhöfer, C. Neill, G. Nondal, A. Oschlies, J. Wohlers and E. Zöllner, 2007: Enhanced biological carbon consumption in a high CO₂ ocean. *Nature*, **450(7169)**, 545-548.
- Ries, J.B.**, A.L. Cohen and D.C. McCorkle, 2009: Marine calcifiers exhibit mixed responses to CO₂-induced ocean acidification. *Geology*, **37(12)**, 1131-1134.
- Robinson, C.**, D.K. Steinberg, T.R. Anderson, J. Arístegui, C.A. Carlson, J.R. Frost, J.F. Ghiglione, S. Hernández-Léon, G.A. Jackson, R. Koppelman, B. Queguiner, O. Ragueneau, F. Rassoulzadegan, B.H. Robison, C. Tamburini, T. Tanaka, K.F. Wishner and J. Zhang, 2010: Mesopelagic zone ecology and biogeochemistry - a synthesis. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **57(16)**, 1504-1518.
- Robinson, R.A.**, J.A. Learmonth, A.M. Hutson, C.D. MacLeod, T.H. Sparks, D.I. Leech, G.J. Pierce, M.M. Rehfish and H.Q.P. Crick, 2005: *Climate Change and Migratory Species, Research Report 414*. BTO, Thetford, UK, 308 pp.
- Rode, K.D.**, E. Peacock, M. Taylor, I. Stirling, E.W. Born, K.L. Laidre and Ø. Wiig, 2012: A tale of two polar bear populations: ice habitat, harvest, and body condition. *Population Ecology*, **54(1)**, 3-18.
- Rodolfo-Metalpa, R.**, F. Houlbrèque, É. Tambutté, F. Boisson, C. Baggini, F.P. Patti, R. Jeffree, M. Fine, A. Foggo, J.-P. Gattuso and J.M. Hall-Spencer, 2011: Coral and mollusc resistance to ocean acidification adversely affected by warming. *Nature Climate Change*, **1(6)**, 308-312.
- Rokitta, S.D.** and B. Rost, 2012: Effects of CO₂ and their modulation by light in the life-cycle stages of the coccolithophore *Emiliana huxleyi*. *Limnology and Oceanography*, **57(2)**, 607-618.
- Romanuk, T.N.**, Y. Zhou, U. Brose, E.L. Berlow, R.J. Williams and N.D. Martinez, 2009: Predicting invasion success in complex ecological networks. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **364**, 1743-1754.
- Rose, G.** and R.L. O'Driscoll, 2002: Capelin are good for cod: can the northern stock rebuild without them? *ICES Journal of Marine Science*, **59(5)**, 1018-1026.
- Rose, J.M.**, Y. Feng, C.J. Gobler, R. Gutierrez, C.E. Hare, K. Leblanc and D.A. Hutchins, 2009: Effects of increased pCO₂ and temperature on the North Atlantic spring bloom. II. Microzooplankton abundance and grazing. *Marine Ecology Progress Series*, **388**, 27-40.
- Rose, K.A.**, J.I. Allen, Y. Artioli, M. Barange, J. Blackford, F. Carlotti, R. Cropp, U. Daewel, K. Edwards, K. Flynn, S.L. Hill, R. HilleRisLambers, G. Huse, S. Mackinson, B. Megrey, A. Moll, R. Rivkin, B. Salihoglu, C. Schrum, L. Shannon, Y.-J. Shin, S.L. Smith, C. Smith, C. Solidoro, M. St. John and M. Zhou, 2010: End-to-end models

- for the analysis of marine ecosystems: challenges, issues, and next steps. *Marine and Coastal Fisheries*, **2(1)**, 115-130.
- Rosegrant**, M.W. and S.A. Cline, 2003: Global food security: challenges and policies. *Science*, **302(5652)**, 1917-1919.
- Rossoll**, D., R. Bermúdez, H. Hauss, K.G. Schulz, U. Riebesell, U. Sommer and M. Winder, 2012: Ocean acidification-induced food quality deterioration constrains trophic transfer. *PLoS ONE*, **7(4)**, e34737.
- Rost**, B., I. Zondervan and D. Wolf-Gladrow, 2008: Sensitivity of phytoplankton to future changes in ocean carbonate chemistry: current knowledge, contradictions and research directions. *Marine Ecology Progress Series*, **373**, 227-237.
- Rost**, B., U. Riebesell, S. Burkhardt and D. Sültemeyer, 2003: Carbon acquisition of bloom-forming marine phytoplankton. *Limnology and Oceanography*, **48(1)**, 55-67.
- Russell**, B.D., C.A. Passarelli and S.D. Connell, 2011: Forecasted CO₂ modifies the influence of light in shaping subtidal habitat. *Journal of Phycology*, **47(4)**, 744-752.
- Russell**, L.M., P.J. Rasch, G.M. Mace, R.B. Jackson, J. Shepherd, P. Liss, M. Leinen, D. Schimel, N.E. Vaughan, A.C. Janetos, P.W. Boyd, R.J. Norby, K. Caldeira, J. Merikanto, P. Artaxo, J. Melillo and M.G. Morgan, 2012: Ecosystem impacts of geoengineering: a review for developing a science plan. *Ambio*, **41(4)**, 350-369.
- Rykaczewski**, R.R. and J.P. Dunne, 2011: A measured look at ocean chlorophyll trends. *Nature*, **472(7342)**, E5-E6.
- Saba**, G.K., O. Schofield, J.J. Torres, E.H. Ombres and D.K. Steinberg, 2012: Increased feeding and nutrient excretion of adult Antarctic krill, *Euphausia superba*, exposed to enhanced carbon dioxide (CO₂). *PLoS ONE*, **7(12)**, e52224.
- Saba**, V.S., C.A. Stock, J.R. Spotila, F.V. Paladino and P.S. Tomillo, 2012: Projected response of an endangered marine turtle population to climate change. *Nature Climate Change*, **2(11)**, 814-820.
- Saba**, V.S., M.A.M. Friedrichs, D. Antoine, R.A. Armstrong, I. Asanuma, M.J. Behrenfeld, A.M. Ciotti, M. Dowell, N. Hoepffner, K.J.W. Hyde, J. Ishizaka, T. Kameda, J. Marra, F. Mélin, A. Morel, J. O'Reilly, M. Scardi, W.O. Smith Jr., T.J. Smyth, S. Tang, J. Uitz, K. Waters and T.K. Westberry, 2011: An evaluation of ocean color model estimates of marine primary productivity in coastal and pelagic regions across the globe. *Biogeosciences*, **8(2)**, 489-503.
- Sabatés**, A., P. Martín, J. Lloret and V. Raya, 2006: Sea warming and fish distribution: the case of the small pelagic fish, *Sardinella aurita*, in the western Mediterranean. *Global Change Biology*, **12(11)**, 2209-2219.
- Saito**, M.A., T.J. Goepfert and J.T. Ritt, 2008: Some thoughts on the concept of colimitation: three definitions and the importance of bioavailability. *Limnology and Oceanography*, **53(1)**, 276-290.
- Saito**, M.A., J.W. Moffett, S.W. Chisholm and J.B. Waterbury, 2002: Cobalt limitation and uptake in *Prochlorococcus*. *Limnology and Oceanography*, **47(6)**, 1629-1636.
- Sala**, E. and N. Knowlton, 2006: Global marine biodiversity trends. *Annual Review of Environment and Resources*, **31(1)**, 93-122.
- Sallée**, J.B., E. Shuckburgh, N. Bruneau, A.J.S. Meijers, T.J. Bracegirdle and Z. Wang, 2013: Assessment of Southern Ocean mixed-layer depths in CMIP5 models: Historical bias and forcing response. *Journal of Geophysical Research: Oceans*, **118(4)**, 1845-1862.
- Salvadeo**, C., D. Lluch-Belda, S. Lluch-Cota and M. Mercuri, 2011: Review of long term macro-fauna movement by multi-decadal warming trends in the Northeastern Pacific. In: *Climate Change: Geophysical Foundations and Ecological Effects* [Blanco, J. and H. Kheradmand (eds.)]. InTech, Rijeka, Croatia, pp. 217-230.
- Salvadeo**, C., S. Lluch-Cota, M. Maravilla-Chávez, S. Álvarez-Castañeda, M. Mercuri and A. Ortega-Rubio, 2013: Impact of climate change on sustainable management of gray whale (*Eschrichtius robustus*) populations: Whale-watching and conservation. *Archives of Biological Sciences*, **65(3)**, 997-1005.
- Salvadeo**, C.J., D. Lluch-Belda, A. Gómez-Gallardo, J. Urbán-Ramírez and C.D. MacLeod, 2010: Climate change and a poleward shift in the distribution of the Pacific white-sided dolphin in the northeastern Pacific. *Endangered Species Research*, **11(1)**, 13-19.
- Santidrián Tomillo**, P., V.S. Saba, G.S. Blanco, C.A. Stock, F.V. Paladino and J.R. Spotila, 2012: Climate driven egg and hatchling mortality threatens survival of eastern Pacific leatherback turtles. *PLoS ONE*, **7(5)**, e37602.
- Sarker**, M.Y., I. Bartsch, M. Olischläger, L. Gutow and C. Wiencke, 2013: Combined effects of CO₂, temperature, irradiance and time on the physiological performance of *Chondrus crispus* (Rhodophyta). *Botanica Marina*, **56(1)**, 63-74.

- Sarmiento, H.**, J.M. Montoya, E. Vázquez-Domínguez, D. Vaqué and J.M. Gasol, 2010: Warming effects on marine microbial food web processes: how far can we go when it comes to predictions? *Philosophical Transactions of the Royal Society B: Biological Sciences*, **365(1549)**, 2137-2149.
- Sarmiento, J.L.** and N. Gruber, 2006: *Ocean Biogeochemical Dynamics*. Princeton University Press, Princeton, NJ, USA, 526 pp.
- Sarmiento, J.L.**, T.M.C. Hughes, R.J. Stouffer and S. Manabe, 1998: Simulated response of the ocean carbon cycle to anthropogenic climate warming. *Nature*, **393(6682)**, 245-249.
- Sarmiento, J.L.**, R.D. Slater, J. Dunne, A. Gnanadesikan and M.R. Hiscock, 2010: Efficiency of small scale carbon mitigation by patch iron fertilization. *Biogeosciences*, **7(11)**, 3593-3624.
- Scheibner, C.** and R.P. Speijer, 2008: Late Paleocene-early Eocene Tethyan carbonate platform evolution - A response to long- and short-term paleoclimatic change. *Earth-Science Reviews*, **90(3-4)**, 71-102.
- Schlüter, M.**, A. Merico, K.H. Wiltshire, W. Greve and H. von Storch, 2008: A statistical analysis of climate variability and ecosystem response in the German Bight. *Ocean Dynamics*, **58(3-4)**, 169-186.
- Schlüter, M.H.**, A. Merico, M. Reginatto, M. Boersma, K.H. Wiltshire and W. Greve, 2010: Phenological shifts of three interacting zooplankton groups in relation to climate change. *Global Change Biology*, **16(11)**, 3144-3153.
- Schöne, B.R.**, W. Oschmann, J. Rössler, A.D.F. Castro, S.D. Houk, I. Kröncke, W. Dreyer, R. Janssen, H. Rumohr and E. Dunca, 2003: North Atlantic Oscillation dynamics recorded in shells of a long-lived bivalve mollusk. *Geology*, **31(12)**, 1037-1040.
- Sciandra, A.**, J. Harlay, D. Lefèvre, R. Lemée, P. Rimmelin, M. Denis and J.-P. Gattuso, 2003: Response of coccolithophorid *Emiliana huxleyi* to elevated partial pressure of CO₂ under nitrogen limitation. *Marine Ecology Progress Series*, **261**, 111-122.
- Seibel, B.A.**, 2011: Critical oxygen levels and metabolic suppression in oceanic oxygen minimum zones. *Journal of Experimental Biology*, **214(2)**, 326-336.
- Seki, O.**, G.L. Foster, D.N. Schmidt, A. Mackensen, K. Kawamura and R.D. Pancost, 2010: Alkenone and boron-based Pliocene pCO₂ records. *Earth and Planetary Science Letters*, **292(1-2)**, 201-211.
- Sen Gupta, A.** and B. McNeil, 2012: Variability and Change in the Ocean. In: *The Future of the World's Climate* [Henderson-Sellers, A. and K. McGuffie (eds.)]. Elsevier, Amsterdam, pp. 141-165.
- Shepherd, J.**, K. Caldeira, P. Cox, J. Haigh, D. Keith, B. Launder, G. Mace, G. MacKerron, J. Pyle, S. Rayner, C. Redgwell, A. Watson, R. Garthwaite, R. Heap, A. Parker and J. Wilsdon, 2009: *Geoengineering the Climate*. The Royal Society, London, UK, 98 pp.
- Sheppard-Brennand, H.**, N. Soars, S.A. Dworjanyn, A.R. Davis and M. Byrne, 2010: Impact of ocean warming and ocean acidification on larval development and calcification in the sea urchin *Tripneustes gratilla*. *PLoS ONE*, **5(6)**, e11372.
- Sherman, K.** and G. Hempel, 2009: *The UNEP Large Marine Ecosystem Report: a perspective on changing conditions in LMEs of the world's Regional Seas*. UNEP Regional Seas Reports and Studies, 872 pp.
- Sherman, K.**, M. Sissenwine, V. Christensen, A. Duda, G. Hempel, C. Ibe, S. Levin, D. Lluch-Belda, G. Matishov, J. McGlade, M. O'Toole, S. Seitzinger, R. Serra, H.R. Skjoldal, Q. Tang, J. Thulin, V. Vandeweerde and K. Zwanenburg, 2005: A global movement toward an ecosystem approach to management of marine resources. *Marine Ecology Progress Series*, **300**, 275-279.
- Short, F.T.** and H.A. Neckles, 1999: The effects of global climate change on seagrasses. *Aquatic Botany*, **63(3-4)**, 169-196.
- Signorini, S.R.** and C.R. McClain, 2012: Subtropical gyre variability as seen from satellites. *Remote Sensing Letters*, **3(6)**, 471-479.
- Signorini, S.R.**, R.G. Murtugudde, C.R. McClain, J.R. Christian, J. Picaut and A.J. Busalacchi, 1999: Biological and physical signatures in the tropical and subtropical Atlantic. *Journal of Geophysical Research*, **104(8)**, 18376-18382.
- Simmonds, M.P.** and S.J. Isaac, 2007: The impacts of climate change on marine mammals: early signs of significant problems. *Oryx*, **41(1)**, 19-26.
- Simpson, S.D.**, S. Jennings, M.P. Johnson, J.L. Blanchard, P.-J. Schön, D.W. Sims and M.J. Genner, 2011: Continental shelf-wide response of a fish assemblage to rapid warming of the sea. *Current Biology*, **21(18)**, 1565-1570.
- Sissener, E.H.** and T. Bjørndal, 2005: Climate change and the migratory pattern for Norwegian spring-spawning herring - implications for management. *Marine Policy*, **29(4)**, 299-309.

- Sluijs, A. and H. Brinkhuis, 2009: A dynamic climate and ecosystem state during the Paleocene-Eocene Thermal Maximum: inferences from dinoflagellate cyst assemblages on the New Jersey Shelf. *Biogeosciences*, **6(8)**, 1755-1781.
- Smetacek, V. and S. Nicol, 2005: Polar ocean ecosystems in a changing world. *Nature*, **437(7057)**, 362-368.
- Smith, K.L., Jr., H.A. Ruhl, B.J. Bett, D.S. Billett, R.S. Lampitt and R.S. Kaufmann, 2009: Climate, carbon cycling, and deep-ocean ecosystems. *Proceedings of the National Academy of Sciences of the United States of America*, **106(46)**, 19211-19218.
- Smith, S.V., 1981: Marine macrophytes as a global carbon sink. *Science*, **211(4484)**, 838-840.
- Snyder, M.A., L.C. Sloan, N.S. Diffenbaugh and J.L. Bell, 2003: Future climate change and upwelling in the California Current. *Geophysical Research Letters*, **30(15)**, 1823.
- Sohm, J.A., E.A. Webb and D.G. Capone, 2011: Emerging patterns of marine nitrogen fixation. *Nature Reviews Microbiology*, **9(7)**, 499-508.
- Sorokin, C. and R.W. Krauss, 1962: Effects of temperature & illuminance on *Chlorella* growth uncoupled from cell division. *Plant Physiology*, **37(1)**, 37-42.
- Soto, C.G., 2001: The potential impacts of global climate change on marine protected areas. *Reviews in Fish Biology and Fisheries*, **11(3)**, 181-195.
- Springer, A.M., J.F. Piatt, V.P. Shuntov, G.B. Van Vliet, V.L. Vladimirov, A.E. Kuzin and A.S. Perlov, 1999: Marine birds and mammals of the Pacific Subarctic Gyres. *Progress in Oceanography*, **43(2-4)**, 443-487.
- Stassen, P., E. Thomas and R.P. Speijer, 2012: The progression of environmental changes during the onset of the Paleocene-Eocene Thermal Maximum (New Jersey Coastal Plain). *Austrian Journal of Earth Sciences*, **105(1)**, 169-178.
- Steinacher, M., F. Joos, T.L. Frölicher, G.K. Plattner and S.C. Doney, 2009: Imminent ocean acidification in the Arctic projected with the NCAR global coupled carbon cycle-climate model. *Biogeosciences*, **6(4)**, 515-533.
- Steinacher, M., F. Joos, T.L. Frölicher, L. Bopp, P. Cadule, V. Cocco, S.C. Doney, M. Gehlen, K. Lindsay, J.K. Moore, B. Schneider and J. Segschneider, 2010: Projected 21st century decrease in marine productivity: a multi-model analysis. *Biogeosciences*, **7(3)**, 979-1005.
- Stenevik, E.K. and S. Sundby, 2007: Impacts of climate change on commercial fish stocks in Norwegian waters. *Marine Policy*, **31(1)**, 19-31.
- Stige, L.C., G. Ottersen, P. Dalpadado, K.-S. Chan, D. Hjermann, D.L. Lajus, N.A. Yaragina and N.C. Stenseth, 2010: Direct and indirect climate forcing in a multi-species marine system. *Proceedings of the Royal Society B: Biological Sciences*, **277(1699)**, 3411-3420.
- Stock, C. and J. Dunne, 2010: Controls on the ratio of mesozooplankton production to primary production in marine ecosystems. *Deep-Sea Research Part I: Oceanographic Research Papers*, **57(1)**, 95-112.
- Stock, C.A., M.A. Alexander, N.A. Bond, K.M. Brander, W.W.L. Cheung, E.N. Curchitser, T.L. Delworth, J.P. Dunne, S.M. Griffies, M.A. Haltuch, J.A. Hare, A.B. Hollowed, P. Lehodey, S.A. Levin, J.S. Link, K.A. Rose, R.R. Rykaczewski, J.L. Sarmiento, R.J. Stouffer, F.B. Schwing, G.A. Vecchi and F.E. Werner, 2011: On the use of IPCC-class models to assess the impact of climate on living marine resources. *Progress in Oceanography*, **88(1-4)**, 1-27.
- Stolper, D.A., N.P. Revsbech and D.E. Canfield, 2010: Aerobic growth at nanomolar oxygen concentrations. *Proceedings of the National Academy of Sciences of the United States of America*, **107(44)**, 18755-18760.
- Stramma, L., G.C. Johnson, J. Sprintall and V. Mohrholz, 2008: Expanding oxygen-minimum zones in the tropical oceans. *Science*, **320(5876)**, 655-658.
- Stramma, L., S. Schmidtko, L.A. Levin and G.C. Johnson, 2010: Ocean oxygen minima expansions and their biological impacts. *Deep-Sea Research Part I: Oceanographic Research Papers*, **57(4)**, 587-595.
- Stramma, L., E.D. Prince, S. Schmidtko, J. Luo, J.P. Hoolihan, M. Visbeck, D.W.R. Wallace, P. Brandt and A. Kortzinger, 2012: Expansion of oxygen minimum zones may reduce available habitat for tropical pelagic fishes. *Nature Climate Change*, **2(1)**, 33-37.
- Strand, S.E. and G. Benford, 2009: Ocean sequestration of crop residue carbon: recycling fossil fuel carbon back to deep sediments. *Environmental Science & Technology*, **43(4)**, 1000-1007.
- Strong, A.E., G. Liu, W. Skirving and C.M. Eakin, 2011: NOAA's Coral Reef Watch program from satellite observations. *Annals of GIS*, **17(2)**, 83-92.
- Stumpp, M., K. Trubenbach, D. Brennecke, M.Y. Hu and F. Melzner, 2012: Resource allocation and extracellular acid-base status in the sea urchin *Strongylocentrotus droebachiensis* in response to CO₂ induced seawater acidification. *Aquatic Toxicology*, **110-111**, 194-207.

- Sumaila**, U.R. and W.W.L. Cheung, 2010: *Development and Climate Change: Cost of Adapting Fisheries to Climate Change*. Discussion Paper Number 5, International Bank for Reconstruction and Development/ World Bank, Washington, D.C., USA, 37 pp.
- Sumaila**, U.R., W.W.L. Cheung, V.W.Y. Lam, D. Pauly and S. Herrick, 2011: Climate change impacts on the biophysics and economics of world fisheries. *Nature Climate Change*, **1(9)**, 449-456.
- Sun**, J., D.A. Hutchins, Y. Feng, E.L. Seubert, D.A. Caron and F.-X. Fu, 2011: Effects of changing $p\text{CO}_2$ and phosphate availability on domoic acid production and physiology of the marine harmful bloom diatom *Pseudo-nitzschia* multiseriis. *Limnology and Oceanography*, **56(3)**, 829-840.
- Sunday**, J.M., A.E. Bates and N.K. Dulvy, 2012: Thermal tolerance and the global redistribution of animals. *Nature Climate Change*, **2(9)**, 686-690.
- Sunday**, J.M., R.N. Crim, C.D.G. Harley and M.W. Hart, 2011: Quantifying rates of evolutionary adaptation in response to ocean acidification. *PLoS ONE*, **6(8)**, e22881.
- Sverdrup**, H.U., 1953: On conditions for the vernal blooming of phytoplankton. *ICES Journal of Marine Science*, **18(3)**, 287-295.
- Sydeinan**, W.J. and S.J. Bograd, 2009: Marine ecosystems, climate and phenology: introduction. *Marine Ecology Progress Series*, **393**, 185-188.
- Takai**, K., A. Inoue and K. Horikoshi, 1999: *Thermaerobacter marianensis* gen. nov., sp. nov., an aerobic extremely thermophilic marine bacterium from the 11000 m deep Mariana Trench. *International Journal of Systematic Bacteriology*, **49(2)**, 619-628.
- Takai**, K., K. Nakamura, T. Toki, U. Tsunogai, M. Miyazaki, J. Miyazaki, H. Hirayama, S. Nakagawa, T. Nunoura and K. Horikoshi, 2008: Cell proliferation at 122°C and isotopically heavy CH_4 production by a hyperthermophilic methanogen under high-pressure cultivation. *Proceedings of the National Academy of Sciences of the United States of America*, **105(31)**, 10949-10954.
- Takasuka**, A. and I. Aoki, 2006: Environmental determinants of growth rates for larval Japanese anchovy *Engraulis japonicus* in different waters. *Fisheries Oceanography*, **15(2)**, 139-149.
- Takasuka**, A., Y. Oozeki and I. Aoki, 2007: Optimal growth temperature hypothesis: why do anchovy flourish and sardine collapse or vice versa under the same ocean regime? *Canadian Journal of Fisheries and Aquatic Sciences*, **64(5)**, 768-776.
- Takasuka**, A., Y. Oozeki and H. Kubota, 2008: Multi-species regime shifts reflected in spawning temperature optima of small pelagic fish in the western North Pacific. *Marine Ecology Progress Series*, **360**, 211-217.
- Tatters**, A.O., F.-X. Fu and D.A. Hutchins, 2012: High CO_2 and silicate limitation synergistically increase the toxicity of *Pseudo-nitzschia fraudulenta*. *PLoS ONE*, **7(2)**, e32116.
- Tatters**, A.O., A. Schnetzer, F. Fu, A.Y.A. Lie, D.A. Caron and D.A. Hutchins, 2013: Short- versus long-term responses to changing CO_2 in a coastal dinoflagellate bloom: implications for interspecific competitive interactions and community structure. *Evolution*, **67(7)**, 1879-1891.
- Taucher**, J. and A. Oschlies, 2011: Can we predict the direction of marine primary production change under global warming? *Geophysical Research Letters*, **38(2)**, L02603.
- Taylor**, A.R., A. Chrachri, G. Wheeler, H. Goddard and C. Brownlee, 2011: A voltage-gated H^+ channel underlying pH homeostasis in calcifying Coccolithophores. *PLoS Biology*, **9(6)**, e1001085.
- Taylor**, C.C., 1958: Cod growth and temperature. *ICES Journal of Marine Science*, **23**, 366-370.
- Teneva**, L., M. Karnauskas, C.A. Logan, L. Bianucci, J.C. Currie and J.A. Kleypas, 2012: Predicting coral bleaching hotspots: the role of regional variability in thermal stress and potential adaptation rates. *Coral Reefs*, **31(1)**, 1-12.
- Tenreiro**, S., M.F. Nobre, F.A. Rainey, C. Miguel and M.S. Da Costa, 1997: *Thermonema rossianum* sp. nov., a new thermophilic and slightly halophilic species from saline hot springs in Naples, Italy. *International Journal of Systematic Bacteriology*, **47(1)**, 122-126.
- Thackeray**, S.J., T.H. Sparks, M. Frederiksen, S. Burthe, P.J. Bacon, J.R. Bell, M.S. Botham, T.M. Brereton, P.W. Bright, L. Carvalho, T. Clutton-Brock, A. Dawson, M. Edwards, J.M. Elliott, R. Harrington, D. Johns, I.D. Jones, J.T. Jones, D.I. Leech, D.B. Roy, W.A. Scott, M. Smith, R.J. Smithers, I.J. Winfield and S. Wanless, 2010: Trophic level asynchrony in rates of phenological change for marine, freshwater and terrestrial environments. *Global Change Biology*, **16(12)**, 3304-3313.
- Thomas**, E., 2003: Extinction and food at the seafloor: a high-resolution benthic foraminiferal record across the Initial Eocene Thermal Maximum, Southern Ocean Site 690. In: *Causes and Consequences of Globally Warm*

- Climates in the Early Paleogene: Geological Society of America Special Paper 369* [Wing, S.L., P.D. Gingerich, B. Schmitz and E. Thomas (eds.)]. Geological Society of America, Boulder, CO, USA, pp. 319-332.
- Thomas, E.**, 2007: Cenozoic mass extinctions in the deep sea: what perturbs the largest habitat on earth? In: *Large Scale Ecosystem Perturbation: Causes and Consequences: Geological Society of America* [Monechi, S., R. Coccioni and M.R. Rampino (eds.)]. Geological Society of America, Boulder, CO, USA, pp. 1-23.
- Thomas, M.K.**, C.T. Kremer, C.A. Klausmeier and E. Litchman, 2012: A global pattern of thermal adaptation in marine phytoplankton. *Science*, **338(6110)**, 1085-1088.
- Thomsen, J.** and F. Melzner, 2010: Moderate seawater acidification does not elicit long-term metabolic depression in the blue mussel *Mytilus edulis*. *Marine Biology*, **157(12)**, 2667-2676.
- Thresher, R.E.**, T. B., S. Fallon, N.C. Wilson and J. Adkins, 2011: Effects of chronic low carbonate saturation levels on the distribution, growth and skeletal chemistry of deep-sea corals and other seamount megabenthos. *Marine Ecology Progress Series*, **442**, 87-99.
- Tittensor, D.P.**, C. Mora, W. Jetz, H.K. Lotze, D. Ricard, E.V. Berghe and B. Worm, 2010: Global patterns and predictors of marine biodiversity across taxa. *Nature*, **466(7310)**, 1098-1101.
- Tortell, P.D.**, C.D. Payne, Y. Li, S. Trimborn, B. Rost, W.O. Smith, C. Riesselman, R.B. Dunbar, P. Sedwick and G.R. DiTullio, 2008: CO₂ sensitivity of Southern Ocean phytoplankton. *Geophysical Research Letters*, **35(4)**, L04605.
- Trick, C.G.**, B.D. Bill, W.P. Cochlan, M.L. Wells, V.L. Trainer and L.D. Pickell, 2010: Iron enrichment stimulates toxic diatom production in high-nitrate, low-chlorophyll areas. *Proceedings of the National Academy of Sciences of the United States of America*, **107(13)**, 5887-5892.
- Trimborn, S.**, N. Lundholm, S. Thoms, K.U. Richter, B. Krock, P.J. Hansen and B. Rost, 2008: Inorganic carbon acquisition in potentially toxic and non-toxic diatoms: the effect of pH-induced changes in seawater carbonate chemistry. *Physiologia Plantarum*, **133(1)**, 92-105.
- Trotter, J.**, P. Montagna, M. McCulloch, S. Silenzi, S. Reynaud, G. Mortimer, S. Martin, C. Ferrier-Pagès, J.-P. Gattuso and R. Rodolfo-Metalpa, 2011: Quantifying the pH 'vital effect' in the temperate zooxanthellate coral *Cladocora caespitosa*: validation of the boron seawater pH proxy. *Earth and Planetary Science Letters*, **303(3-4)**, 163-173.
- Tseng, Y.-C.**, M.Y. Hu, M. Stumpp, L.-Y. Lin, F. Melzner and P.-P. Hwang, 2013: CO₂-driven seawater acidification differentially affects development and molecular plasticity along life history of fish (*Oryzias latipes*). *Comparative Biochemistry and Physiology A: Molecular and Integrative Physiology*, **165(2)**, 119-130.
- Ulloa, O.**, D.E. Canfield, E.F. DeLong, R.M. Letelier and F.J. Stewart, 2012: Microbial oceanography of anoxic oxygen minimum zones. *Proceedings of the National Academy of Sciences of the United States of America*, **109(40)**, 15996-16003.
- UNWTO**, 2008: *Climate Change and Tourism - Responding to Global Challenges*. World Tourism Organization and the United Nations Environment Programme, Madrid, 272 pp.
- Urban, M.C.**, J.J. Tewksbury and K.S. Sheldon, 2012: On a collision course: competition and dispersal differences create no-analogue communities and cause extinctions during climate change. *Proceedings of the Royal Society B: Biological Sciences*, **279**, 2072-2080.
- Utne-Palm, A.C.**, A.G. Salvanes, B. Currie, S. Kaartvedt, G.E. Nilsson, V.A. Braithwaite, J.A. Stecyk, M. Hundt, M. van der Bank, B. Flynn, G.K. Sandvik, T.A. Klevjer, A.K. Sweetman, V. Bruchert, K. Pittman, K.R. Peard, I.G. Lunde, R.A. Strandabo and M.J. Gibbons, 2010: Trophic structure and community stability in an overfished ecosystem. *Science*, **329(5989)**, 333-336.
- Valdimarsson, H.**, O.S. Astthorsson and J. Palsson, 2012: Hydrographic variability in Icelandic waters during recent decades and related changes in distribution of some fish species. *ICES Journal of Marine Science*, **69(5)**, 816-825.
- Van Houtan, K.S.** and O.L. Bass, 2007: Stormy oceans are associated with declines in sea turtle hatching. *Current Biology*, **17(15)**, 590-591.
- Van Houtan, K.S.** and J.M. Halley, 2011: Long-term climate forcing in loggerhead sea turtle nesting. *PLoS ONE*, **6(4)**, e19043.
- Vaquer-Sunyer, R.** and C.M. Duarte, 2008: Thresholds of hypoxia for marine biodiversity. *Proceedings of the National Academy of Sciences of the United States of America*, **105(40)**, 15452-15457.
- Vaquer-Sunyer, R.** and C.M. Duarte, 2011: Temperature effects on oxygen thresholds for hypoxia in marine benthic organisms. *Global Change Biology*, **17(5)**, 1788-1797.

- Vargas, F.H., R.C. Lacy, P.J. Johnson, A. Steinfurth, R.J.M. Crawford, P. Dee Boersma and D.W. Macdonald, 2007: Modelling the effect of El Niño on the persistence of small populations: the Galápagos penguin as a case study. *Biological Conservation*, **137(1)**, 138-148.
- Vásquez-Bedoya, L.F., A.L. Cohen, D.W. Oppo and P. Blanchon, 2012: Corals record persistent multidecadal SST variability in the Atlantic Warm Pool since 1775 AD. *Paleoceanography*, **27**, PA3231.
- Vaughan, N.E. and T.M. Lenton, 2011: A review of climate geoengineering proposals. *Climatic Change*, **109(3-4)**, 745-790.
- Vecchi, G.A. and B.J. Soden, 2007: Increased tropical Atlantic wind shear in model projections of global warming. *Geophysical Research Letters*, **34**, L08702.
- Vélez-Belchí, P., A. Hernández-Guerra, E. Fraile-Nuez and V. Benítez-Barrios, 2010: Changes in temperature and salinity tendencies of the upper subtropical North Atlantic ocean at 24.5°N. *Journal of Physical Oceanography*, **40(11)**, 2546-2555.
- Venn, A.A., E. Tambutté, M. Holcomb, J. Laurent, D. Allemand and S. Tambutté, 2013: Impact of seawater acidification on pH at the tissue-skeleton interface and calcification in reef corals. *Proceedings of the National Academy of Sciences of the United States of America*, **110(5)**, 1634-1639.
- Ventura, S., C. Viti, R. Pastorelli and L. Giovannetti, 2000: Revision of species delineation in the genus *Ectothiorhodospira*. *International Journal of Systematic and Evolutionary Microbiology*, **50(2)**, 583-591.
- Vermeij, G.J. and E.J. Petuch, 1986: Differential extinction in tropical American molluscs: endemism, architecture, and the Panama land bridge. *Malacologia*, **27(1)**, 29-41.
- Veron, J.E.N., O. Hoegh-Guldberg, T.M. Lenton, J.M. Lough, D.O. Obura, P. Pearce-Kelly, C.R. Sheppard, M. Spalding, M.G. Stafford-Smith and A.D. Rogers, 2009: The coral reef crisis: the critical importance of < 350 ppm CO₂. *Marine Pollution Bulletin*, **58(10)**, 1428-1436.
- Veron, J.E.N., 2011: Ocean acidification and coral reefs: an emerging big picture. *Diversity*, **3(2)**, 262-274.
- Vetter, R.D., E.A. Lynn, M. Garza and A.S. Costa, 1994: Depth zonation and metabolic adaptation in Dover sole, *Microstomus pacificus*, and other deep-living flatfishes: factors that affect the sole. *Marine Biology*, **120(1)**, 145-159.
- Vezzoli, A., M. Gussoni, F. Greco, L. Zetta and P. Cerretelli, 2004: Temperature and pH dependence of energy balance by ³¹P- and ¹H-MRS in anaerobic frog muscle. *Biochimica et Biophysica Acta*, **1608(2-3)**, 163-170.
- Vezzulli, L., C. Pruzzo, A. Huq and R.R. Colwell, 2010: Environmental reservoirs of *Vibrio cholerae* and their role in cholera. *Environmental Microbiology Reports*, **2(1)**, 27-33.
- Vitasse, Y., C.C. Bresson, A. Kremer, R. Michalet and S. Delzon, 2010: Quantifying phenological plasticity to temperature in two temperate tree species. *Functional Ecology*, **24(6)**, 1211-1218.
- Vogt, M., M. Steinke, S. Turner, A. Paulino, M. Meyerhofer, U. Riebesell, C. LeQuéré and P. Liss, 2008: Dynamics of dimethylsulphoniopropionate and dimethylsulphide under different CO₂ concentrations during a mesocosm experiment. *Biogeosciences*, **5**, 407-419.
- Votier, S.C., B.J. Hatchwell, A. Beckerman, R.H. McCleery, F.M. Hunter, J. Pellatt, M. Trinder and T.R. Birkhead, 2005: Oil pollution and climate have wide-scale impacts on seabird demographics. *Ecology Letters*, **8(11)**, 1157-1164.
- Walther, K., K. Anger and H.-O. Pörtner, 2010: Effects of ocean acidification and warming on the larval development of the spider crab *Hyas araneus* from different latitudes (54° vs. 79°N). *Marine Ecology Progress Series*, **417**, 159-170.
- Walther, K., F.J. Sartoris and H.-O. Pörtner, 2011: Impacts of temperature and acidification on larval calcification of the spider crab *Hyas araneus* from different latitudes (54° vs. 79°N). *Marine Biology*, **158(9)**, 2043-2053.
- Walther, K., F.J. Sartoris, C. Bock and H.-O. Pörtner, 2009: Impact of anthropogenic ocean acidification on thermal tolerance of the spider crab *Hyas araneus*. *Biogeosciences*, **6(10)**, 2207-2215.
- Wara, M.W., A.C. Ravelo and M.L. Delaney, 2005: Permanent El Niño-like conditions during the Pliocene warm period. *Science*, **309(5735)**, 758-761.
- Ware, D.M. and R.E. Thomson, 2005: Bottom-up ecosystem trophic dynamics determine fish production in the northeast Pacific. *Science*, **308(5726)**, 1280-1284.
- Watanabe, Y.W., M. Shigemitsu and K. Tadokoro, 2008: Evidence of a change in oceanic fixed nitrogen with decadal climate change in the North Pacific subpolar region. *Geophysical Research Letters*, **35(1)**, L01602.
- Watson, A.J., U. Schuster, D.C.E. Bakker, N.R. Bates, A. Corbière, M. González-Dávila, T. Friedrich, J. Hauck, C. Heinze, T. Johannessen, A. Körtzinger, N. Metzl, J. Olafsson, A. Olsen, A. Oschlies, X.A. Padin, B. Pfeil, J.M.

- Santana-Casiano, T. Steinhoff, M. Telszewski, A.F. Rios, D.W.R. Wallace and R. Wanninkhof, 2009: Tracking the variable North Atlantic sink for atmospheric CO₂. *Science*, **326(5958)**, 1391-1393.
- Webb**, A.E., L.R. Leighton, S.A. Schellenberg, E.A. Landau and E. Thomas, 2009: Impact of the Paleocene-Eocene thermal maximum on deep-ocean microbenthic community structure: using rank-abundance curves to quantify paleoecological response. *Geology*, **37(9)**, 783-786.
- Weinbauer**, M.G., X. Mari and J.-P. Gattuso, 2011: Effects of ocean acidification on the diversity and activity of heterotrophic marine microorganisms. In: *Ocean Acidification* [Gattuso, J.-P. and L. Hansson (eds.)]. Oxford University Press, Oxford, UK, pp. 83-98.
- Wernberg**, T., D.A. Smale, F. Tuya, M.S. Thomsen, T.J. Langlois, T. de Bettignies, S. Bennett and C.S. Rousseaux, 2013: An extreme climatic event alters marine ecosystem structure in a global biodiversity hotspot. *Nature Climate Change*, **3(1)**, 78-82.
- Westbrook**, G.K., K.E. Thatcher, E.J. Rohling, A.M. Piotrowski, H. Pälike, A.H. Osborne, E.G. Nisbet, T.A. Minshull, M. Lanoisellé, R.H. James, V. Hühnerbach, D. Green, R.E. Fisher, A.J. Crocker, A. Chabert, C. Bolton, A. Beszczynska-Möller, C. Berndt and A. Aquilina, 2009: Escape of methane gas from the seabed along the West Spitsbergen continental margin. *Geophysical Research Letters*, **36(15)**, L15608.
- Wethey**, D.S., S.A. Woodin, T.J. Hilbish, S.J. Jones, F.P. Lima and P.M. Brannock, 2011: Response of intertidal populations to climate: effects of extreme events versus long term change. *Journal of Experimental Marine Biology and Ecology*, **400(1-2)**, 132-144.
- Whiteley**, N.M., 2011: Physiological and ecological responses of crustaceans to ocean acidification. *Marine Ecology Progress Series*, **430**, 257-271.
- Whitney**, F.A., 2011: Nutrient variability in the mixed layer of the subarctic Pacific Ocean, 1987-2010. *Journal of Oceanography*, **67(4)**, 481-492.
- Williams**, J.W. and S.T. Jackson, 2007: Novel climates, no-analog communities, and ecological surprises. *Frontiers in Ecology and the Environment*, **5(9)**, 475-482.
- Williamson**, P. and C. Turley, 2012: Ocean acidification in a geoengineering context. *Philosophical Transactions of the Royal Society A: Mathematical, Physical, and Engineering Sciences*, **370(1974)**, 4317-4342.
- Williamson**, P., R. Watson, P. Artaxo, R. Bodle, V. Galaz, G. Mace, A. Parker, D. Santillo, C. Vivian, D. Cooper, J. Webbe, A. Cung and E. Woods, 2012: *Impacts of climate-related geoengineering on biological diversity*. Convention on Biological Diversity, 83 pp.
- Wilson**, K.J., J. Falkingham, H. Melling and R. De Abreu, 2004: Shipping in the Canadian Arctic: other possible climate change scenarios. *Geoscience and Remote Sensing Symposium Proceedings*, **3**, 1853-1856.
- Wilson**, R., A. Tudhope, P. Brohan, K. Briffa, T. Osborn and S. Tett, 2006: Two-hundred-fifty years of reconstructed and modeled tropical temperatures. *Journal of Geophysical Research*, **111(C10)**, C10007.
- Wilson**, S.E., D.K. Steinberg and K.O. Buesseler, 2008: Changes in fecal pellet characteristics with depth as indicators of zooplankton repackaging of particles in the mesopelagic zone of the subtropical and subarctic North Pacific Ocean. *Deep-Sea Research Part II: Topical Studies in Oceanography*, **55(14-15)**, 1636-1647.
- Wiltshire**, K., A. Kraberg, I. Bartsch, M. Boersma, H.-D. Franke, J. Freund, C. Gebühr, G. Gerdtts, K. Stockmann and A. Wichels, 2010: Helgoland roads, North Sea: 45 years of change. *Estuaries and Coasts*, **33(2)**, 295-310.
- Witt**, M.J., L.A. Hawkes, M.H. Godfrey, B.J. Godley and A.C. Broderick, 2010: Predicting the impacts of climate change on a globally distributed species: the case of the loggerhead turtle. *Journal of Experimental Biology*, **213**, 901-911.
- Wittmann**, A.C. and H.-O. Pörtner, 2013 in press: Sensitivities of extant animal taxa to ocean acidification. *Nature Climate Change*.
- Woese**, C.R., O. Kandler and M.L. Wheelis, 1990: Towards a natural system of organisms: proposal for the domains Archaea, Bacteria, and Eucarya. *Proceedings of the National Academy of Sciences of the United States of America*, **87(12)**, 4576-4579.
- Wohlrs-Zöllner**, J., P. Breithaupt, K. Walther, K. Jürgens and U. Riebesell, 2011: Temperature and nutrient stoichiometry interactively modulate organic matter cycling in a pelagic algal-bacterial community. *Limnology and Oceanography*, **56(2)**, 599-610.
- Wolf**, S.G., M.A. Snyder, W.J. Sydeman, D.F. Doak and D.A. Croll, 2010: Predicting population consequences of ocean climate change for an ecosystem sentinel, the seabird Cassin's auklet. *Global Change Biology*, **16(7)**, 1923-1935.

- Wolff, G.A., D.S.M. Billett, B.J. Bett, J. Holtvoeth, T. FitzGeorge-Balfour, E.H. Fisher, I. Cross, R. Shannon, I. Salter, B. Boorman, N.J. King, A. Jamieson and F. Chaillan, 2011:** The effects of natural iron fertilisation on deep-sea ecology: The Crozet plateau, Southern Indian Ocean. *PLoS ONE*, **6(6)**, e20697.
- Wood, H.L., J.I. Spicer and S. Widdicombe, 2008:** Ocean acidification may increase calcification rates, but at a cost. *Proceedings of the Royal Society B: Biological Sciences*, **275(1644)**, 1767-1773.
- Wood, R., 1999:** *Reef Evolution*. Oxford University Press, Oxford, U. K., 414 pp.
- Woodworth-Jefcoats, P.A., J.J. Polovina, J.P. Dunne and J.L. Blanchard, 2013:** Ecosystem size structure response to 21st century climate projection: large fish abundance decreases in the central North Pacific and increases in the California Current. *Global Change Biology*, **19(3)**, 724-733.
- Wootton, J.T., C.A. Pfister and J.D. Forester, 2008:** Dynamic patterns and ecological impacts of declining ocean pH in a high-resolution multi-year dataset. *Proceedings of the National Academy of Sciences of the United States of America*, **105(48)**, 18848-18853.
- Worm, B., R. Hilborn, J.K. Baum, T.A. Branch, J.S. Collie, C. Costello, M.J. Fogarty, E.A. Fulton, J.A. Hutchings, S. Jennings, O.P. Jensen, H.K. Lotze, P.M. Mace, T.R. McClanahan, C. Minto, S.R. Palumbi, A.M. Parma, D. Ricard, A.A. Rosenberg, R. Watson and D. Zeller, 2009:** Rebuilding global fisheries. *Science*, **325(5940)**, 578-585.
- Zachos, J.C., U. Röhl, S.A. Schellenberg, A. Sluijs, D.A. Hodell, D.C. Kelly, E. Thomas, M. Nicolo, I. Raffi, L.J. Lourens, H. McCarren and D. Kroon, 2005:** Rapid acidification of the ocean during the Paleocene-Eocene Thermal Maximum. *Science*, **308(5728)**, 1611-1615.
- Zeebe, R.E. and P. Westbroek, 2003:** A simple model for the CaCO₃ saturation state of the ocean: The "Strangelove", the "Neritan", and the "Cretan" Ocean. *Geochemistry Geophysics Geosystems*, **4(12)**, 1104.
- Zeebe, R.E., J.C. Zachos and G.R. Dickens, 2009:** Carbon dioxide forcing alone insufficient to explain Palaeocene-Eocene Thermal Maximum warming. *Nature Geoscience*, **2(8)**, 576-580.
- Zhang, J., D. Gilbert, A.J. Gooday, L. Levin, S.W.A. Naqvi, J.J. Middelburg, M. Scranton, W. Ekau, A. Peña, B. Dewitte, T. Oguz, P.M.S. Monteiro, E. Urban, N.N. Rabalais, V. Ittekkot, W.M. Kemp, O. Ulloa, R. Elmgren, E. Escobar-Briones and A.K. Van der Plas, 2010:** Natural and human-induced hypoxia and consequences for coastal areas: synthesis and future development. *Biogeosciences*, **7(5)**, 1443-1467.
- Zondervan, I., B. Rost and U. Riebesell, 2002:** Effect of CO₂ concentration on the PIC/POC ratio in the coccolithophore *Emiliania huxleyi* grown under light-limiting conditions and different daylengths. *Journal of Experimental Marine Biology and Ecology*, **272(1)**, 55-70.
- Zondervan, I., R.E. Zeebe, B. Rost and U. Riebesell, 2001:** Decreasing marine biogenic calcification: a negative feedback on rising atmospheric pCO₂. *Global Biogeochemical Cycles*, **15(2)**, 507-516.

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

Table 6-1: To assess how a changing climate will alter the ocean's biological pump (Figure 6-4) and determine the resulting biogeochemical feedbacks on global climate, changes in a wide range of processes from cells to ocean basins, and from epipelagic to mesopelagic, must be quantified. This table illustrates the complexity of the integrated knowledge platform needed to provide evidence of these biogeochemical ramifications and thus the present limits to clear conclusions about climate-induced effects on the biological pump (C, Carbon; TEP, Transparent Exopolymer Particle; DOM, Dissolved Organic Matter; POM, Particulate Organic Matter).

Alteration of physiological rates	Biogeographical changes / community shifts	Altered foodweb structure - Trophodynamics	Changes to particle dynamics	Biogeochemical changes / climatic feedbacks
NPP (Bopp <i>et al.</i> , 2002, 2013)	Microbial community structure (Giovannoni and Vergin, 2012)	Altered prey-predator linkages (Lewandowska, and Sommer, 2010)	Faecal pellet geometry (Wilson <i>et al.</i> , 2008)	Particle flux/C sequestration (Bopp <i>et al.</i> , 2002)
Particle solubilization through bacterial ectoenzymes (Christian and Karl, 1995)	Phytoplankton community structure – biomes (Boyd and Doney, 2002)		C partitioning between DOM vs. POM – TEP (Riebesell <i>et al.</i> , 2007)	Shifts in elemental stoichiometry of planktonic communities (Karl <i>et al.</i> , 2003)
TEP production (Engel <i>et al.</i> , 2004)	Alteration of zooplankton biomes (Beaugrand <i>et al.</i> , 2009)		Sinking rates/seawater viscosity (Lam and Bishop, 2008)	Remineralization rate – [O ₂]; hypoxia; nutrient resupply (Gruber, 2011)
Microzooplankton grazing rates (Rose <i>et al.</i> , 2009)	Faunistic shifts at depth (Jackson and Burd, 2001)		Ballasting - calcite versus opal (Klaas and Archer, 2002)	Activity of the microbial loop; vertical carbon export (Grossart <i>et al.</i> , 2006; Piontek <i>et al.</i> , 2010)

Table 6-2: Selected examples of species responses and underlying mechanisms to changing temperature, oxygen level and ocean acidification (OA). References are indicated by superscript numbers and in the footnote.

Phenomenon	Key drivers	Mechanism / Sensitivity
<i>Biogeography</i>		
Northward shift in the distribution of North Sea cod (<i>Gadus morhua</i>) stocks between 1977 and 2001 ^{1,2}	Temperature	Bottlenecks of high sensitivity during early life stages as well as adult spawning stage in winter/early spring
Shift from sardines (<i>Sardinops melanostictus</i>) to anchovies (<i>Engraulis japonicus</i>) in the western North Pacific observed between 1993 and 2003 ^{3,4}	Temperature	Thermal windows of growth and reproductive output are found at higher temperatures for anchovies than sardines, food preferences of the competing species being similar.
Variable sensitivity of Pacific tuna species to the availability of dissolved O ₂ . Bigeye tuna routinely reach depths where ambient O ₂ content is below 1.5 ml L ⁻¹ (≈ 60 μmoles kg ⁻¹) ^{5,6}	Oxygen	Oxygen transport via hemoglobin is adapted to be highly efficient supporting high metabolic rates as needed during feeding in the OMZ.
Northward movement of species and the conversion of polar into more temperate and temperate into more subtropical system characteristics in the European Large Marine Ecosystems between 1958–2005 ^{7,8}	Warming and current advection	Effects are attributed to climate change but may be influenced by nutrient enrichment and overfishing.

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

<u>Abundance</u>		
Increase in abundance of arctic boreal plankton species, notably the copepods <i>Calanus hyperboreus</i> , <i>Calanus glacialis</i> and the dinoflagellate <i>Ceratium arcticum</i> between 1960 and 2000 in the Newfoundland Shelf, Northwest Atlantic ^{9,10}	Temperature	Temperature sensitivity of phyto- and zooplankter resulting from cooling due to increased influx of Arctic water
A benthic fish species, the eelpout (<i>Zoarces viviparus</i>) at its southern distribution limit, the German Wadden Sea, displayed abundance losses during warming periods and rising summer extreme temperatures between 1993 and 2005, with early disappearance of the largest individuals ¹¹	Temperature	Temperature extremes exceed organism's thermal windows, with largest individuals being relatively less tolerant to high temperature than smaller individuals.
Variable sensitivities to OA within and across animal phyla (Figure 6-10 B) ^{12 - 21}	Anthropogenic OA, Sea water acidification by elevated $p\text{CO}_2$ in OMZs, upwelling areas, involving anthropogenic ocean acidification.	<ul style="list-style-type: none"> – Lowered extracellular (blood plasma) pH causing a lowering of the rates of ion exchange and metabolism in muscle or liver (hepatocytes) of vertebrates and invertebrates. – High sensitivity at reduced energy turnover in tissues and/or whole organism by reduced ion exchange, – use of more energy efficient transport mechanisms, – reduced protein synthesis, enhanced nitrogen release from amino acid catabolism and protein degradation, – slower growth
<u>Phenology</u>		
Migration time of pink salmon (<i>Oncorhynchus gorbuscha</i>) in Alaska is almost two weeks earlier in 2010s relative to 40 years ago ²²	Warming	Rapid microevolution for earlier migration timing.
In the waters around the U.K., during a period of warming between 1976 and 2005, the seasonal timing of biological events of all major marine taxonomic groups (plant/phytoplankton, invertebrate and vertebrates) advanced on average, by 0.31 to 0.43 days year ⁻¹ ²³	Warming	Sensitivity to seasonal temperature changes as a result of specific thermal windows of different organisms;
<u>Body size and growth</u>		
Asymptotic size of different populations of Atlantic cod (<i>Gadus morhua</i>) and Atlantic Herring (<i>Clupea harengus</i>) are negatively related to temperature ^{24, 25}	Warming	At large body size, oxygen supply limitations are exacerbated and the organism reaches its long-term heat tolerance limits at lower temperatures, thus limiting the maximum body size that can be reached.

1. Perry *et al.*, 2005; 2. Pörtner *et al.*, 2008; 3. Takasuka *et al.*, 2007; 4. Takasuka *et al.*, 2008; 5. Lehodey *et al.*, 2011; 6. Seibel, 2011; 7. Beaugrand *et al.*, 2009; 8. Philippart *et al.*, 2011; 9. Johns *et al.*, 2001; 10. Greene and Pershing, 2003; 11. Pörtner and Knust, 2007; 12. Reipschläger and Pörtner, 1996; 13. Pörtner *et al.*, 2000; 14. Vezzoli *et al.*, 2004; 15. Langenbuch and Pörtner, 2003; 16. Fernández-Reiriz *et al.*, 2011; 17. Langenbuch and Pörtner, 2002; 18. Langenbuch *et al.*, 2006; 19. Michaelidis *et al.*, 2005; 20. Pörtner *et al.*, 1998; 21. Stumpp *et al.*, 2012; 22. Kovach *et al.*, 2012; 23. Thackeray *et al.*, 2010; 24. Taylor 1958; 25. Brunel and Dickey-Collas, 2010.

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

Table 6-3: Tolerances to ocean acidification in marine taxa, assessed from laboratory and field studies of species in the $p\text{CO}_2$ range from <650 to $>10\,000$ μatm , compared to present day atmospheric levels of 400 μatm . (It should be noted that anthropogenic CO_2 emissions add to the natural variability of CO_2 concentrations in marine environments, which can reach much higher than atmospheric levels). Variables studied include growth, survival, calcification, metabolic rate, immune response, development, abundance, behavior and others. Neither all life stages, nor all variables, including the entire range of CO_2 concentrations, were studied in all species. *Confidence* is based on the number of studies, the number of species studied and the agreement of results within one group. +: denotes that possibly more species or strains (genetically distinct populations of the same species) were studied, as only genus or family were specified; beneficial: most species were positively affected; vulnerable: more than 5% of species in a group will be negatively affected by 2100; tolerant: more than 95% of species will not be affected by 2100. RCP 6.0: representative concentration pathway with projected atmospheric $p\text{CO}_2 = 670$ μatm ; RCP 8.5: $p\text{CO}_2 = 936$ μatm in 2100 (Meinshausen *et al.*, 2011). *Confidence* is limited by the short- to medium-term nature of various studies and the lack of sensitivity estimates on evolutionary timescales, i.e., across generations (see separate reference list, online supplementary material). Note that the assessment of variability between species from the same animal phylum has revealed an increase in the fraction of sensitive species with rising CO_2 levels, see Figure 6-10. *Rather than a sensitivity threshold the entire range of investigated $p\text{CO}_2$ values is given for groups of photosynthetic organisms. In all studies photosynthetic rates are stimulated to different, species-specific degrees by elevated $p\text{CO}_2$, indicating low vulnerability. Coccolithophores and calcifying algae are assessed as being more sensitive than other photosynthetic organisms due to reduced calcification and shell dissolution. NA, not available, *confidence levels for fishes were converted from medium to low, in light of uncertainty on the long-term implications of behavioral disturbances.

Taxon	No. of studies	No. of parameters studied	Total no. of species studied	$p\text{CO}_2$ where the most vulnerable species is negatively affected or investigated $p\text{CO}_2$ range* (μatm)	Assessment of tolerance to RCP 6.0 (<i>confidence</i>)	Assessment of tolerance to RCP 8.5 (<i>confidence</i>)
Cyanobacteria	17	5	9+	180-1250*	Beneficial (<i>low</i>)	Beneficial (<i>low</i>)
Coccolithophores	35	6	7+	740	Tolerant (<i>low</i>)	Vulnerable (<i>medium</i>)
Diatoms	22	5	28+	150-1500*	Tolerant (<i>low</i>)	Tolerant (<i>low</i>)
Dinoflagellates	12	4	11+	150-1500*	Beneficial (<i>low</i>)	Tolerant (<i>low</i>)
Foraminifers	11	4	22	588	Vulnerable (<i>low</i>)	Vulnerable (<i>medium</i>)
Seagrasses	6	6	5	300-21000*	Beneficial (<i>medium</i>)	Beneficial (<i>low</i>)
Macroalgae (non-calcifying)	21	5	21+	280-20812*	Beneficial (<i>medium</i>)	Beneficial (<i>low</i>)
Macroalgae (calcifying)	38	10	36+	365	Vulnerable (<i>medium</i>)	Vulnerable (<i>high</i>)
Warm-water corals	45	13	31	467	Vulnerable (<i>medium</i>)	Vulnerable (<i>high</i>)
Cold-water corals	10	13	6	445	Vulnerable (<i>low</i>)	Vulnerable (<i>medium</i>)
Annelids	10	6	17+	1200	Tolerant (<i>medium</i>)	Tolerant (<i>medium</i>)
Echinoderms	54	14	35	510	Vulnerable (<i>medium</i>)	Vulnerable (<i>high</i>)
Mollusks (benthic)	72	20	38+	508	Vulnerable (<i>medium</i>)	Vulnerable (<i>high</i>)
Mollusks (pelagic)	7	8	8	550	Vulnerable (<i>low</i>)	Vulnerable (<i>medium</i>)
Mollusks (cephalopods)	10	8	5	2200 (850 for trace elements)	Tolerant (<i>medium</i>)	Tolerant (<i>medium</i>)
Bryozoans	7	3	8+	549	Tolerant (<i>low</i>)	Vulnerable (<i>low</i>)
Crustaceans	47	27	44+	700	Tolerant (<i>medium</i>)	Tolerant (<i>low</i>)
Fish ⁺	51	16	40	700	Vulnerable (<i>low</i>)	Vulnerable (<i>low</i>)

Table 6-4: Potential interactions between modes of anthropogenic forcing (environmental; foodwebs, harvesting) on different levels of biological organisation. These interactions, from simple to complex, are illustrated with examples from the published literature. E, O, and M denote studies using manipulation Experiments (lab or field), Observations, or Modelling approaches, respectively. Unknown denotes no published information is available for each of these categories. NA denotes Not Applicable for this category.

Biological Organisation studied at ecosystem level	Anthropogenic Forcing			
	Single Environmental Driver	Multiple Environmental Drivers	Fishing / Foodwebs	Fishing / Climate change
<i>Individuals</i>	Lab experiments and field observations show that warming alters organismal physiology and thereby, growth (Pörtner and Knust, 2007) E & O	Shipboard manipulation experiment addressing interactive effects of temperature and CO ₂ on coccolithophore calcification (Feng <i>et al.</i> , 2009) E	NA	Unknown
<i>Population</i>	Physiological effects of warming change population abundance <i>in situ</i> , (Pörtner and Knust, 2007) E & O. Lab cultures show how altered pH elicits different responses of coccolithophore species (Langer <i>et al.</i> , 2006) E	Lab cultures show differential responses of cyanobacterial groups to temperature and CO ₂ (Fu <i>et al.</i> , 2007) E	Altered maturation age and growth rate of populations due to fishing (Fairweather <i>et al.</i> , 2006; Hseih <i>et al.</i> , 2006) O	Interactive effects on cod populations of fishing and alteration of salinity (Lindegren <i>et al.</i> , 2010) M
<i>Ecosystem</i>	Mesocosm experiments simulating the effect of individual drivers (e.g. ocean acidification effects on benthos: Christen <i>et al.</i> 2013; and on pelagic communities: Riebesell <i>et al.</i> , 2013) E, O or M	Mesocosm experiments studying differential effects of light and temperature, on copepods versus diatoms (Lewandowska and Sommer, 2010) E	Effects of fishing on ecosystem structure – trophic cascades (Frank <i>et al.</i> , 2005) O	Interplay of fishing and climate pressures on ecosystems promotes lower trophic levels (Kirby <i>et al.</i> , 2009) O; enhances diversity loss in benthic communities (Griffith <i>et al.</i> , 2011) M
<i>Biome</i>	Time-series observations on warming and geographical shifts of zooplankton biomes (Beaugrand <i>et al.</i> , 2009) O	Unknown	Unknown	Unknown

Table 6-5: Challenges for the oceans that will arise from the employment of a range of geoengineering methods (SRM, solar radiation management, CDR, carbon dioxide removal).

Topic	Brief Description	Challenge and Impact	References
Solar radiation management techniques	Deflection of approximately 1.8 per cent of sunlight, by various techniques, is able to offset the global mean temperature effects of a doubling of atmospheric carbon dioxide content from preindustrial values	Will leave ocean acidification unabated (<i>high confidence</i>). Response of primary production to light reduction unclear.	Crutzen, 2006; Caldeira and Wood, 2008
Ocean storage by direct injection	Capture of CO ₂ post-combustion from mainly coastal power plants, followed by injection of liquid CO ₂ by pipeline or from a ship into the deep ocean.	Will add to ocean acidification and create localized harm to marine life (<i>high confidence</i>). Quantities will be small relative to the atmospheric invasion signal. CO ₂ injected will dissolve and be transported by ocean circulation with eventual surface exposure.	Caldeira <i>et al.</i> , 2005
Sub-sea geologic storage	Capture of CO ₂ from extracted gas or from post-combustion followed by well injection into a porous submarine aquifer beneath impermeable geologic strata.	Extensive experience in place from the Norwegian Sleipner field activity in the North Sea. No evidence of ocean impact from leakage to date.	Benson <i>et al.</i> , 2005
Ocean Fertilization	Spreading of trace amounts of reduced iron, over very large areas of the surface ocean where excess nutrients occur. Overcoming the local iron deficiency creates extensive phytoplankton blooms drawing down sea surface pCO ₂ . Fertilization can also be carried out by using direct or indirect (ocean pipes) addition of macronutrients to oceanic regions where they are depleted.	Much of the exported organic matter is remineralized at shallow depths creating local oxygen stress and shallow CO ₂ enrichment, methane and N ₂ O production. These effects are temporary and the effective retention time is short. If sustained, reduced surface-ocean and increased deep-ocean acidification. O ₂ loss in ocean interior (<i>medium confidence</i>).	de Baar <i>et al.</i> , 1995; de Baar <i>et al.</i> , 2005; Pörtner <i>et al.</i> , 2005; Boyd <i>et al.</i> , 2007; Buesseler <i>et al.</i> , 2008; Law, 2008; Cao and Caldeira, 2010
Artificial upwelling or downwelling	Ocean fertilization by bringing nutrient rich deep water (from 200 - 1000 m) to the surface. Downwelling occurs in parallel, transporting physically dissolved CO ₂ into the deep ocean.	Deep water contains high levels of CO ₂ , which if released counteracts the binding of CO ₂ by fertilization. No evidence available.	Lovelock and Rapley, 2007 Oschlies <i>et al.</i> , 2010
Sequestration of organic carbon	Storage of terrestrial biomass in the coastal or deep ocean	Physical impact, regional loss of oxygen, CO ₂ accumulation and acidification during degradation, increases in methane, N ₂ O and H ₂ S. No evidence available.	Metzger and Benford, 2001; Strand and Benford, 2009
Carbonate neutralization	Dissolution of power plant flue gas into sea water yielding an acidic solution which is neutralized by addition of crushed limestone. The resulting bicarbonate rich fluid is discharged to the ocean.	Involves the transport and crushing to fine scale of large quantities of limestone and the processing of very large quantities of sea water. Environmental impact issues not yet explored.	Rau, 2011
Accelerated olivine weathering	Uses wind powered electrochemical processes to remove HCl from the ocean and neutralizes the acid with silicate minerals such as olivine for disposal. The net result is to add alkalinity to the ocean akin to natural silicate weathering processes.	Complex system as yet untested in pilot processes. Involves mining and crushing large quantities of silicate minerals. Very long time scale consequences uncertain.	House <i>et al.</i> , 2007; Köhler <i>et al.</i> , 2010

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

Table 6-6: Coastal and oceanic key risks from climate change and the potential for risk reduction through mitigation and adaptation. Key risks are identified based on assessment of the literature and expert judgments made by authors of the various WGII AR5 chapters, with supporting evaluation of evidence and agreement in the referenced chapter sections. Each key risk is characterized as very low, low, medium, high, or very high. Risk levels are presented for the near-term era of committed climate change (here, for 2030-2040), in which projected levels of global mean temperature increase do not diverge substantially across emissions scenarios. Risk levels are also presented for the longer-term era of climate options (here, for 2080-2100), for global mean temperature increase of 2°C and 4°C above preindustrial levels. For each timeframe, risk levels are estimated for the current state of adaptation and for a hypothetical highly adapted state. As the assessment considers potential impacts on different physical, biological, and human systems, risk levels should not necessarily be used to evaluate relative risk across key risks. Relevant climate variables are indicated by symbols. Acronyms for oceans sub-regions are as follows: HLSBS: High-Latitude Spring Bloom Systems; EUS: Equatorial Upwelling Systems; SES: Semi-Enclosed Seas; CBS: Coastal Boundary Systems; EBUE: Eastern Boundary Upwelling Ecosystems; STG: Sub-Tropical Gyres, DS: Deep Sea (>1000 m).

Key risk	Adaptation issues and prospects	Climatic drivers	Supporting ch. sections	Timeframe	Risk for current and high adaptation			
Risks to ecosystems and adaptation options								
Changes in ecosystem productivity associated with the redistribution and loss of net primary productivity in open oceans (medium confidence)	Adaptation options are limited to the translocation of industrial fishing activities due to regional decreases (low latitude) versus increases (high latitude) in productivity, or to the expansion of aquaculture.		6.5.1, 6.3.4, Box CC-NPP	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Distributional shift in fish and invertebrate species, fall in fishery catch potential at low latitudes, e.g., in EUS, CBS and STG regions (high confidence)	Evolutionary adaptation potential of fish and invertebrate species to warming is limited as indicated by their ongoing latitudinal shifts. Human adaptation options involve the large scale translocation of industrial fishing activities following the regional decreases (low latitude) versus (possibly transient) increases (high latitude) in catch potential as well deploying flexible management that can react to variability and change. Further options include improving fish resilience to thermal stress by reducing other stressors like pollution and eutrophication, or the expansion of aquaculture.		6.3.1, Box CC-Bio	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
High mortalities and loss of habitat to larger fauna including commercial species due to hypoxia expansion and effects (high confidence)	Human adaptation options involve the large scale translocation of industrial fishing activities as a consequence of the hypoxia induced decreases in biodiversity and fisheries catch of pelagic fish and squid. Special fisheries may benefit (Humboldt squid). Reducing the amount of organic carbon running of coastlines by controlling nutrients and pollution running off agricultural areas can reduce microbial activity and consequently limit the extent of the oxygen drawdown and the formation of coastal dead zones.		6.3.3, 30.5.3.2, 30.5.4.1-2	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Ocean acidification: Reduced growth and survival of commercially valuable shellfish and other calcifiers, e.g., reef building corals, calcareous red algae (high confidence)	Evidence for differential resistance and evolutionary adaptation of some species exists but is likely limited by the CO2 concentrations and high temperatures reached; adaptation options include the shift to exploiting more resilient species or the protection of habitats with low natural CO2 levels, as well as the reduction of other stresses, mainly pollution and limiting pressures from tourism and fishing.		5.3.3.5, 6.X.X., 30.X.X, Box CC-OA	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Reduced biodiversity, fisheries abundance and coastal protection by coral reefs due to heat induced mass coral bleaching and mortality increases, e.g., in CBS, SES, and STG regions (high confidence)	Evidence of rapid evolution by corals is very limited or non-existent. Some corals may migrate to higher latitudes. However, the movement of entire reef systems is unlikely given estimates that they need to move at the speed of 10-20 km per year. Human adaptation options are limited to reducing other stresses, mainly enhancing water quality and limiting pressures from tourism and fishing. This option will delay the impacts of climate change by a few decades but is likely to disappear as thermal stress increases.		5.4.2.4, 6.4.2, 30.3.1.1, 30.5.2, 30.5.3, 30.5.4, 30.5.6, Box CC-CR	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Coastal inundation and habitat loss due to sea level rise and intensified precipitation events, e.g., in CBS and STG subregions (medium to high confidence)	Options to maintain ecosystem integrity are limited to the reduction of other stresses, mainly pollution and limiting pressures from tourism, fishing and aquaculture. Loss of ecosystems such as sea grass, mangroves and coral reefs can be reduced by reducing deforestation and increasing reforestation of river catchments and coastal areas to retain sediments and nutrients.		5.5.2, 5.5.4, CC-CR, 30.5.6.1.3, 30.6.2.2	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Marine biodiversity loss with high rate of climate change (medium confidence)	Adaptation options are limited to the reduction of other stresses, mainly to reducing pollution and to limiting pressures from tourism and fishing.		Box CC-Bio, 6.3.1, 30.X.X	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Climatic drivers of impacts				Risk & potential for adaptation				
								Potential for adaptation to reduce risk Risk level with high adaptation Risk level with current adaptation

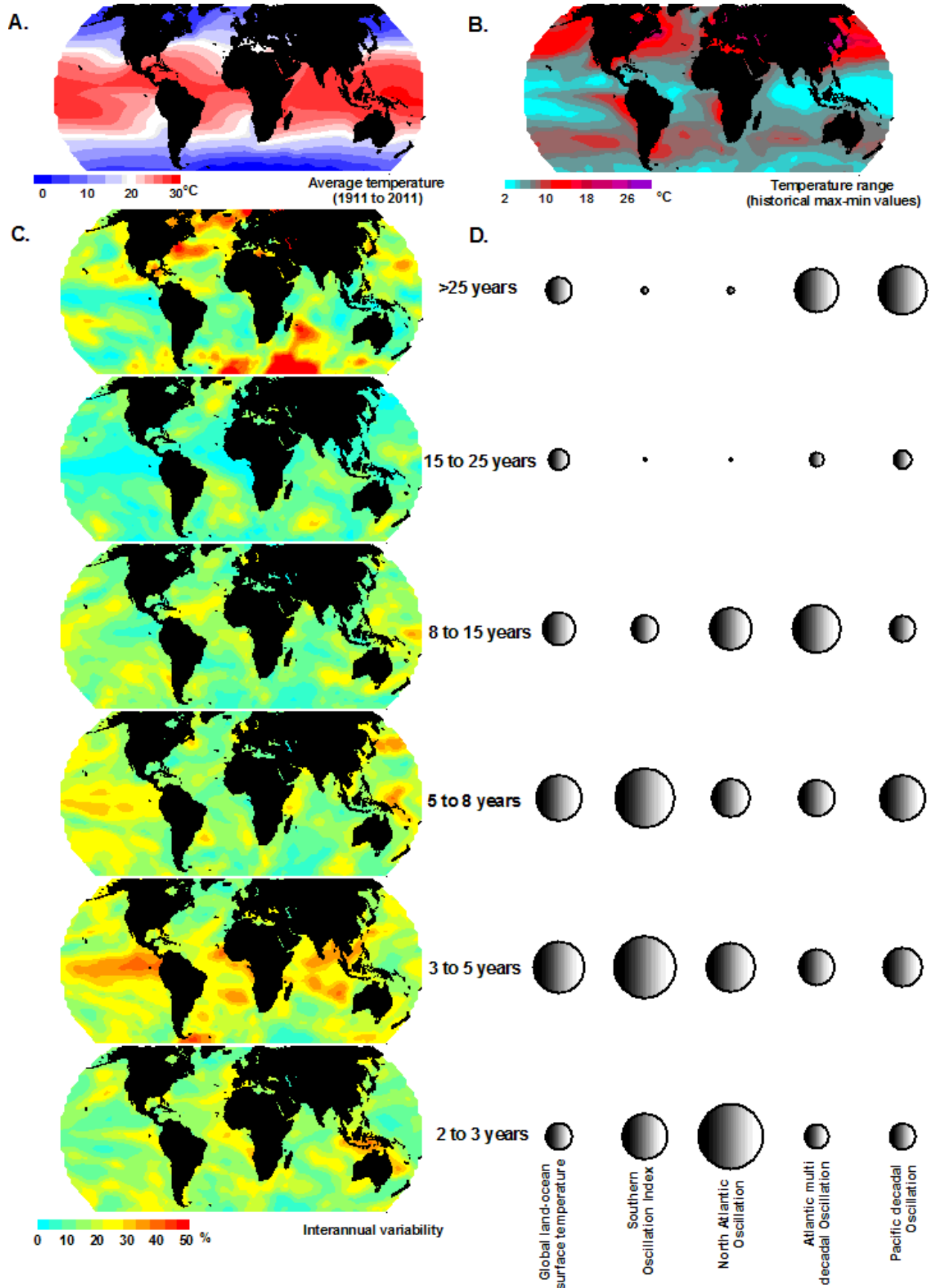
Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

Key risk	Adaptation issues and prospects	Climatic drivers	Supporting ch. sections	Timeframe	Risk for current and high adaptation			
Risks to fisheries								
Decreased production of global shellfish fisheries (<i>high confidence</i>)	Effective shift to alternative livelihoods, changes in food consumption patterns and adjustment of (global) markets.		6.3.2, 6.3.5, 6.4.1.1, 30.5.5, 30.6.2.1, CC-OA	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Global redistribution and decrease of low latitude fisheries yields are paralleled by a global trend to catches having smaller fishes (<i>medium confidence</i>)	Increasing coastal poverty at low latitudes as fisheries becomes smaller - partially compensated by the growth of aquaculture and marine spatial planning, as well as enhanced industrialized fishing efforts.		6.3.1, 6.4.1, 6.5.3, 30.5.4, 30.5.6, 30.6.2	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Redistribution of catch potential of large pelagic-highly migratory fish resources, such as tropical Pacific tuna fisheries (<i>high confidence</i>)	International fisheries agreements and instruments, such as the tuna commissions, may have limited success in establishing sustainable fisheries yields.		6.3.1, 6.4.3, Table 30.4	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Variability of small pelagic fishes in Eastern Boundary Upwelling systems is becoming more extreme at interannual to multidecadal scales, making industry and management decisions more uncertain (<i>medium confidence</i>)	Development of new and specific management tools and models may have limited success to sustain yields. Reduction in fishing intensity increases resilience of the fisheries.		6.3.2, 6.3.3, 30.5.2, 30.5.5, CC-UP	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Decrease in catch and species diversity of fisheries in tropical coral reefs, exacerbated by interactions with other human drivers such as eutrophication and habitat destruction (<i>high confidence</i>)	Restoration of overexploited fisheries and reduction of other stressors on coral reefs delay ecosystem changes. Human adaptation includes the usage of alternative livelihoods and food sources (e.g., coastal aquaculture).		6.4.1, 30.5.3-4, 30.5.6, CC-CR	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Current spatial management units, especially the MPAs may fail in the future due to shifts in species distribution and community structure (<i>high confidence</i>)	Continuous revision and shifts of MPA borders, and of MPA goals and performance.		6.3.1, 6.4.2.1, 30.5.1, CC-BIO	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Risks to humans and infrastructure								
Coastal socio-economic security (<i>high confidence</i>)	Human adaptation options involve (1) Protection using coastal defences (e.g. seawalls) and soft measures (e.g. mangrove replanting and enhancing coral growth), (2) Accommodation to allow continued occupation of coastal areas by making changes to human activities and infrastructure, and (3) Managed retreat as a last viable option. Vary from large-scale engineering works to smaller scale community projects. Options are available under the more traditional CZM (coastal zone management) framework but increasingly under DRR (disaster risk reduction) and CCA (climate change adaptation) frameworks.		5.5.2, 5.5.4, 30.6.5, 30.7.1	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
*high confidence in existence of adaptation measures, low confidence in magnitude of risk reduction								
Reduced livelihoods and increased poverty (<i>medium confidence</i>)	Human adaptation options involve the large scale translocation of industrial fishing activities following the regional decreases (low latitude) versus increases (high latitude) in catch potential and shifts in biodiversity. Artisanal local fisheries are extremely limited in their adaptation options by available financial resources and technical capacities, except for their potential shift to other species of interest.		6.4.1-2, 30.6.2, 30.6.5	Present Near-term (2030-2040) Long-term (2080-2100) 2°C 4°C	Very low, Medium, Very high			
Climatic drivers of impacts				Risk & potential for adaptation				
Warming trend	Extreme temperature	Precipitation	Extreme precipitation	Damaging cyclone	Sea level	Hypoxia	Ocean acidification	<p>Potential for adaptation to reduce risk</p> <p>Risk level with high adaptation</p> <p>Risk level with current adaptation</p>

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

Key risk	Adaptation issues and prospects	Climatic drivers	Supporting ch. sections	Timeframe	Risk for current and high adaptation			
Risks to humans and infrastructure (continued)								
Impacts due to increased frequency of harmful algal blooms (<i>medium confidence</i>)	Adaptation options include improved monitoring and early warning system, reduction of stresses favouring harmful algal blooms, mainly pollution and eutrophication, as well as the avoidance of contaminated areas and fisheries products.		6.4.2.3		Very low Medium Very high			
				Present				
				Near-term (2030-2040)				
				Long-term (2080-2100)	2°C 4°C 			
Impacts on marine resources threatening regional security as territorial disputes and food security challenges increase (<i>limited evidence, medium agreement</i>)	Decrease in marine resources, movements of fish stocks and opening of new seaways, and impacts of extreme events coupled with increasing populations will increase the potential for conflict in some regions, drive potential migration of people and increase humanitarian crises.		AR5 SREX, 30.6.5, 12.4-12.6, 29.3		Very low Medium Very high			
				Present				
				Near-term (2030-2040)				
				Long-term (2080-2100)	2°C 4°C 			
Impacts on shipping and infrastructure for energy and mineral extraction increases as storm intensity and wave height increase in some regions (e.g., high latitudes) (<i>high confidence</i>)	Adaptation options are to limit activities to particular times of the year and/or develop strategies to decrease the vulnerability of structures and operations.		AR5 SREX, 30.6.5, 12.4-12.6, 29.3		Very low Medium Very high			
				Present				
				Near-term (2030-2040)				
				Long-term (2080-2100)	2°C 4°C 			
Climatic drivers of impacts				Risk & potential for adaptation				
Warming trend	Extreme temperature	Precipitation	Extreme precipitation	Damaging cyclone	Sea level	Hypoxia	Ocean acidification	

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014



Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

Figure 6-1: Sea surface temperature variability between 1911 and 2011. The top left map shows the sea surface temperature average for the period. The top right map illustrates the temperature range calculated as the difference between the maximum and minimum values for each grid component during the century. The spatial distribution of variability by time scales (left hand map series, based on the Extended Reynolds Sea Surface Temperature, NOAA, 2012) corresponds to the multidecadal (25 to 40 years), bidecadal (15 to 25 years), decadal (8 to 15 years), low ENSO (El Niño Southern Oscillation) frequency (5 to 8 years), high ENSO frequency (3 to 5 years), and very high frequency (2 to 3 years) scales. The summed variabilities from the same 2°x2° box in all six maps corresponds to 100% of the time series variability. The areas of the right hand bubbles show the spectral density of some of the most widely used climate indices, accumulated in the same frequency windows: the Global Average Temperature and SST Anomalies (GSST), the Southern Oscillation Index (SOI), the North Atlantic Oscillation (NAO), the Multidecadal Atlantic Oscillation (AMO), and the Pacific Decadal Oscillation (PDO). The sum of bubble surface areas for each vertical column (each climate index) corresponds to 100% of the time series variability between the 2 and 40 year periods. Climate indices were obtained from the NOAA ESRL Physical Sciences Division website. **[Illustration to be redrawn to conform to IPCC publication specifications.]**

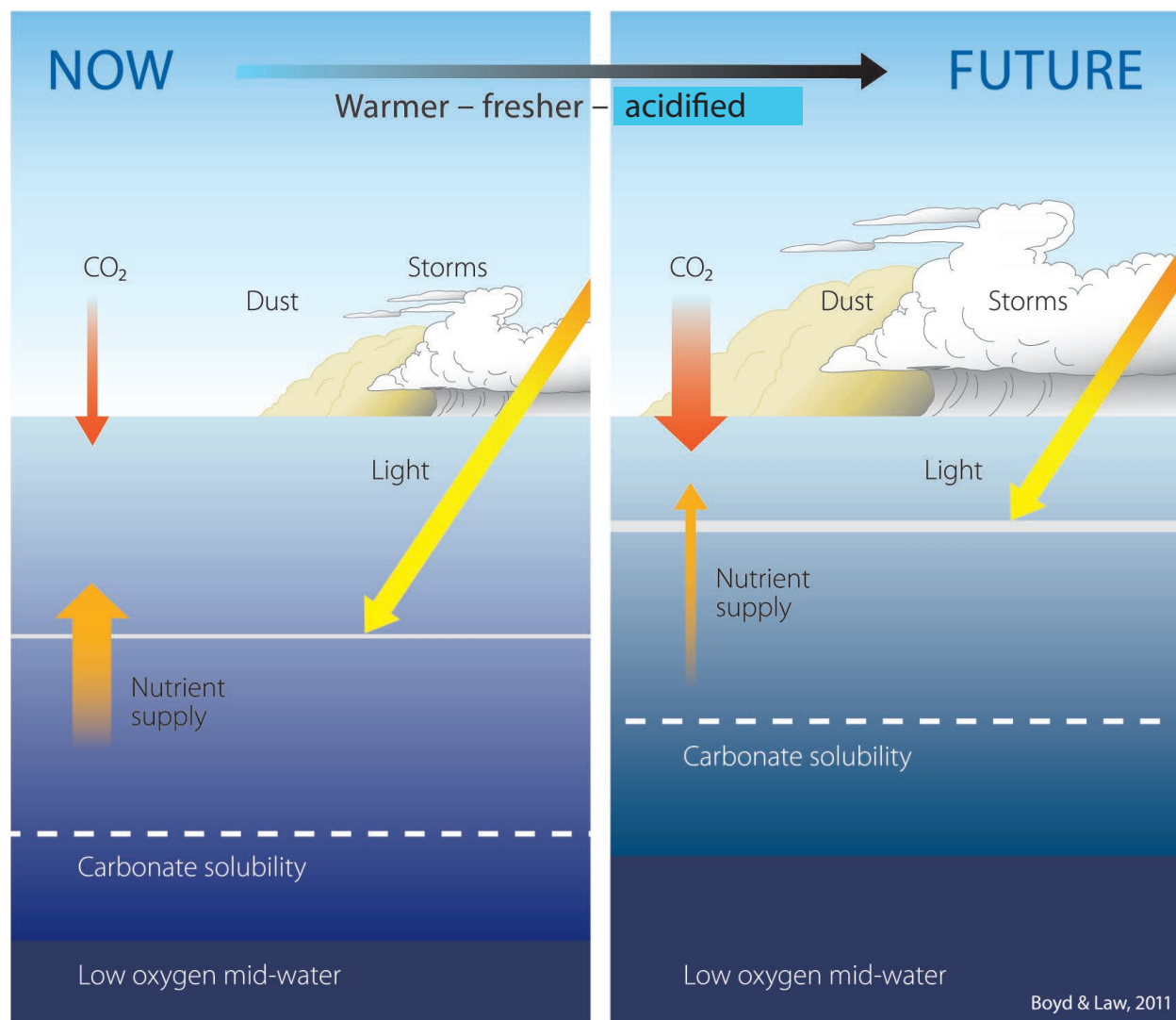


Figure 6-2: Projected alteration (magnitude and frequency) of oceanic fluxes and atmospheric events due to a changing climate in the coming decades. Ocean properties will be altered from the sunlit surface layer to the mid-water stratum. In the surface ocean, the depth of the mixed layer (solid horizontal line) will shallow resulting in higher mean light levels. Increased density stratification (i.e., a strengthening sea water density gradient represented by the increasing thickness of the solid horizontal line) will reduce the vertical supply of nutrients for photosynthesizing organisms residing in the mixed layer. Anthropogenic CO_2 will acidify, i.e., lower the pH of the surface ocean (note this happens in a pH range higher than 7 such that oceans will remain alkaline but less so due to acidification). The penetration of acidified waters to depth will result in a shallower depth (dashed horizontal line) at which calcium carbonate structures, such as shells, dissolve. At depth, the location of low oxygen waters will progressively become shallower. In addition, changes in storm activity and dust deposition will influence ocean physics and chemistry, with consequent effects on ocean biota and hence ecosystems (courtesy of Reusch and Boyd, 2013). **[Illustration to be redrawn to conform to IPCC publication specifications.]**

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

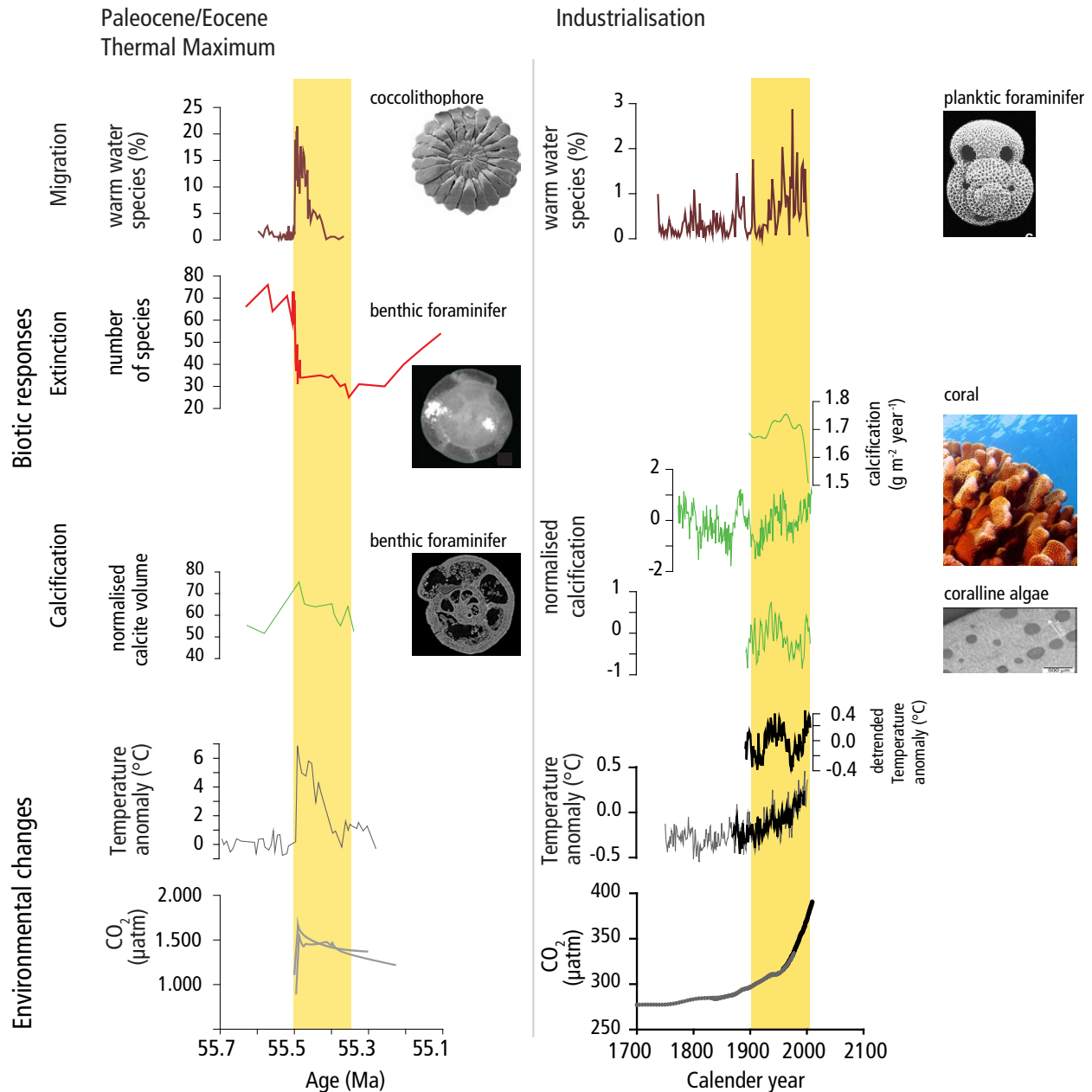


Figure 6-3: Environmental changes (bottom) and associated biological responses (top) for the Paleocene Eocene Thermal Maximum (PETM, left) and the industrial era (right). The PETM represents the best geological analogue for the future ocean in terms of biological responses. Episodes of largest environmental change are indicated with yellow bars. Note the different time scale between the two columns. Both time intervals are characterized by rapid warming both on land and in the ocean and increases in CO₂. Atmospheric CO₂ and temperature are described with direct measurements (black), proxy reconstructions (grey) and model results (light grey). For the recent anthropocenic record, the AMO is shown to highlight high frequency natural temperature fluctuations (Enfield *et al.*, 2001) and their influence on marine biota. Biological responses to the environmental forcing are divided into calcification, extinction and migration. Note the highly group-specific responses to the environmental change, especially with regards to calcification with decreases, increases and high variability. While there was extinction during the PETM, there is currently no evidence for climate-related extinction in the marine record. The warming led to migration of warm water species into previous cold water habitats indicating range expansion due to warming

(modern: planktic foraminifera, *Gs. ruber* Field *et al.*, 2006; PETM: coccolithophores, *Discoaster* spp. Bralower, 2002). CO₂ data: Mauna Loa (Keeling *et al.*, 2005), ice core records (Etheridge *et al.* 1996) and model output for the PETM (Zeebe *et al.*, 2009; Ridgwell and Schmidt, 2010). Sea surface temperatures modern: Wilson *et al.*, 2006 and PETM: Kennett and Stott, 1991. Biotic responses: growth changes in coralline algae (Halfar *et al.*, 2011, bottom) and corals (Vásquez-Bedoya *et al.*, 2012, middle, De'ath *et al.*, 2009, top); calcification changes (Foster *et al.*, 2013) and extinction in benthic foraminifers (Thomas, 2003) for the PETM.

[Illustration to be redrawn to conform to IPCC publication specifications.]

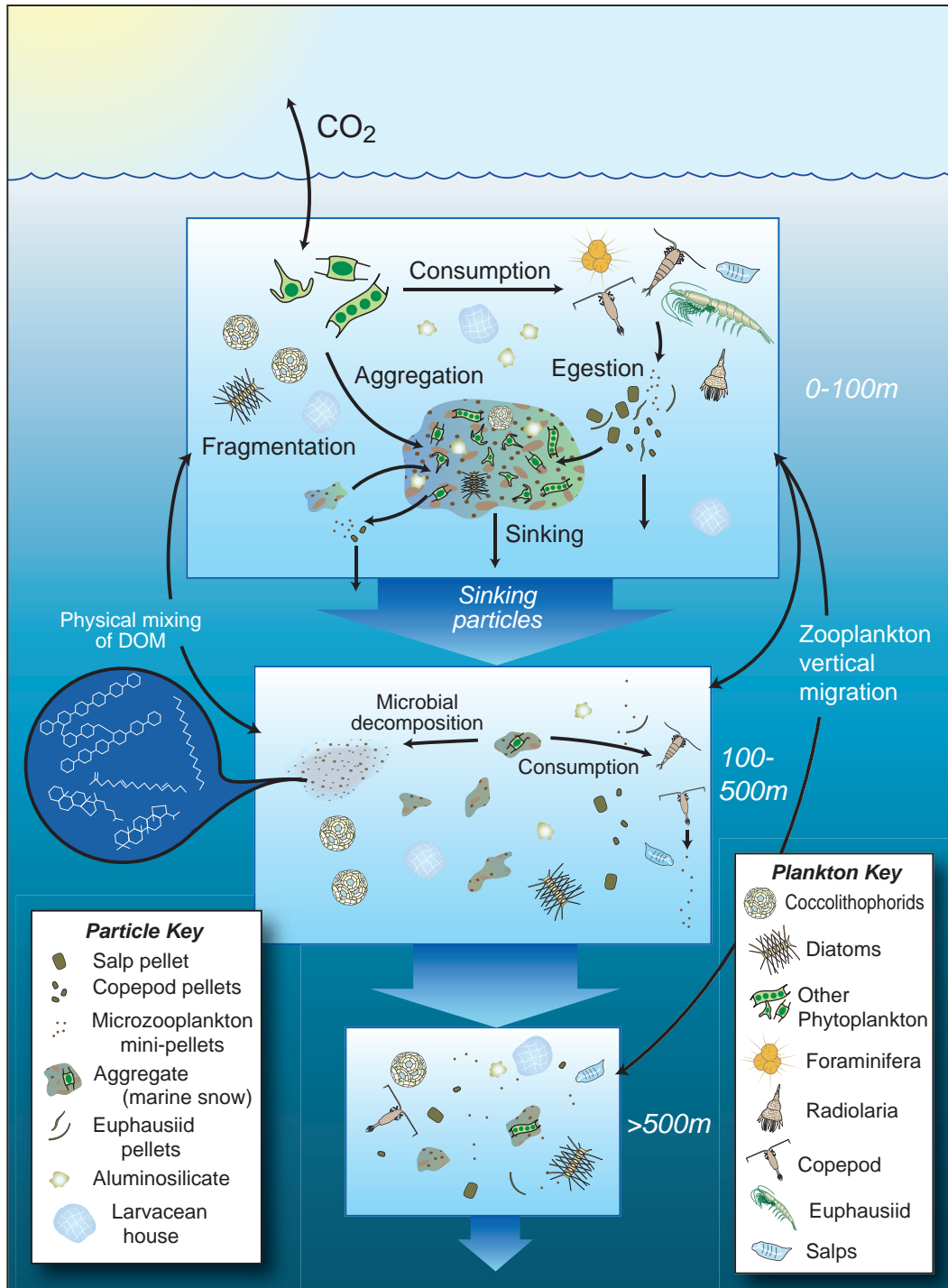


Figure 6-4: A schematic representation of the ocean's biological pump, which will be influenced by climate change and is a conduit for carbon sequestration. It is difficult to project how the pump might be altered (Table 6-1) and whether it would represent a positive or negative feedback to climate change through the cumulative effects of affected processes, surface to depth (Table 6-1): shifts in Net Primary Production, floristic and faunistic community composition in the pelagic realm, and in grazing rates; alterations to the ballasting of settling particles and the proportion of NPP released as DOM (Dissolved Organic Matter); modified bacterial enzymatic rates and particle solubilization; faunistic shifts at depth. Modified from Buesseler *et al.* (2008) by J. Cook (WHOI).

[Illustration to be redrawn to conform to IPCC publication specifications.]

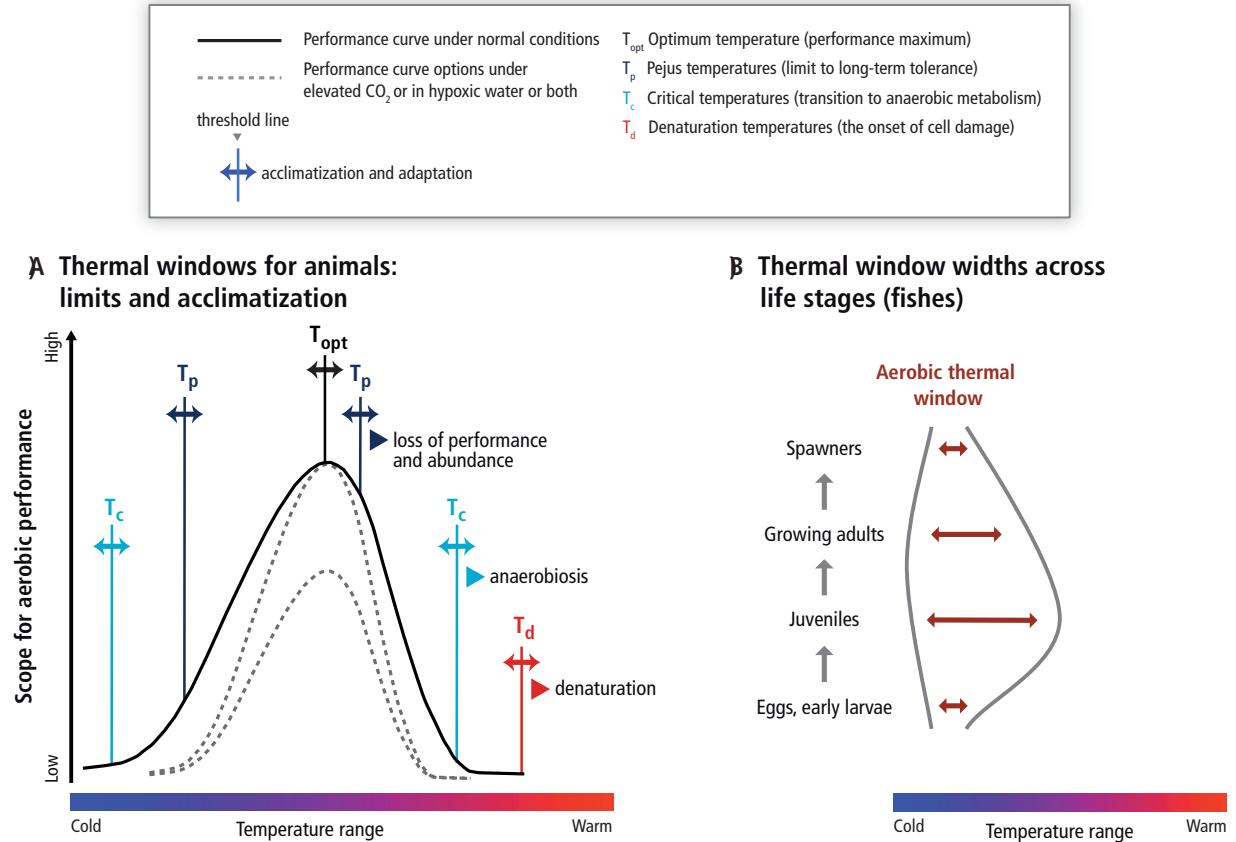


Figure 6-5: Thermal specialization of an organism explains the why, how, when, and where of climate sensitivity. (A) The thermal tolerance range and performance levels of an organism are described by its performance curve (exemplified for an animal). Each Performance (e.g., exercise, growth, reproduction) is maximal at its optimum temperature (T_{opt}), and becomes progressively constrained during cooling or warming. Surpassing the first low and high temperature thresholds (T_p, p, pejus: getting worse) means going into time-limited tolerance. Once further cooling or warming surpasses the next low or high thresholds (T_c, c, critical), oxygen availability becomes insufficient and an anaerobic metabolism begins. Denaturation temperatures (T_d) are even more extreme and characterized by the onset of damage to cells and proteins. Horizontal arrows indicate that T_p, T_c, and T_d-thresholds of an individual can shift, within limits, between summer and winter (seasonal acclimatization) or when the species adapts to a cooler or warmer climate over generations (evolutionary adaptation). Under elevated CO₂ levels (ocean acidification) and in hypoxic waters performance levels can decrease and thermal windows narrow (dashed grey curves). (B) The width of the thermal range (horizontal arrows) also changes over time when an individual develops from egg to larva to adult and gains weight and size. Blue to red colour gradients illustrate the range between cold to warm temperatures (after Pörtner, 2002a, 2012; Pörtner and Farrell, 2008).

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

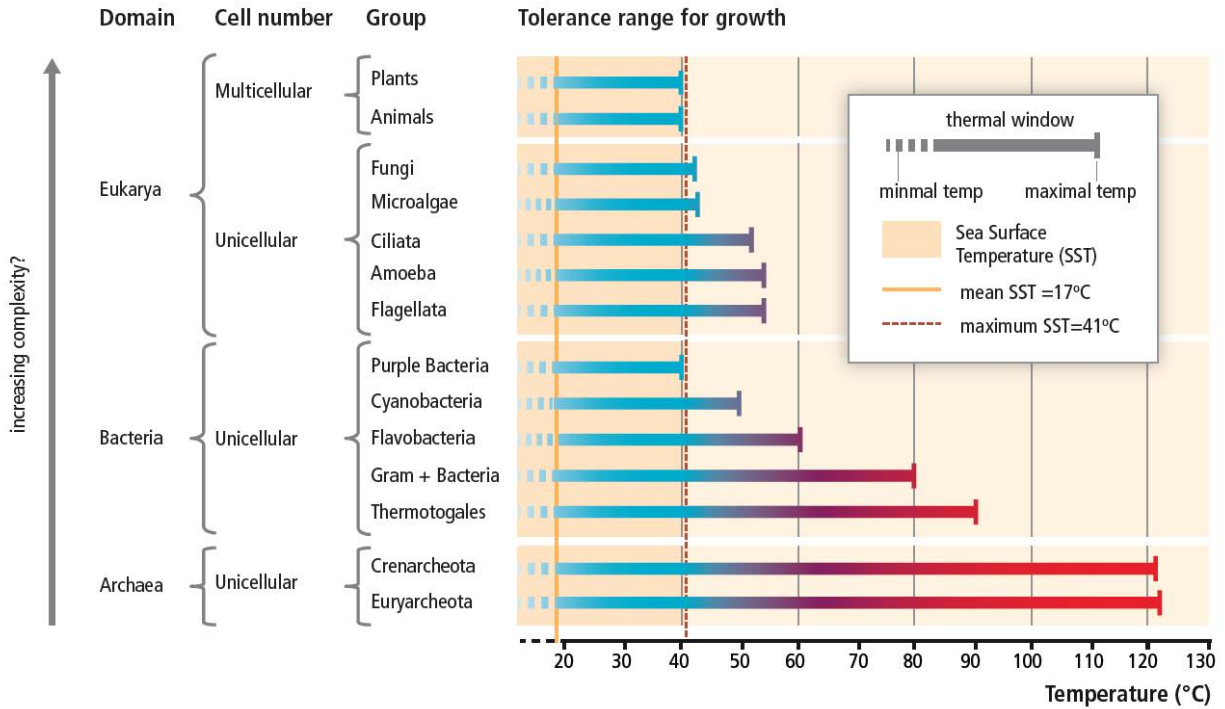


Figure 6-6: Maximal values of temperature covered by various domains and groups of free-living marine organisms (bacteria to animals, domains, and groups modified after Woese *et al.*,1990). High organizational complexity is hypothesized to be associated with decreasing tolerance to heat and to enable an increase in body size which in turn, decreases heat tolerance further (Alker *et al.*, 2001; Baumgartner *et al.*, 2002; Campbell *et al.*, 2006; Chevaldonné *et al.*, 2000; De Jonckheere *et al.*, 2009, 2011; Pörtner, 2002a,b; Sorokin and Kraus, 1962). In the domain Bacteria, the Thermotogales are less complex and most tolerant to high temperatures (Abed *et al.*, 2002; Huber *et al.*, 1986; Takai *et al.*, 1999; Tenreiro *et al.*, 1997; Ventura *et al.*, 2000). The highest temperature, at which growth can occur is 122°C for hydrothermal vent archaea, seen under elevated hydrostatic pressure in laboratory experiments (Kashefi and Lovley, 2003; Takai *et al.*, 2008.)

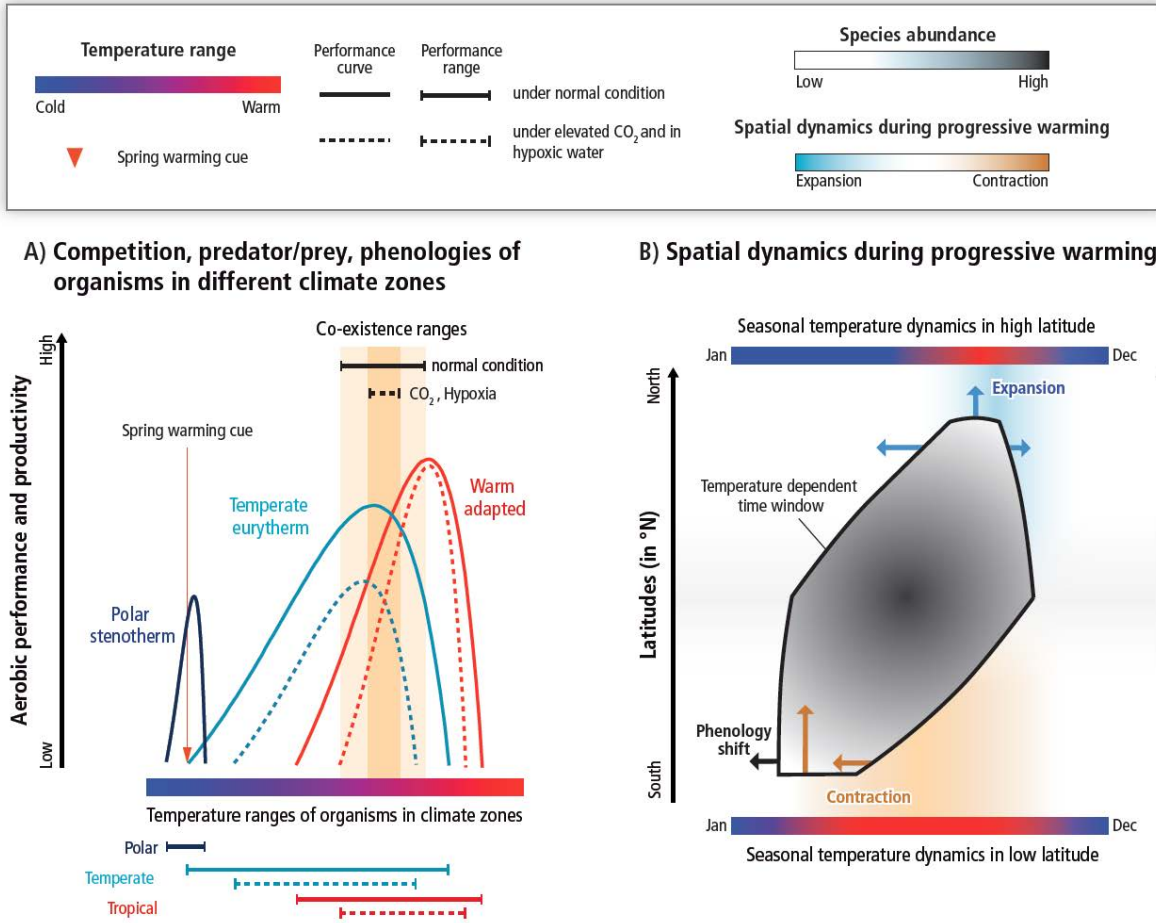
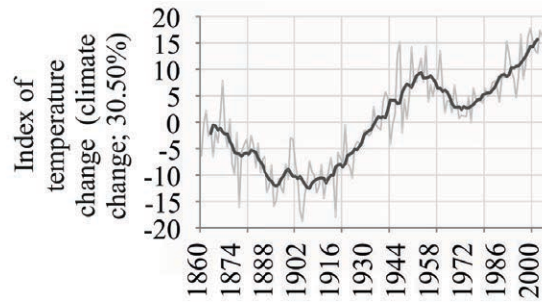


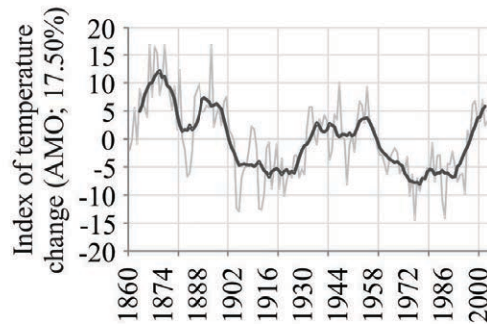
Figure 6-7: Role of thermal tolerance and performance of organisms at ecosystem level. (A) Thermal tolerance ranges (Figure 6-5) differ between species across polar, temperate and tropical climate zones, then overlapping between coexisting species. Shifting temperatures and specific effects of additional drivers on the respective performance curves (dashed lines) change the fitness of coexisting species relative to each other as well as their temperature range of coexistence (after Pörtner and Farrell, 2008). Warming alters the timing of seasonal activities (e.g., elicited by spring warming cues) earlier, or can benefit only one of two interacting species (e.g. in predator-prey dynamics or competition) causing shifts in predominance. (B) During climate warming a largely unchanged thermal range of a species causes it to follow its normal temperatures as it moves or is displaced, typically resulting in a poleward shift of the biogeographic range (exemplified for the Northern hemisphere, modified after Beaugrand, 2009). The polygon delineates the distribution range in space and seasonal time; the level of grey denotes abundance. The southern time window of tolerated temperatures shifts to earlier and contracts, while the northern one dilates (indicated by arrows). Species display maximum productivity in low latitude spring, wide seasonal coverage in the center, and a later productivity maximum in the North. The impact of photoperiod (length of daily exposure to light) increases with latitude (grey arrow). Water column characteristics or photoperiod may overrule temperature control in some organisms (e.g. diatoms), limiting northward displacement.

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

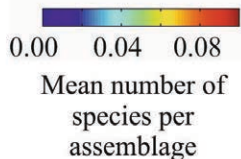
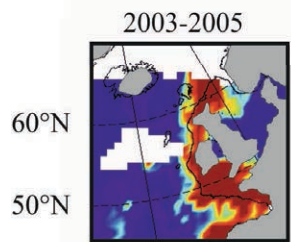
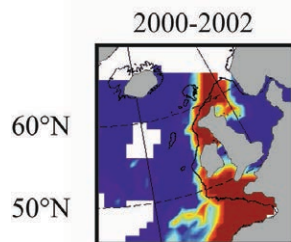
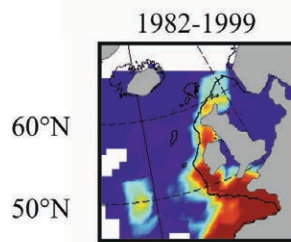
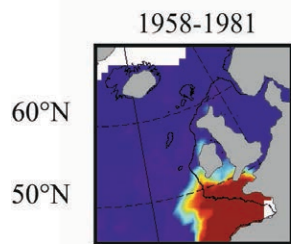
A. Climatic warming



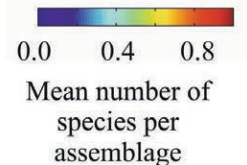
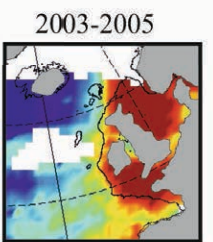
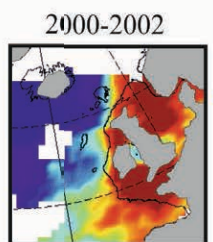
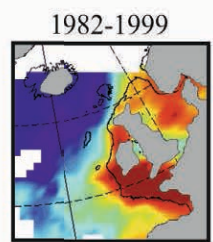
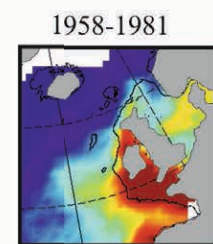
B. Atlantic Multidecadal Oscillation



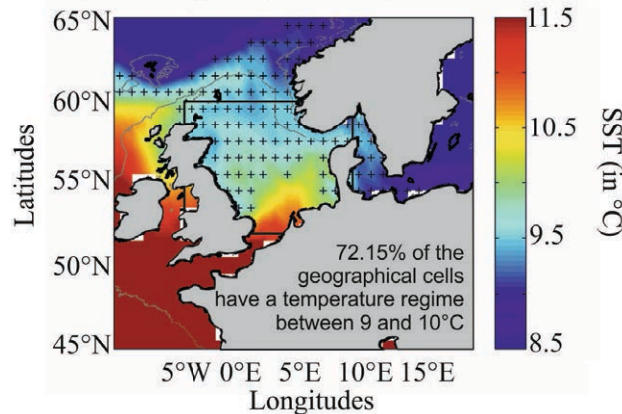
E. Warm-temperate pseudo-oceanic species



F. Temperate pseudo-oceanic species



C. Temperature regime (1960-1981)



D. Temperature regime (1988-2005)

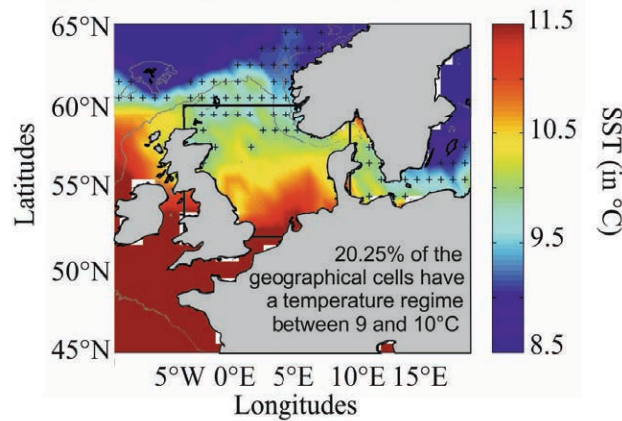


Figure 6-8: Multi-decadal changes in ecosystem structure in the NE Atlantic driven by warming from both anthropogenic climate change and natural climate variability. **A.** Index of temperature change over the North Atlantic (31°N-65°N and 99°W-11°W) reflecting climate change. This index is the first principal component (i.e., explaining 30.5% of observed variability) based on a Principal Component Analysis (PCA) performed on sea surface temperature. **B.** Index of temperature change (17.5% of observed variability) reflecting the Atlantic Multidecadal Oscillation (AMO). The index is the second principal component. **C-D.** Observed mean annual sea surface temperature in the North Sea during 1960-1981 (**C**) and 1988-2005 (**D**). The location of the critical thermal boundary (9-10°C) is indicated by '+'. **E.** Long-term changes in the mean number of warm-temperate pseudo-oceanic species from 1958 to 2005. **F.** Long-term changes in the mean number of temperate pseudo-oceanic species from 1958 to 2005. The period 1958-1981 was a period of relative stability and the period 1982-1999 was a period of rapid northward shifts, indicating that the abrupt ecosystem shift observed in the North Sea was part of a large-scale response of the zooplankton biodiversity to warming temperatures (see **A-D**). Average values are below 1 because they are annual averages. Note that the color bar is 10-fold smaller for warm-temperate pseudo-oceanic species because these species are less frequently observed than their temperate counterparts. Panels A. and B. from Edwards *et al.* (2013), and C. to F. from Beaugrand *et al.* (2008), and Beaugrand *et al.* (2009).
[Illustration to be redrawn to conform to IPCC publication specifications.]

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

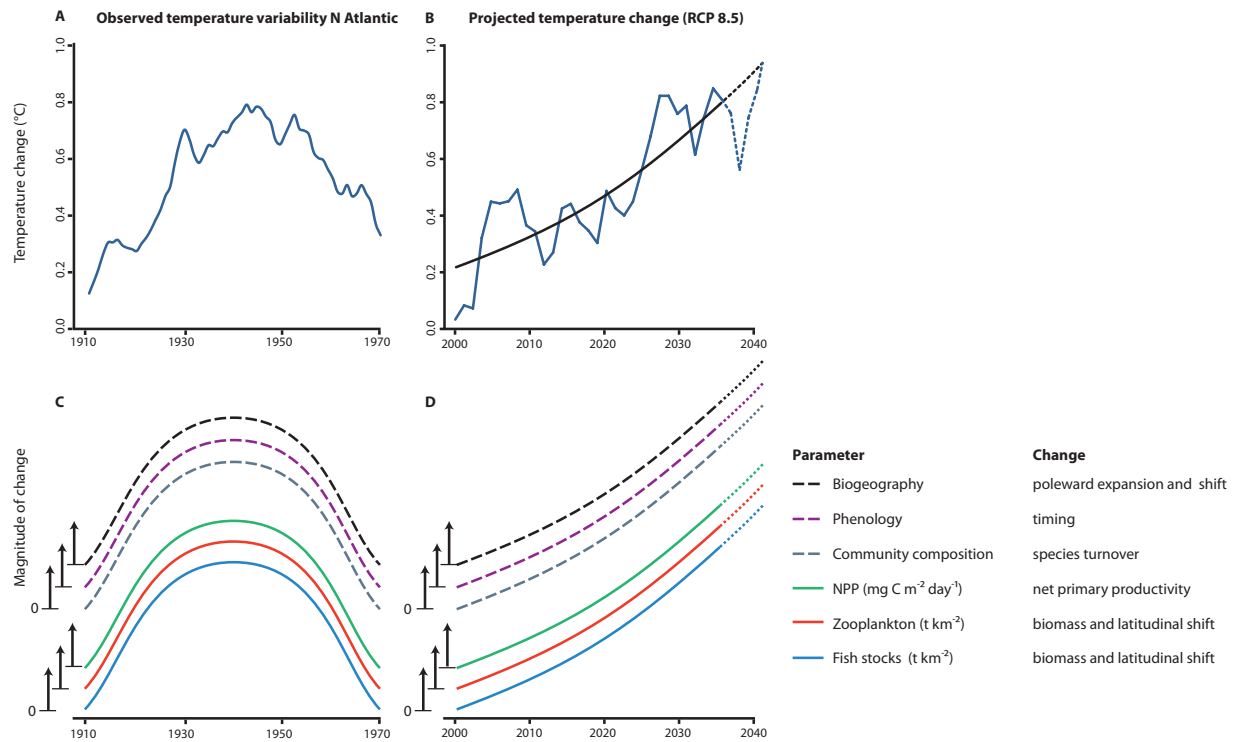
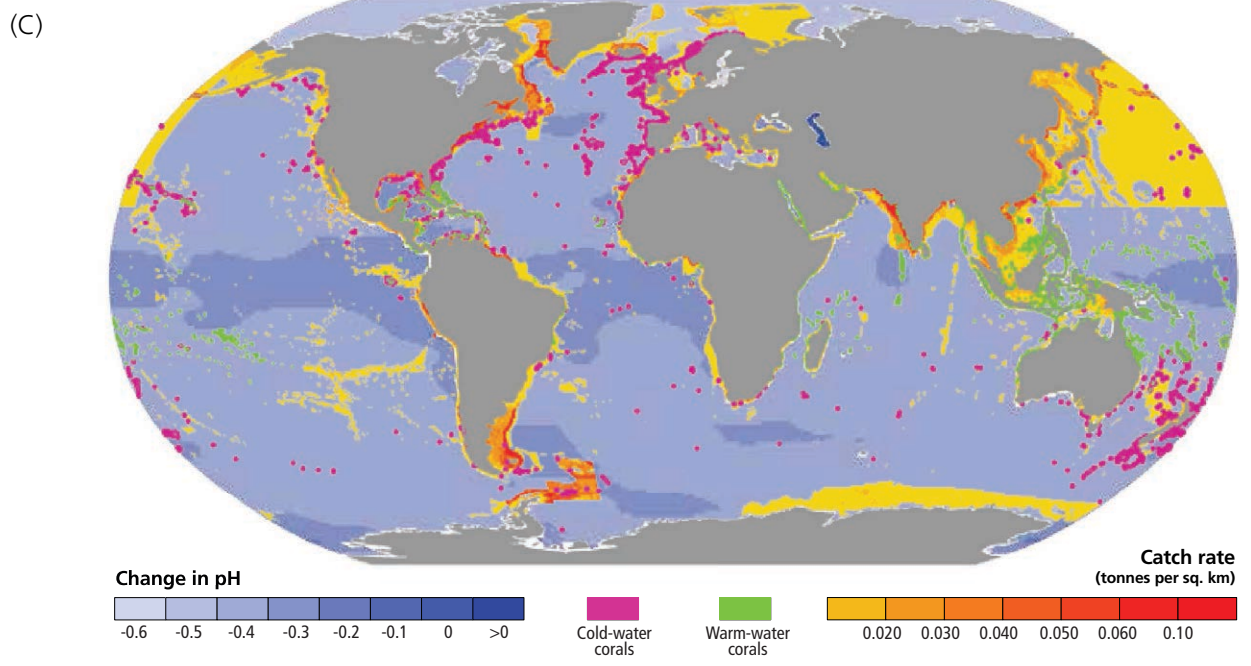
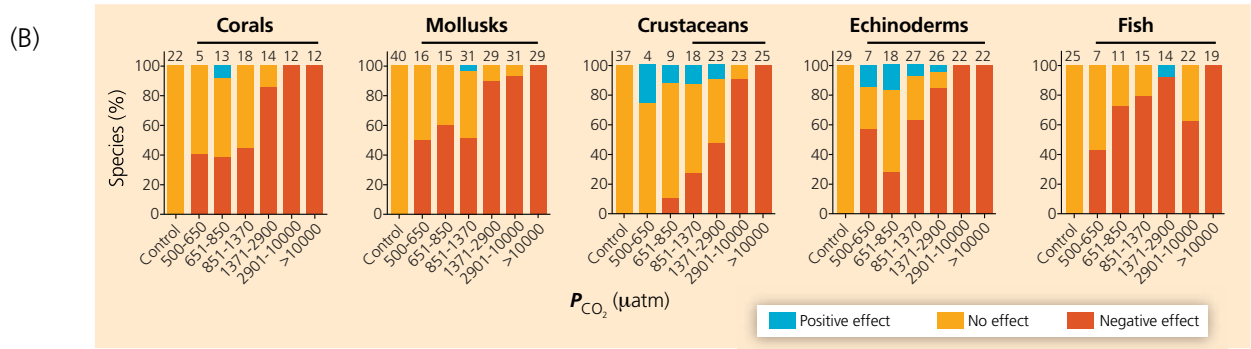
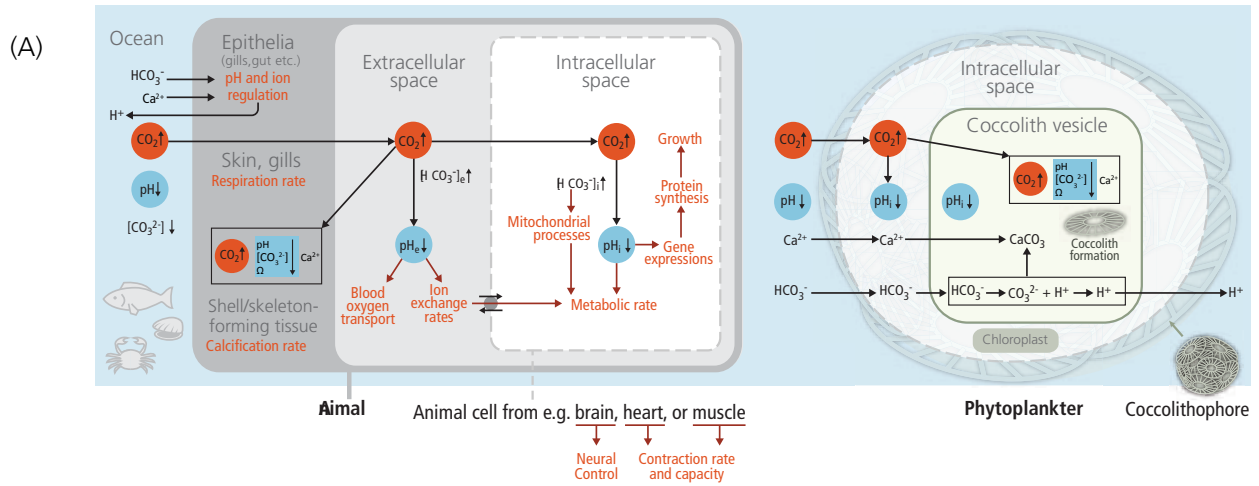


Figure 6-9: Schematic depiction of observed effects of $\sim 1^{\circ}\text{C}$ ocean warming in the northern North Atlantic driven by climate variability (A, C) versus effects expected from anthropogenic climate change (B, D). A) Transient warming and cooling associated with AMO variability (Drinkwater, 2006), based on the Kola Section temperatures (0-200 m; Stations 3-7, 71.5° - 72.5°N , 33.5°E) in the Barents Sea obtained from <http://www.pinro.ru> and filtered using a 20-year running mean. Similar trends occurred across most of the northern North Atlantic although the amplitude and timing of the peaks and troughs varied spatially. B) Warming driven by climate change for the same region (RCP 8.5 simulations averaged from CMIP5 models, computed as the mean over the upper 200 m in the grid box (2.5×2.5 degrees) centered at 71.25°N and 33.75°E). C) Warming and subsequent cooling in the northern North Atlantic during the period shown in A resulted in complex multi-faceted changes (shown schematically) in NPP (green curve), zooplankton biomass (red curve) and fish stock abundances (blue curve). There was a general poleward shift and range expansion of many commercial (e.g., Atlantic herring, Atlantic cod, haddock) and non-commercial species, reversed during the subsequent cooling period (denoted by black dashed curve). Polewards shifts in spawning areas (e.g., Atlantic cod) were also reversed as the waters cooled. The purple and grey dashed curve represents shifts in seasonal timing (phenology) and community composition, respectively, that were influenced by earlier arrivals and later retreat of migratory fish (not shown). For more details see Drinkwater (2006). D) Projected effects of climate mediated warming on northern sub-polar and polar biota based on model projections of altered NPP (Bopp *et al.*, 2013), and of the range shift of exploited fishes and invertebrates (Cheung *et al.*, 2009, 2013a). The projected trends in panel D will differ with latitude, e.g., decreased NPP at lower latitudes and no significant change to NPP in temperate waters (Bopp *et al.*, 2013). Higher NPP supported and is projected to support higher trophic levels at high latitudes (C, D, 6.3.4). Note that climate variability will be superimposed on anthropogenic warming (B, see Figure 6-1, 6-8A,B). Dotted lines indicate projected changes to continue beyond the range of historical observations.

[Illustration to be redrawn to conform to IPCC publication specifications.]

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014



Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

Figure 6-10: A: Responses of a schematized marine animal (left) and a phytoplankter (right) to ocean acidification (OA). Effects are mediated via diffusive CO₂ entry (black arrows) into body and cell compartments, resulting in a rise in pCO₂ (highlighted in red), a drop in compartmental pH (highlighted in blue) and their effects (red arrows) on various processes (red text) in tissues and cellular compartments, as well as on calcium carbonate saturation state (Ω) at calcification sites (after Pörtner, 2008; Taylor *et al.*, 2011). Variable sensitivity relates to the degree of pH decline and compensation, depending on the capacity of pH and ion regulation. B: Distribution of sensitivities across species within animal phyla, under progressively rising water CO₂ levels, as percent of studied scleractinian coral, echinoderm, molluscan, crustacean and fish species affected negatively, positively or not at all (for effects considered see text). As not all life stages, variables and pCO₂ ranges were covered in all species two assumptions partially compensate for missing data: 1) Negative effects at low pCO₂ will remain negative at high pCO₂. 2) A positive or neutral outcome at both low and high pCO₂ will be the same at intermediate pCO₂. As responses reported for each species vary for each CO₂ range, variable species numbers result (on top of columns). The total number of species studied in a group is shown as the number above the control column. Horizontal lines above columns represent frequency distributions significantly different from controls (Wittmann and Pörtner, 2013). C: Areas with reported annual catches of marine calcifiers (crustaceans and mollusks) >0.001 t km⁻² depicted on a global map showing the distribution of ocean acidification in 2100 according to RCP8.5 (WGI AR5 SPM) as well as the distribution of warm-water (green dots) and cold-water coral communities (pink dots).

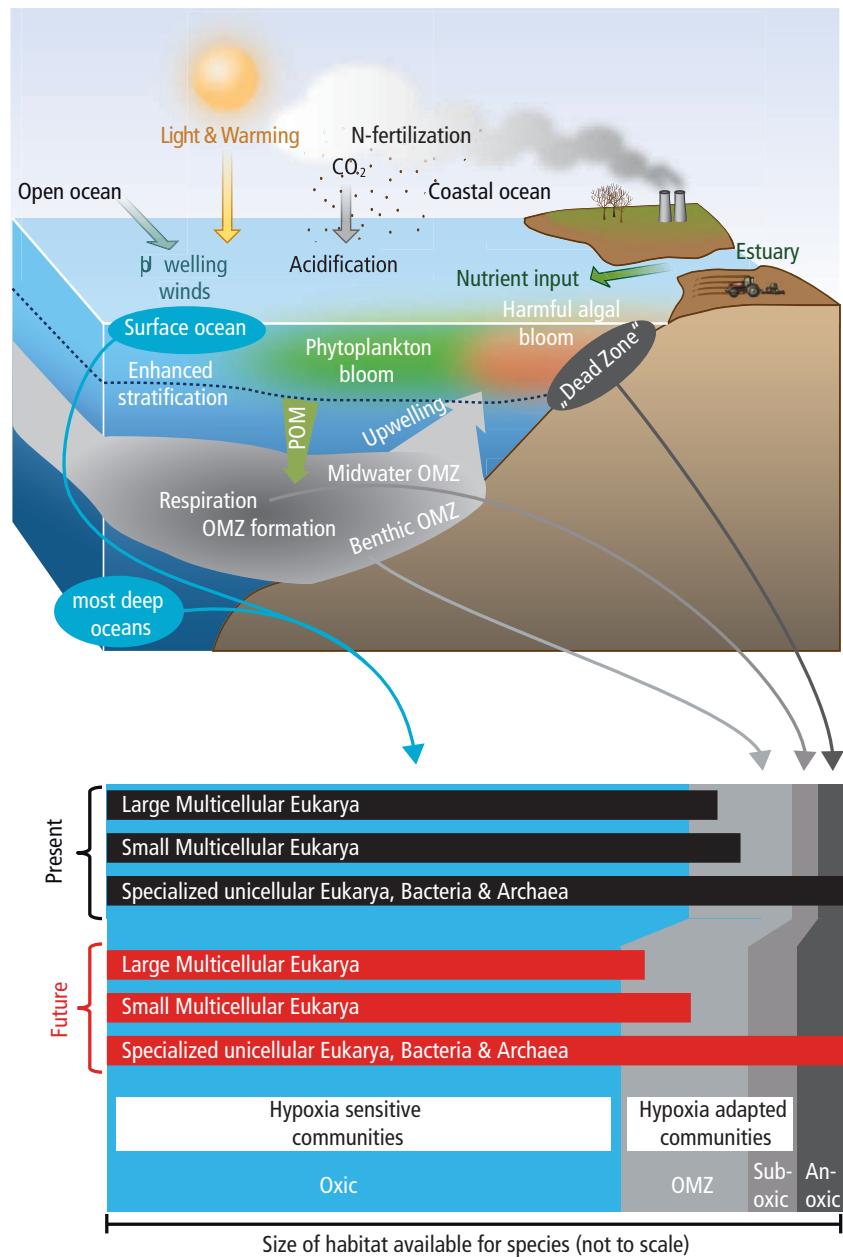


Figure 6-11: (A) Principal mechanisms underlying the formation of hypoxic conditions and their biological background (modified from Levin *et al.*, 2009; Levin and Sibuet, 2012). The buoyancy flux from fluvial discharges produces sharp density stratification at the base of the freshened layer (also valid for ice melt and high precipitation) near the surface and, hence, vertical mixing is greatly reduced. In consequence, the nutrient inputs from the river and the atmosphere accumulate in a narrow upper layer, leading to phytoplankton blooms. The enhancement of oxygen consumption due to aerobic decomposition of sinking organic matter results in hypoxic conditions of OMZs. OMZs also form in the open oceans. Heating of the upper layer increases stratification, while the wind-driven upwelling of hypoxic, nutrient-rich water from deeper layers adds to the formation of the OMZs (Box CC-UP) (B) Distribution of various free-living marine organisms (microbes such as archaea, bacteria, protists, small and large multicellular animals, and plants) across the ranges of O₂ concentrations in various water layers. Hypoxia tolerance is enhanced in small compared to large organisms, allowing unicellular species and small animals to thrive in extremely hypoxic habitats. Species richness and body size of animals increases with rising O₂ levels. **[Illustration to be redrawn to conform to IPCC publication specifications.]**

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

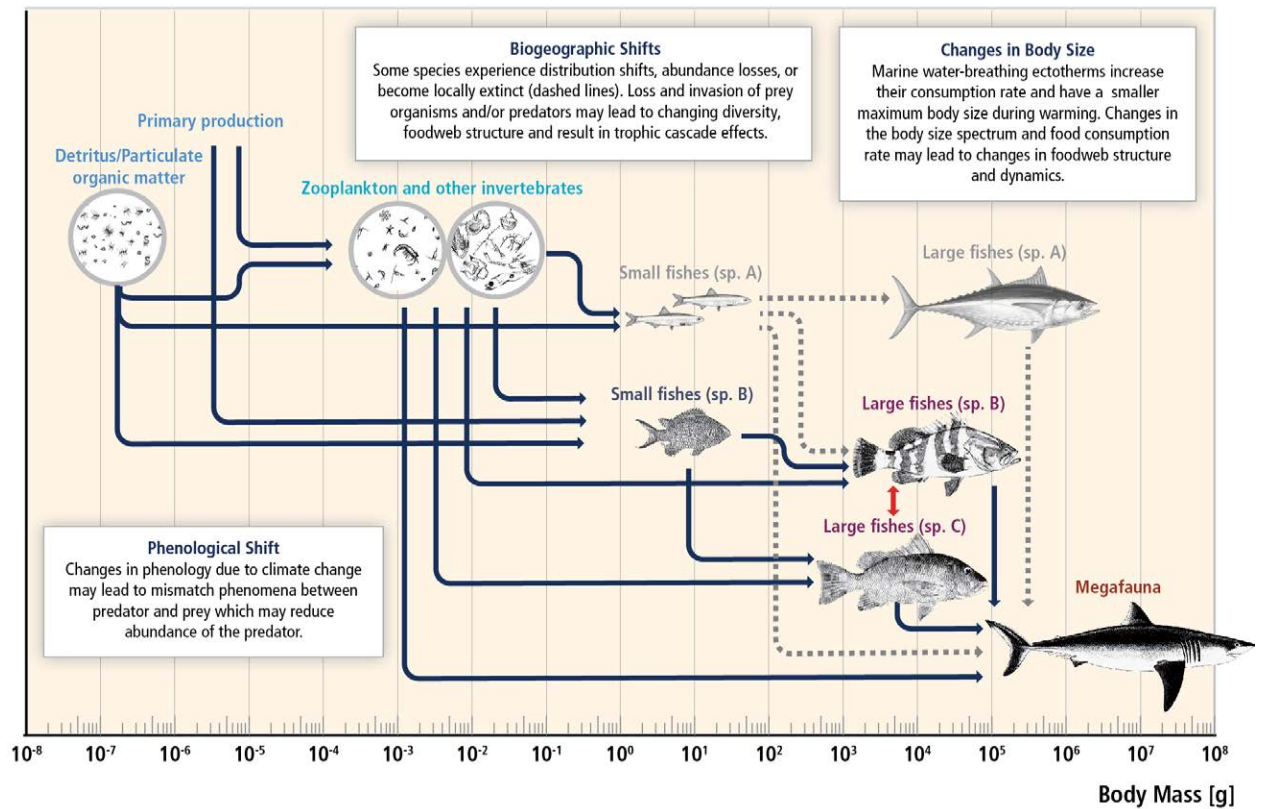


Figure 6-12: Schematic diagram of expected responses to climate change in a marine food web. A coupled pelagic and benthic food web is structured by the body size spectrum of species. Combined warming, hypoxia and ocean acidification reduce body size, shift biogeographies, change species composition and abundance, and reconfigure trophic linkages and interaction dynamics. Fishing generally removes large-bodied species and truncates the body-size spectrum of the community. This confounds the detection and attribution of food web responses to climate change. Arrows represent species interactions (e.g., between predator and prey or competitors for food or space). Broken lines reflect the potential loss of populations and trophic linkages due to climate change.

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

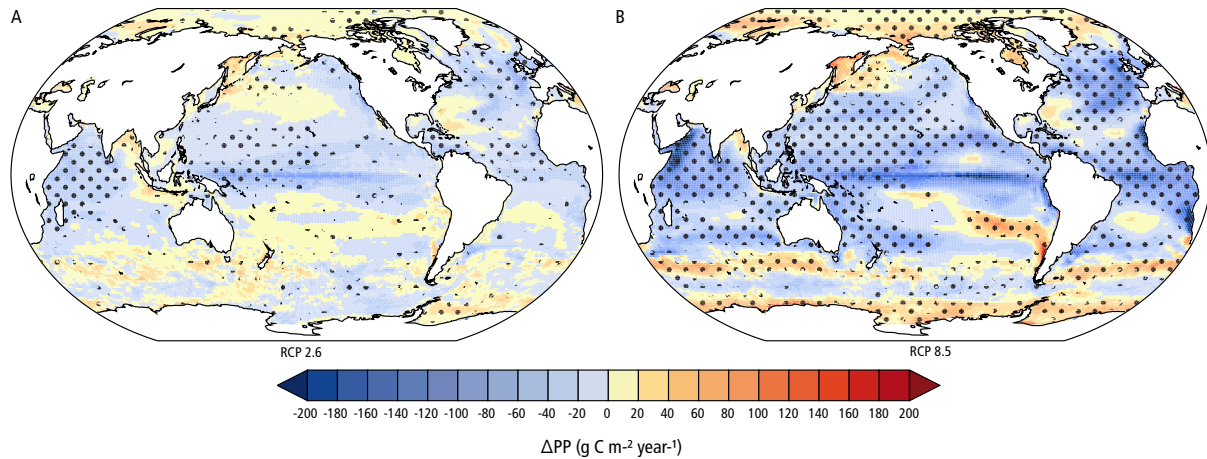
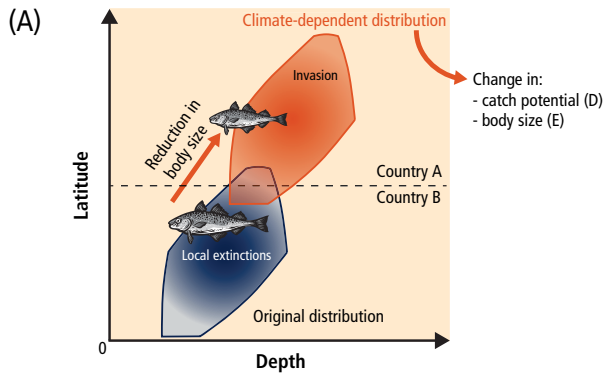
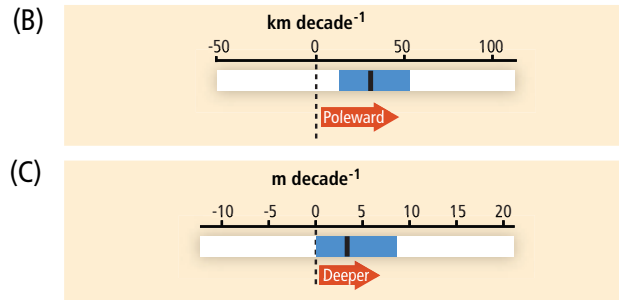


Figure 6-13: Multi-model annual mean changes of projected vertically-integrated net primary production (small and large phytoplankton) under the low emission scenario RCP2.6 (A) and the high emission scenario RCP8.5 (B) for the period 2090 to 2099 relative to 1990 to 1999 (after Bopp *et al.*, 2013). To indicate consistency in the sign of change, regions are stippled where 80% of the ten models from the Coupled Model Intercomparison Project Phase 5 (Bopp *et al.* 2013) agree on the sign of change.

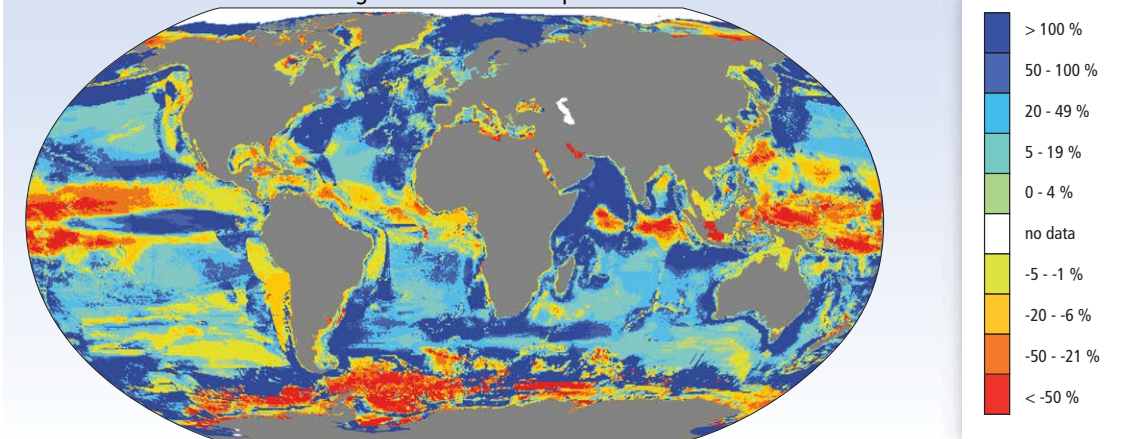
[Illustration to be redrawn to conform to IPCC publication specifications.]



(B) Shifting distribution to cooler water



(D) Change in maximum catch potential



(E) Change in maximum body weight

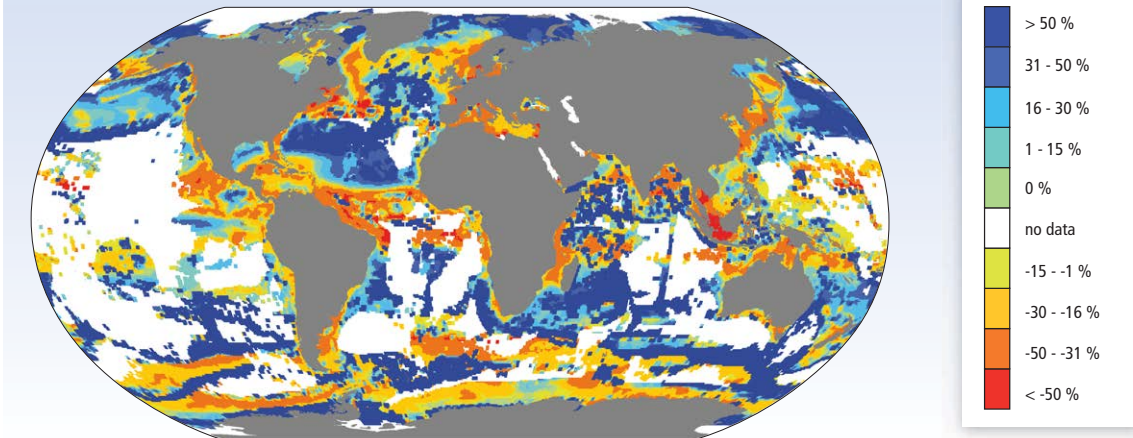


Figure 6-14: Climate change effects on the biogeography, body size and fisheries catch potential of marine fishes and invertebrates. (A) Shifts in distribution range and reduction in body size of exploited fish driven by projected warming, oxygen depletion and sea-ice retreat (cf. Figure 6-7). Whenever the shift in distribution does not fully compensate for warming and hypoxia, the result will be a decrease in body size. Shifts in (B) latitudinal and (C) depth distribution of 610 exploited demersal fishes are projected to have a median (central line of the box) of 31 km decade⁻¹ and 3.3 m decade⁻¹, respectively, with variation between species (box boundary: 25th and 75th percentiles) from 1991-2010 to 2041 - 2060 under the SRES A2 (between RCP6.0 - 8.5) scenario (Cheung *et al.*, 2011; Cheung *et al.*, 2013b). (D) Combining species' range shift with projected changes in NPP leads to a projected global redistribution of maximum catch potential (Analysis includes ~1,000 species of exploited fishes and invertebrates, under warming by 2°C according to SRES A1B (~RCP6.0), comparing the 10-year averages 2001-2010 and 2051-2060; redrawn from Cheung *et al.* 2010). (E) Changes in species distribution and individual growth are projected to lead to reduced maximum body size of fish communities at a certain site. The analysis includes 610 species of marine fishes, from 1991-2010 to 2041-2060 under the SRES A2 (~RCP6.0 to 8.5, Cheung *et al.*, 2013b). Key assumptions of the projections are that current distribution ranges reflect the preferences and tolerances of species for temperature and other environmental conditions and that these preferences and tolerances do not change over time. Catch potential is determined by species range and net primary production. Growth and maximum body size of fishes are a function of temperature and ambient oxygen level.

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

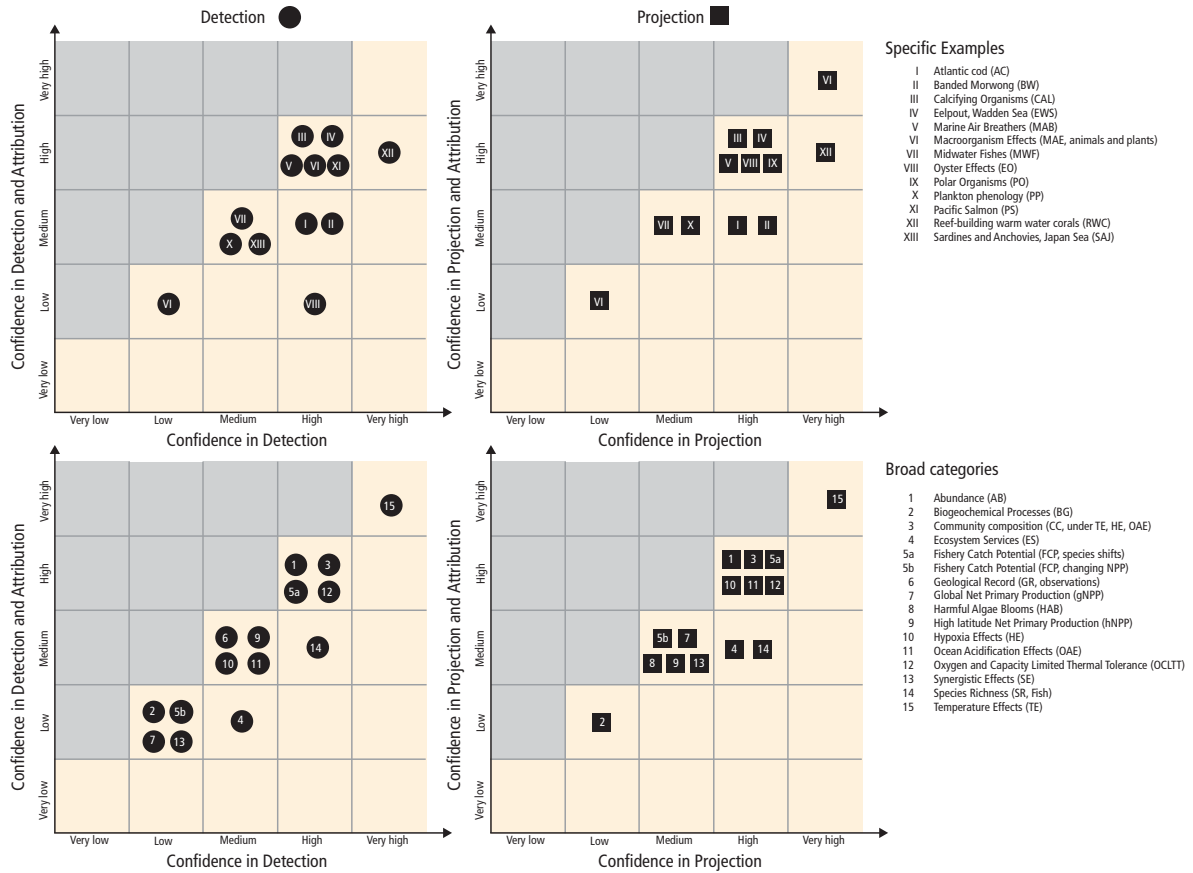


Figure 6-15: Overview of the levels of confidence in detection (left), as well as in projection (right) of climate change effects on ocean systems, in relation to the levels of confidence in attributing these effects to the respective climate forcings. Case studies, processes, and concepts relevant in assessing the effects of climate change are represented by their acronyms in both text and figure. While confidence in the presence of effects is often high, the direct attribution to one driver in field experiments is difficult, as drivers are often highly correlated with each other (e.g., warming with changes in stratification and hence reduced nutrient supply). Some climate change impacts have been condensed into broad categories to avoid overpopulating the figures (e.g., Bio-Geochemical processes, BG). Note that the term “attribution” is used for both present-day detections in the field and future projections, the latter including qualitative and quantitative extrapolations and simulations from fundamental principles and models. Firm knowledge from experiments (field, laboratory and modeling) simulating future conditions enhances the respective confidence levels to those for detection or projection. The empirical observations resulting from those experiments are directly attributable to the respective drivers. Confidence in attribution is enhanced if these experiments identify the underlying mechanisms and their responses. See text for the discussion of depicted examples and categories. Confidence assignments focus on the nature and size of effects, not on model capacity to reliably quantify their magnitude.

[Illustration to be redrawn to conform to IPCC publication specifications.]

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

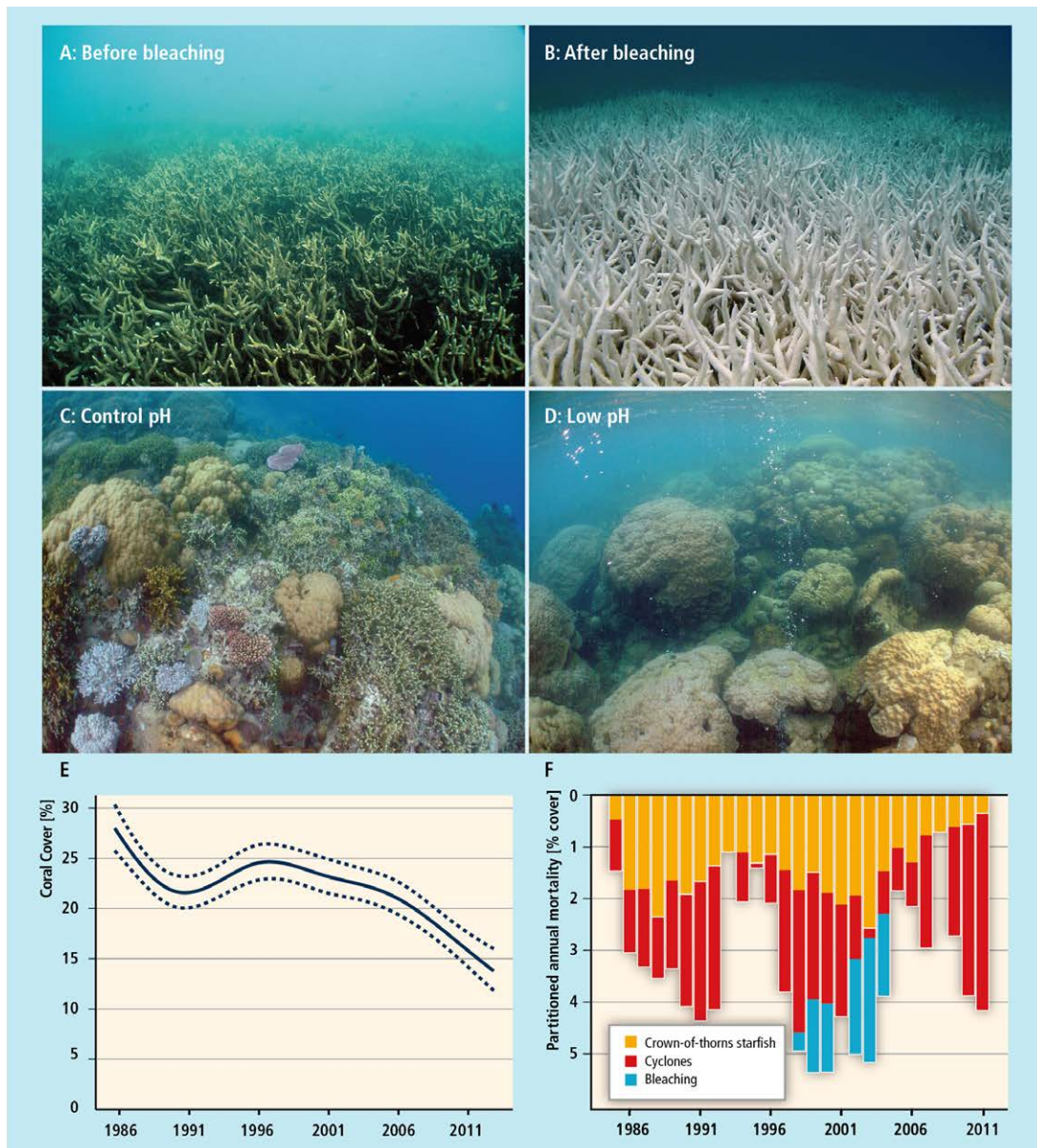


Figure CR-1: A and B: the same coral community before and after a bleaching event in February 2002 at 5 m depth, Halfway Island, Great Barrier Reef. Coral cover at the time of bleaching was 95% bleached almost all of it severely bleached, resulting in mortality of 20.9% (Elvidge *et al.*, 2004). Mortality was comparatively low due in part because these coral communities were able to shuffle their symbiont to more thermo-tolerant types (Berkelmans and van Oppen, 2006; Jones *et al.*, 2008). C and D: three CO₂ seeps in Milne Bay Province, Papua New Guinea show that prolonged exposure to high CO₂ is related to fundamental changes in the ecology of coral reefs (Fabricius *et al.*, 2011), including reduced coral diversity (-39%), severely reduced structural complexity (-67%), lower density of young corals (-66%) and fewer crustose coralline algae (-85%). At high CO₂ sites (panel D; median pH_T ~7.8), reefs are dominated by massive corals while corals with high morphological complexity are underrepresented compared with control sites (D; median pH ~8.0). Reef development ceases at pH_T values below 7.7. pH_T: pH on the total scale. E: temporal trend in coral cover for the whole Great Barrier Reef over the period 1985–2012 (N, number of reefs, mean ± 2 standard errors; De'ath *et al.*, 2012). F: composite bars indicate the estimated mean coral mortality for each year, and the sub-bars indicate the relative mortality due to crown-of-thorns starfish, cyclones, and bleaching for the whole Great Barrier Reef (De'ath *et al.*, 2012). Photo credit: R. Berkelmans (A and B) and K. Fabricius (C and D).

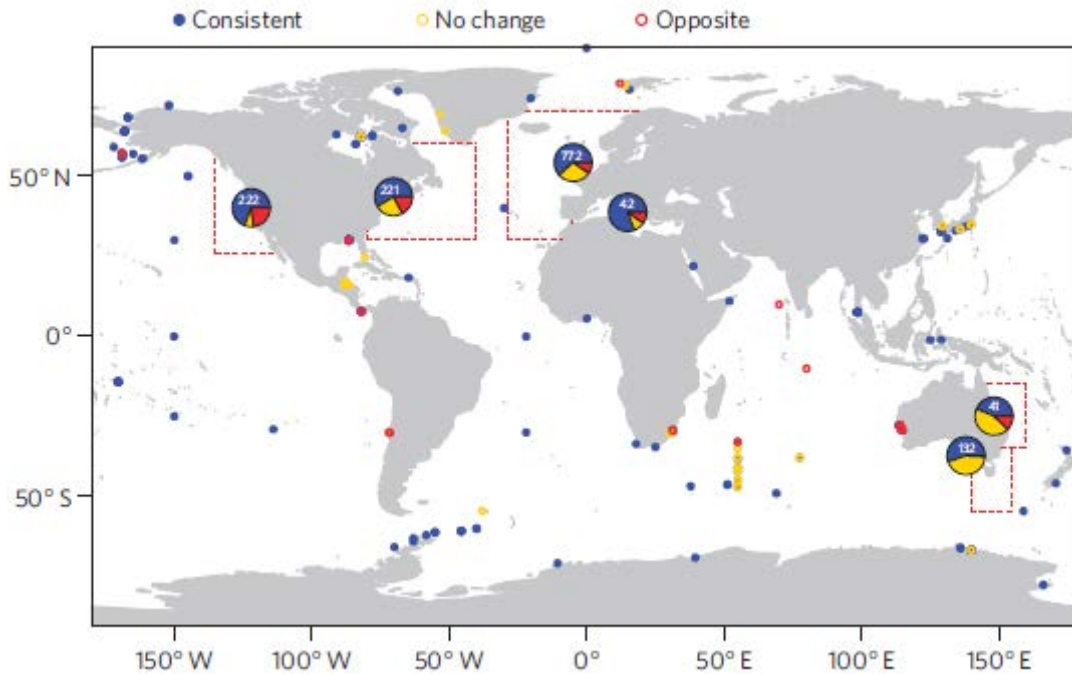


Figure MB-1: 1735 observed responses to climate change from 208 single- and multi-species studies. Changes attributed to climate change (blue), inconsistent with climate change (red) and are equivocal (yellow). Each circle represents the centre of a study area. Where points fall on land, it is because they are centroids of distribution that surround an island or peninsula. Pie charts show the proportions within regions bounded by red squares and in the Mediterranean; numbers indicate the total (consistent, opposite or equivocal) observations within each region. Note: 57% of the studies included were published since AR4 (from Poloczanska *et al.*, 2013).

[Illustration to be redrawn to conform to IPCC publication specifications.]

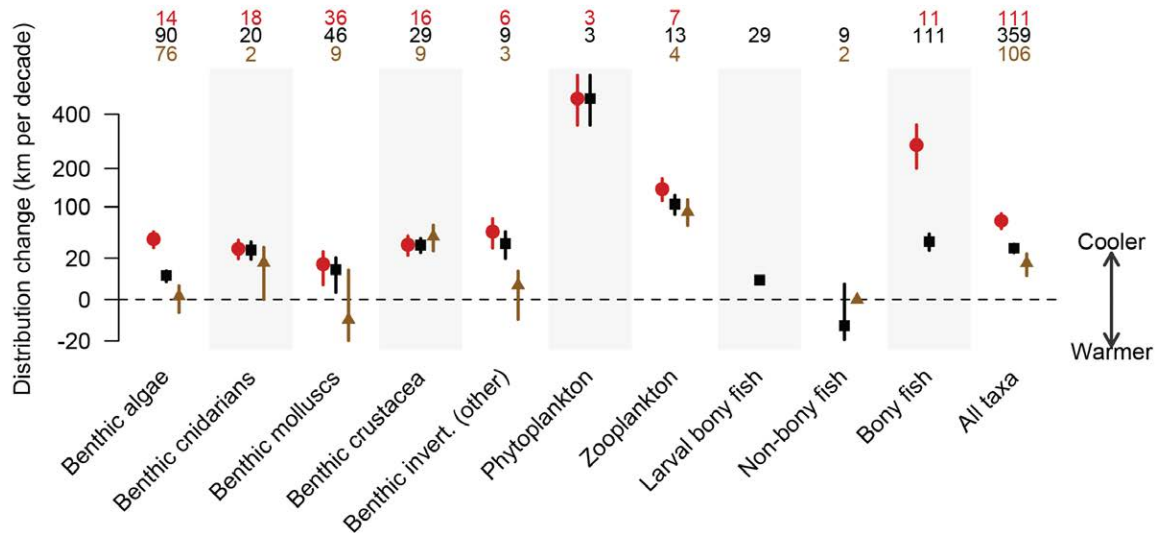


Figure MB-2. Rates of change in distribution (km decade^{-1}) for marine taxonomic groups, measured at the leading edges (red) and trailing edges (brown). Average distribution shifts calculated using all data, regardless of range location, are in black. Distribution rates have been square-root transformed; standard errors may be asymmetric as a result. Positive distribution changes are consistent with warming (into previously cooler waters, generally poleward). Means \pm standard error are shown, along with number of observations (from Poloczanska *et al.*, 2013).

[Illustration to be redrawn to conform to IPCC publication specifications.]

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

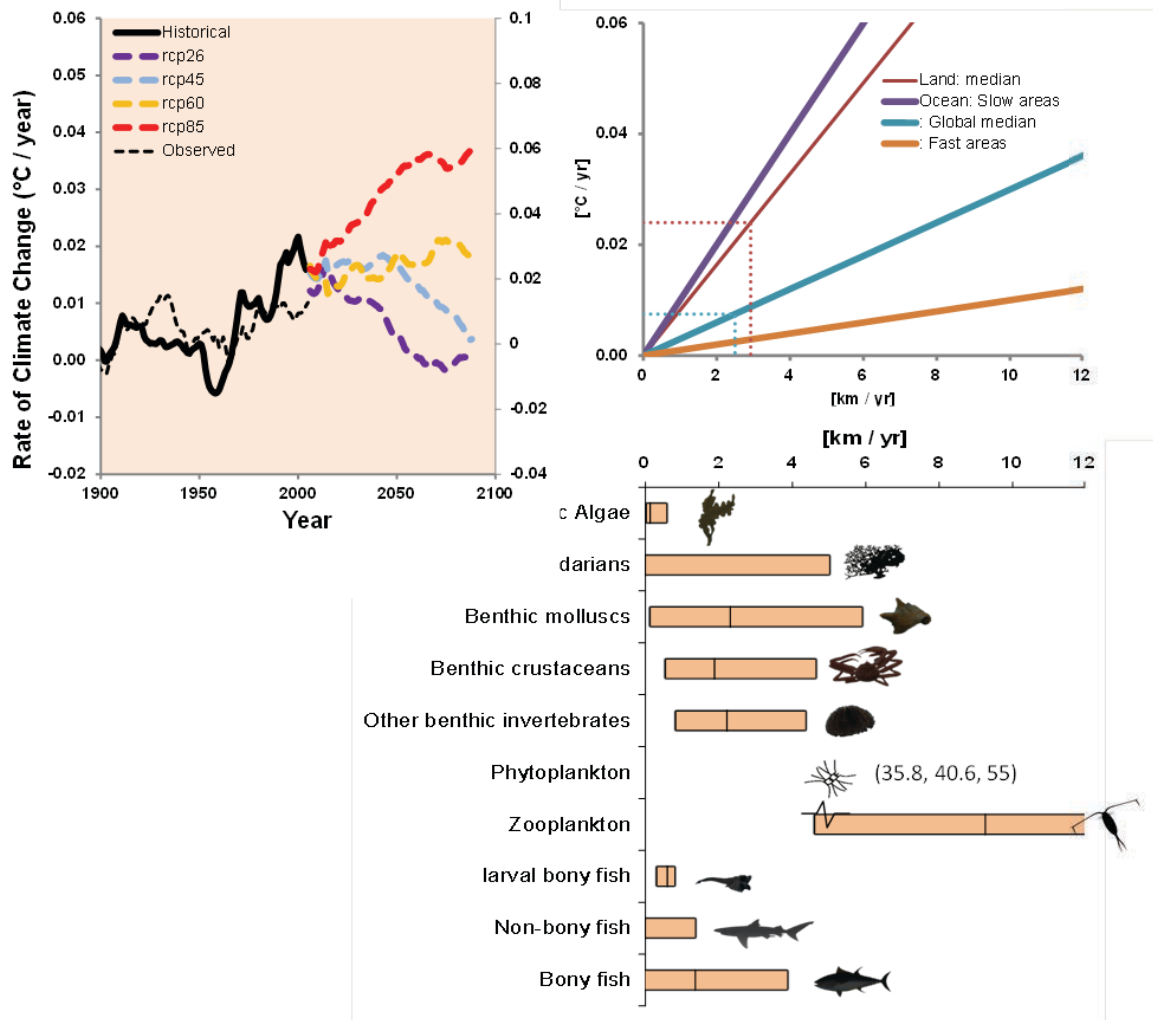
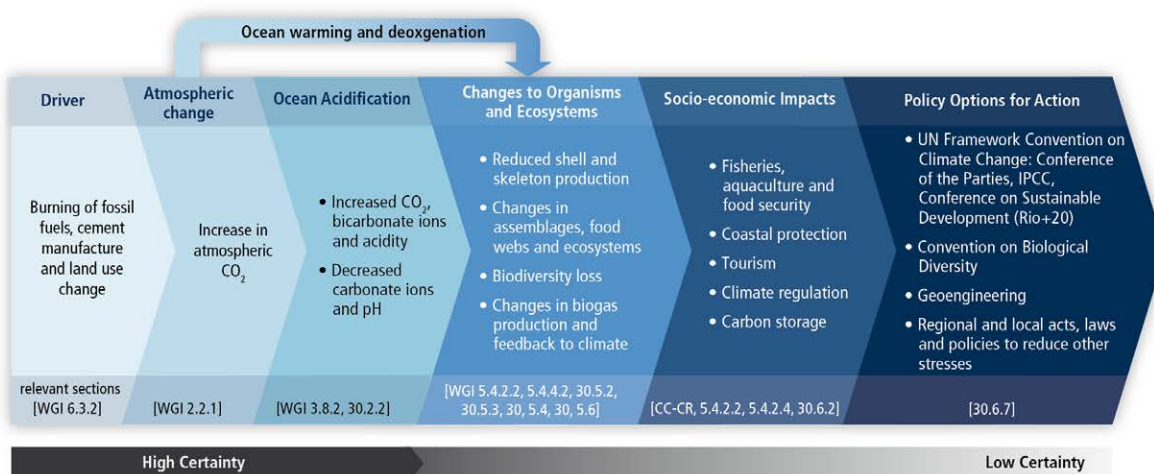
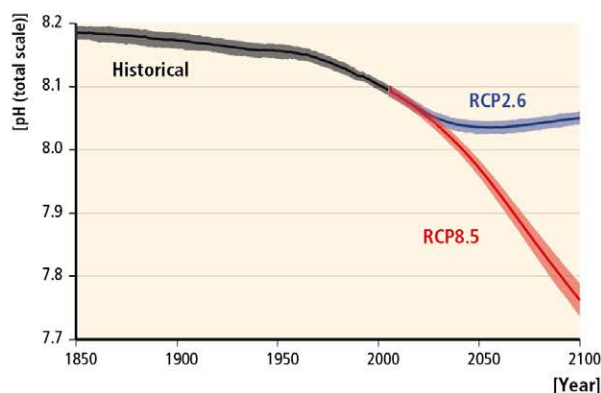


Figure MB-3. A. Rate of climate change for the Ocean (sea surface temperature (SST) °C); B. corresponding climate velocities for the Ocean and median velocity from land (adapted from Burrows et al., 2011); and C. observed rates of displacement of marine taxonomic groups over several decades until 2010. The thin dotted red arrows give an example of interpretation. Rates of climate change of $0.008 \text{ } ^\circ\text{C yr}^{-1}$ correspond to ca. 2.4 km yr^{-1} median climate velocity in the Ocean. When compared to observed rates of displacement, many marine taxonomic groups have been able to track these velocities, except phyto- and zooplankton where rates of displacement greatly exceed climate velocity. All values are calculated for ocean surface with the exclusion of polar seas (Figure 30-1a). (A) Observed rates of climate change for Ocean SST (Black dotted line) are derived from HadISST1.1 data set, all other rates are calculated based on the average of the CMIP5 climate model ensembles (Table S30-3) for the historical period and for the future based on the four RCP emissions scenarios. Data were smoothed using a 20-year sliding window. (B) Median climate velocity calculated from HadISST1.1 dataset over 1960–2010 using the methods of Burrows et al., 2011. The three axes represent estimated median climate velocities are representative of areas of slow velocities such as Pacific subtropical gyre (STG) system (Purple line), the global Ocean surface (excluding polar seas, Blue line), and areas of high velocities such as the Coral Triangle and North Sea (Orange line). Figure 30-3 shows climate velocities over the ocean surface calculated over 1960–2010. The Red line corresponds to the median rate over global land surface calculated using historical surface temperatures from the CMIP5 model ensemble (Table S30-3). (C) Rates of displacement for marine taxonomic groups estimated by Poloczanska et al. 2013 using published studies (Figure MB-2 Black data set). Note the displacement rates for phytoplankton exceed the axis, so values are given. **[Illustration to be redrawn to conform to IPCC publication specifications.]**

A.



B.



C.

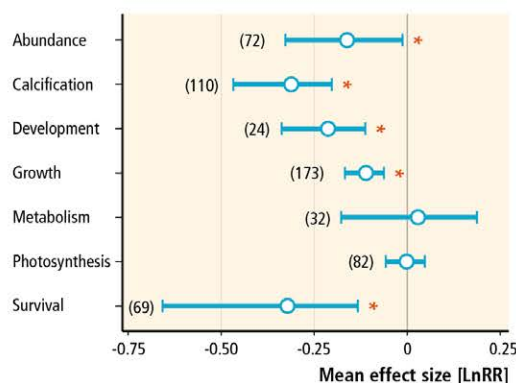


Figure OA-1: A: Overview of the chemical, biological, socio-economic impacts of ocean acidification and of policy options (adapted from Turley and Gattuso, 2012). B: Multi-model simulated time series of global mean ocean surface pH (on the total scale) from CMIP5 climate model simulations from 1850 to 2100. Projections are shown for emission scenarios RCP2.6 (blue) and RCP8.5 (red) for the multi-model mean (solid lines) and range across the distribution of individual model simulations (shading). Black (grey shading) is the modelled historical evolution using historical reconstructed forcings. The models that are included are those from CMIP5 that simulate the global carbon cycle while being driven by prescribed atmospheric CO₂ concentrations. The number of CMIP5 models to calculate the multi-model mean is indicated for each time period/scenario (WGI AR5 Figure 6.28). C: Effect of near future acidification (seawater pH reduction of 0.5 unit or less) on major response variables estimated using weighted random effects meta-analyses, with the exception of survival which is not weighted (Kroeker et al., 2013). The log-transformed response ratio (LnRR) is the ratio of the mean effect in the acidification treatment to the mean effect in a control group. It indicates which process is most uniformly affected by ocean acidification but large variability exists between species. Significance is determined when the 95% bootstrapped confidence interval does not cross zero. The number of experiments used in the analyses is shown in parentheses. * denotes a statistically significant effect.

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

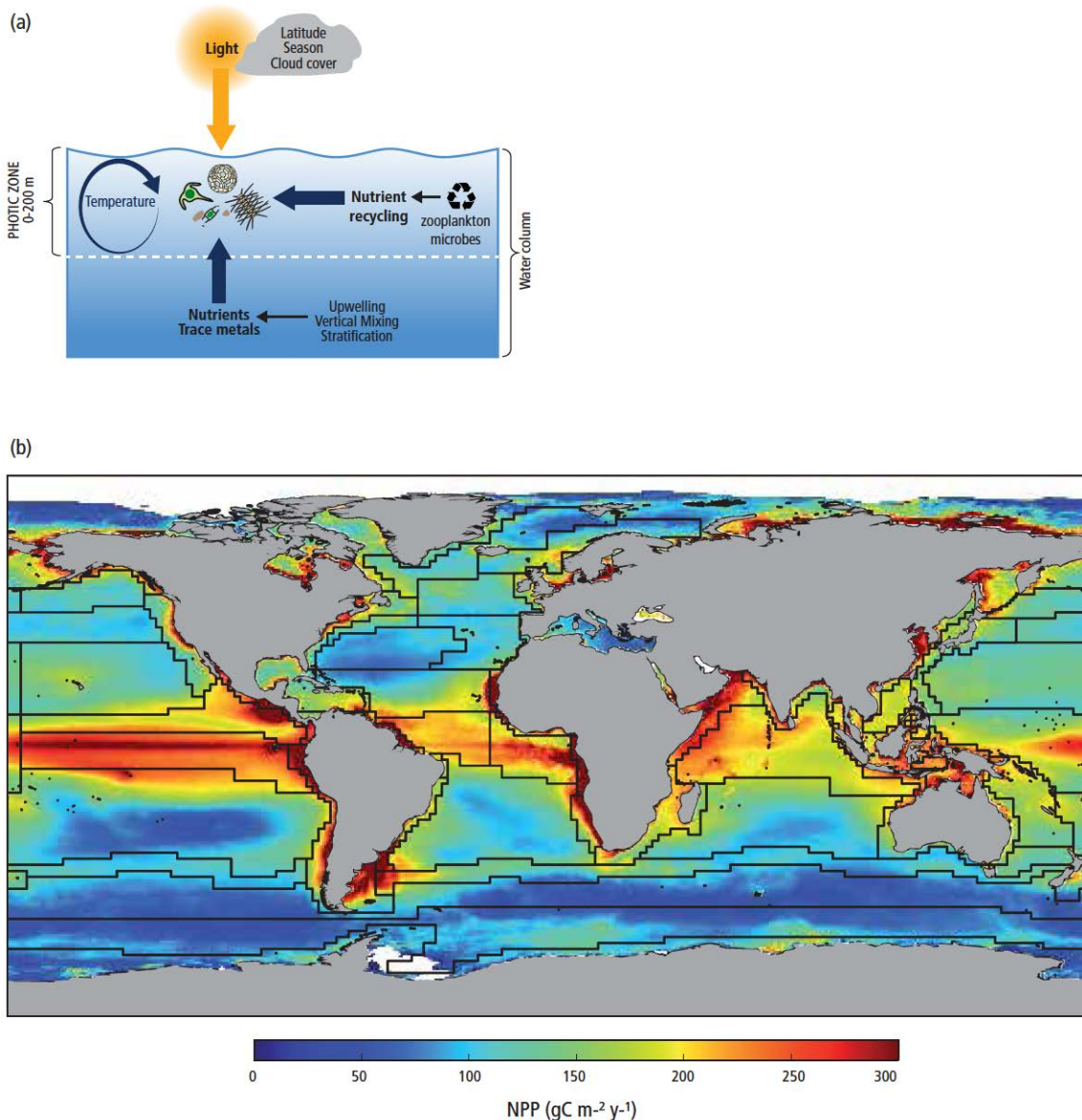


Figure PP-1: A) Environmental factors controlling Net Primary Production (NPP). NPP is mainly controlled by three basic processes: 1) Light conditions in the surface ocean, i.e. the photic zone where photosynthesis occurs, 2) upward flux of nutrients and micronutrients from underlying waters into the photic zone, 3) Regeneration of nutrients and micronutrients via the breakdown and recycling of organic material before it sinks out of the photic zone. All three processes are influenced by physical, chemical and biological processes and vary across regional ecosystems. In addition, water temperature strongly influences the upper rate of photosynthesis for cells that are resource-replete. Predictions of alteration of primary productivity under climate change depend on correct parameterizations and simulations of each of these variables and processes for each region. B) Annual composite map of global areal NPP rates (derived from MODIS Aqua satellite climatology from 2003-2012; NPP was calculated with the Carbon-based Production Model (CbPM, Westberry *et al.*, 2008)). Overlaid is a grid of (thin black lines) that represent 51 distinct global ocean biogeographical provinces (after Longhurst, 1998 and based on Boyd and Doney, 2002). The characteristics and boundaries of each province are primarily set by the underlying regional ocean physics and chemistry. Figure courtesy of Toby Westberry (OSU) and Ivan Lima (WHOI), satellite data courtesy of NASA Ocean Biology Processing Group.

[Illustration to be redrawn to conform to IPCC publication specifications.]

Do Not Cite, Quote, or Distribute Prior to Public Release on 31 March 2014

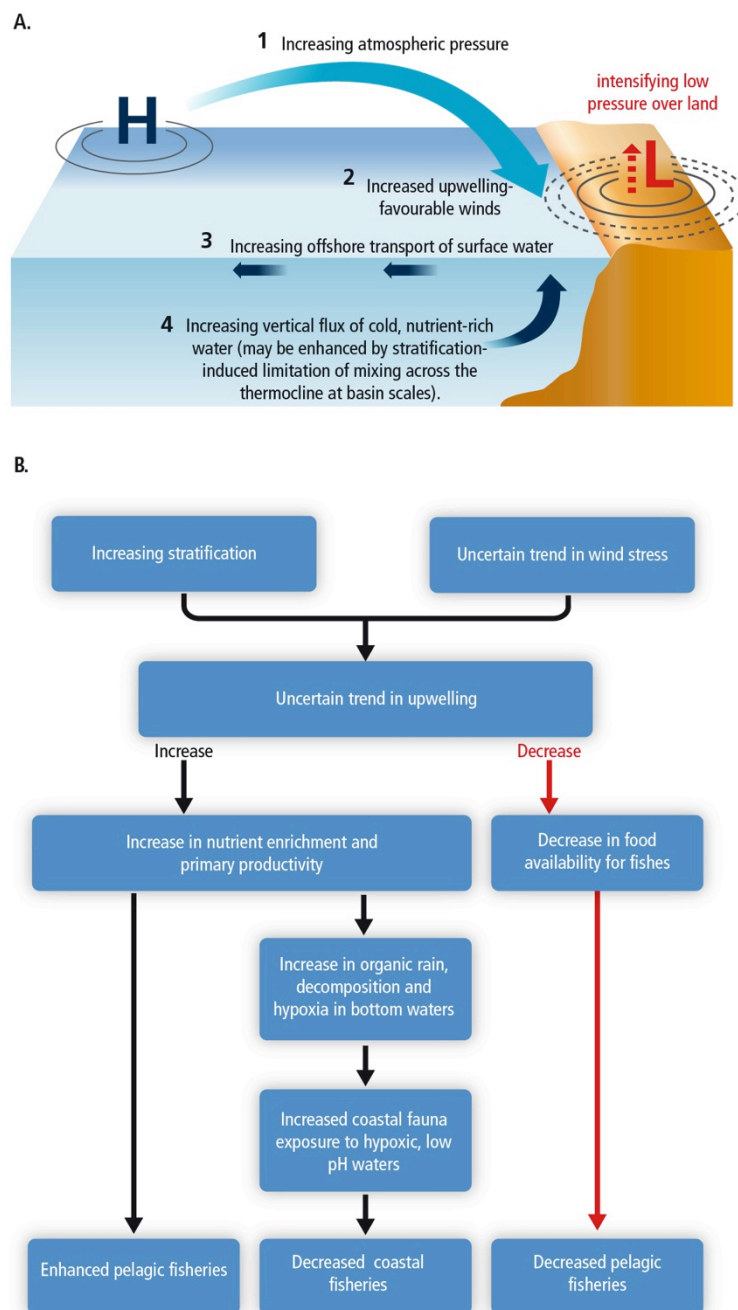


Figure UP-1: Upper panel: Schematic hypothetical mechanism of increasing coastal wind-driven upwelling at eastern boundary systems, where differential warming rates between land and ocean results in increased land-ocean pressure gradients (1) that produce stronger alongshore winds (2) and offshore movement of surface water through Ekman transport (3), and increased upwelling of deep cold nutrient rich waters to replace it (4). Lower panel: potential consequences of climate change in upwelling systems. Increasing stratification and uncertainty in wind stress trends result in uncertain trends in upwelling. Increasing upwelling may result in higher input of nutrients to the euphotic zone, and increased primary production, which in turn may enhance pelagic fisheries, but also decreased coastal fisheries due to an augmented exposure of coastal fauna to hypoxic, low pH waters. Decreased upwelling may result in lower primary production in these systems with direct impacts on pelagic fisheries productivity.